www.cambridge.org/qrd

Perspective

Cite this article: Wilson E, Kaushik A, Dutta S, Singharoy A (2025). The dawn of biophysical representations in computational immunology. *QRB Discovery*, **6**: e19, 1–7 https://doi.org/10.1017/qrd.2025.7.

Keywords:

Computational biophysics; Molecular; immunological, and structural biology; Bioinformatics; Vaccine development

Corresponding author: Abhishek Singharoy; Email: asinghar@asu.edu

A.K and S.D have contributed equally to this work.

The dawn of biophysical representations in computational immunology

Eric Wilson¹, Akshansh Kaushik², Soumya Dutta³ and Abhishek Singharoy³ D

¹Department of Immunology and Immunotherapy, Icahn School of Medicine at Mount Sinai, New York, NY, USA; ²School of Molecular Sciences, Arizona State University, Tempe, AZ, USA and ³Biodesign Institute, Center for Applied Structural Discovery

Abstract

Computational immunology has been the breeding ground of some of the best bioinformatics work of the day. By melding diverse data types, these approaches have been successful in associating genotypes with phenotypes. However, the representations (or spaces) in which these associations are mapped have primarily been constructed from some omics-oriented sequence data typically derived from high-throughput experiments. In this perspective, we highlight the importance of biophysical representations for performing the genotype–phenotype map. We contend that using biophysical representations reduces the dimensionality of a search problem, dramatically expedites the algorithm, and more importantly, offers physical interpretability to the classes of clustered sequences across different layers of complexity – molecular, cellular, or macro-level. Such biophysical interpretations offer a firm basis for the future of bioengineering and cell-based therapies.

Introduction

The core responsibility of our immune system is to protect the body from pathogens and cancers. The need to target and activate the immune system reproducibly has been underscored by the recent pandemic and the rise of anticancer therapies that rely on immunological mechanisms. This has generated a focused enthusiasm for gaining a detailed description of the immune system. However, the human immune system is incredibly complex, and often regarded as one of the most challenging topics in biology. The sheer size of sequence and population diversity in proteins associated with the immune system presents a formidable obstacle to mapping their network of interactions within a tractable space. For instance, T-cell recognition of antigens is driven by human leukocyte antigens (HLA) genes encoding Major Histocompatibility Complexes (or MHCs), which are among the most polymorphic germline genes in the human genome that contain tens of thousands of variants across populations (Barker et al., 2023). Moreover, the somatic hypermutations involved in the function of T-cell and B-cell receptors make them the most polymorphic human proteins in known existence, with theoretical estimates of T-cell receptor (or TCR) diversity reaching over 10⁶¹ potential sequences. A more conservative estimate places TCR diversity in the range of 10⁷ receptors (Mora and Walczak, 2018), which still offers an incredibly vast range of human variations. The desire to account for this diversity and predict its associated non-linear relationships has motivated the genesis of the field of computational immunology to develop methods to analyze and predict immune outcomes based on this data (Figure 1). Computational immunology has transformed our understanding of the immune system by enabling the integration of massive amounts of biochemical and biological data. Simple mathematical models to study disease transmission can be traced to the early 20th century (Ross, 1911; Brauer, 2017). By leveraging population data, it clarified the relationship between the size of mosquito populations and malaria incidence, which led to improved malaria control. The power of computational immunology expanded significantly in the information age with the advent of high-throughput sequencing, proteomics, and the growing availability of experimental and clinical data further empowered by advances in computational technology. These advancements have enabled computational techniques to tackle more complex immunological questions. Consequently, computational immunology has now been applied to a broad spectrum of applications including vaccine design (He and Zhu, 2015), predicting population-level mortality rates (Wilson et al., 2021), and forecasting the outcomes of immune checkpoint blockade therapies (Chowell et al., 2018).

Due to the availability and ease of collection of protein and amino acid sequence information, most computational immunology approaches primarily rely on sequence data for their predictions (Ansari and Raghava, 2010; Jespersen *et al.*, 2017; Peters *et al.*, 2020). However, recent advances in machine learning and protein modeling have caused an explosion in the synergistic incorporation of biophysical information and modeling into existing computational immunology approaches (Andersen *et al.*, 2006; Wilson *et al.*, 2024). Such integrations have already

© Arizona State University, 2025. Published by Cambridge University Press. This is an Open Access article, distributed under the terms of the Creative Commons Attribution licence (http://creativecommons.org/licenses/by/4.0), which permits unrestricted re-use, distribution and reproduction, provided the original article is properly cited.





Figure 1. The scales of computational immunology models from atomistic to macroscales.

shown a profound improvement in the accuracy of models, but also enable novel insights into previously inscrutable mechanisms, advancing computational immunology models into the next era. In the following perspective, we will explore immune-related models ranging from atomistic environments to macro-level systems, demonstrating how biophysics can be used to enhance predictive accuracy and improve our overall understanding of immune responses.

A perspective on biophysical models

Computational immunology has been dominated by bioinformatics, primarily due to a push from recent findings in genomic and proteomic technologies that compose around 31 different databases today (Rigden and Fernández, 2023). Historically, it allows the study of complex protein–protein interactions across a diversity of sequences (Petrovsky and Brusic, 2002). Recently, deep learning approaches have offered rapid access to molecular structures from sequences (Jumper *et al.*, 2021), which has extended the realm of bioinformatics to structure-guided models of immune interactions (Bradley, 2023). However, the physical formulation of intermolecular interactions is statistical, which entails an ensemble description of conformations that remains obscure in the bioinformatics approaches. These ensembles capture transition in the order-disorder transition of the molecules, flexibility, and thermal effects, as well as solvation and microenvironmental impacts on structure. Attempts to overcome such limitations of traditional computational immunology open the doors for employing biophysical tools to take MHC, TCRs, and antibody predictions beyond the sequence-only or sequence-structure paradigm (Raha et al., 2022; Deng et al., 2023; Demerdash and Smith, 2024). Notwithstanding the computationally expensive biophysical simulations, it generates unique representations and metrics that connect collective molecular properties with phenotypic and even population outcomes. We break down the biophysical advances in the realm of atomistic, molecular, wholecell, and macro-level modeling, and highlight how biophysical entities of Figure 1 are acting or can be leveraged as novel representations for learning in computational immunology, as

Illustrations	Immunological Problems	Biophysical represntation of sequences
	1. Antibody/Biologics design	1. Pocket fields, FEP+
	2. Structural modelling of MHC	2. Semi-emperical Quantum mechanical representations
	3. TRC predictions	3. Catch bond modelling
	4. Vaccine design	4. Contact matrices
CDB T CR	5. Whole cell modelling	5. Binding affinities
	6. Epitope predictions and disease association	6. Electrostatic and VDW representation

Figure 2. A comprehensive list of immunological problems and their biophysical representations. Illustrations – 1. Antibody (PDB-1IGT), 2. MHC (PDB-1HHK), 3. TCR (from RCSB-PDB), 4. Viral vector ChAdOx1, 5. Whole-cell illustration, and 6. Epitope (PDB-3PP4).

complements to the traditional sequence or structural methods (Figure 2).

Atomistic description

We start with biophysical descriptors in computational biology arising from detailed interactions of antibodies, MHCs, and TCRs.

Free energy description of antibodies

Since the first antibody structure was deposited in 1976, the number of antibody structures in the Protein Data Bank (PDB) has grown, and it now represents approximately 2.1% of the total entries (Ferdous and Martin, 2018). Many computational tools now use only the antibody data, as opposed to general protein data, due to the increased performance (Ponomarenko and Bourne, 2007; Młokosiewicz *et al.*, 2022). To this end, the Structural Antibody Database (or SAbDab) collects, curates, and presents an ensemble of antibody structures from the PDB (Schneider *et al.*, 2022). Such databases allow for the

prediction of the affinity of antibody-antigen interfaces by combining the biophysics of protein-protein interactions with deep learning approaches (Hummer et al., 2023). In fact, a significant improvement in the ranking and prediction of affinity predictions is observed by combining all-atom free energy methods like Free energy perturbation or FEP+ with focused machine learning approaches like QuanSA (Cleves and Jain, 2018). Using such a combination of biophysics and informatics, the affinity of the CR3022 antibody is optimized to the spike protein of the SARS-CoV-2 Omicron strain, achieving a high success rate with up to a 17-fold affinity increase (Cai et al., 2024). Going beyond simple geometric 3D coordinate representations of ligands (Cleves and Jain, 2018), a novel metric of multiple-ligand alignment is employed using so-called pocket fields to learn affinities. Unlike the learning of real geometries that are quite high-dimensional, the learning of smoother functions like the 3D fields (with known map to the SMILE or peptide sequences) offers learning across a broad diversity of molecular identity and conformation, without overfitting the loss function. In conclusion, the

application of free energy-augmented antibody design underscores the growing power of biophysical modeling to not only understand but also engineer biological systems for specific therapeutic outcomes.

Structural modeling of MHC (Major Histocompatibility Complex)

In 1968, Snell examined the concept of transplantation and came across the term histocompatibility polymorphism (Hull, 1970; Garrido, 2024). MHC proteins play a crucial role in immune mechanisms due to their involvement in activating T cells and B cells (Janeway et al., 2001; Wieczorek et al., 2017). Structural modeling of these complexes offers insights into the mechanism of the several pathways relevant to immunogenicity (Keller et al., 2022). The MHC protein is one of the most polymorphic proteins in humans (Barker et al., 2023), but despite the high polymorphism, the structure of the MHC binding groove is highly conserved (Wilson et al., 2024). Researchers found that the second and last residues are key anchors for peptide binding to the MHC class-I binding groove (Janeway et al., 2001), a discovery made through X-ray diffraction studies (Zhang et al., 1998). Since countless peptides can bind to MHC, many generated by frameshift events, and lack evolutionary context for multi-sequence alignments, crystallizing all polymorphic complexes is unfeasible. A biophysical approach is thus needed to model MHC-peptide complexes for further study.

Conventionally, there are three ways to model structures: molecular dynamics, molecular docking, and homology modeling (Bertoline et al., 2023). The unifying protocol to design a model for MHC is as follows: the first part is to generate a peptide conformation using a PDB template, the second step involves docking of the peptide, and finally optimizing the overall structure. Multiple sources are available to model MHC-I complexes such as Dock-Tope, GradDock, APE-Gen, AlphaFold2, and RoseTTAfold (Rigo et al., 2015; Kyeong et al., 2018; Abella et al., 2019; Bryant et al., 2022). Although these methods are highly accurate, some of them are highly computationally heavy or applicable only to the MHC class-I molecule due to the heterodimeric binding pocket observed in MHC class-II molecules. Recently, a state-of-the-art method, PANDORA, shows potential to design even MHC class-II molecules, and also offers some tunability while modeling. Its energybased definition of loop conformations is shown to outperform most of the methods previously introduced in terms of accuracy and computational efficiency (Parizi et al., 2023). However, there still is a need for a tool that models complex structures by capturing the biophysical attributes of the peptide-MHC complex instead of exploiting sequence similarity and templates. Large datasets to benchmark biophysical properties across a range of MHC systems - similar to MISATO (for MD simulations of 20,000 protein-ligand systems) or 100-protein NMR spectra (for protein dynamics) - do not yet exist in this space. A very promising result is that semiempirical quantum mechanical representations can now be embedded in these data sets to refine the associated protein structures. Once similar datasets start existing for the broad class of MHC proteins, such quantum chemistry representations can likely be extended to the peptide-MHC predictions, for example, with PANDORA or other tools. Ultimately, improvement to MHC modeling and subsequent extraction of generalizable biophysical properties will lead to better predictions of immunogenicity. Highlighting this point, a thorough structural study demonstrated that a non-anchor position mutation in an MHC-I peptide, presented by an ovarian cancer tumor, modified both the structural and dynamic properties of the bound complex. These changes resulted in optimal confirmations for interaction and subsequent

activation of cognate T cells (Devlin *et al.*, 2020). Such an observation would be difficult, if not impossible to determine from sequence alone and emphasizes the value of structural considerations when studying immunogenicity.

Catch bond description of TCRs

Catch bonds have been referred to as the interaction between various biomolecules and biomolecular surfaces, where the lifetime of the bond increases with the application of tensile force on the bond (Marshall et al., 2003; Hertig and Vogel, 2012). The atomistic detail of catch bond formation had remained elusive for a long period of time, but the general explanation was given by a two-state model or a two-pathway model. In the two-state model, the receptor-ligand complex is theorized to exist in two distinct states, a short-lived and a long-lived state. The application of force loosens the interaction between the binding site and a regulatory site, which drives the whole complex toward the long-lifetime state (Hertig and Vogel, 2012). In the two-pathway model, the receptor-ligand complex undergoes unbinding via two distinct pathways with different K_{off} values, and the application of tensile force triggers the allosteric change that leads the unbinding to happen via the pathway with a high energy barrier, thereby the long-lifetime (Sokurenko et al., 2008). Such catch bonding has been observed at the TCR-peptide-MHC immune synapse, and more importantly, immunogenicity has been attributed to the strength of the catch bond formation (Choi et al., 2023). Hence, catch bonds offer a biophysical descriptor of MHC alleles for presenting peptides to the TCRs. Interestingly, unlike binding affinity, catch bonds uniquely capture the system's out-of-equilibrium properties. Therefore, it can capture the state of the immune synapse under stress, which rectifies the frozen stationary picture of complexes drawn by the affinity measures. This descriptor is computable using Steered MD simulations (Schoeler et al., 2014) and more recently using metadynamics methodologies (Ccoa and Hocky, 2022), offering insights into how sequence changes reflect in non-equilibrium interaction changes. However, both the experimental and computational biophysical methods for tracking catch bonds are resource-intensive, so high-throughput measurements are yet missing, in turn impacting the extensive use of this information in immunology models. The advent of reinforcement learning with Jarzynski's equality and so-called stiff-spring approximations (Park and Schulten, 2004) to formulate a space of molecular actions using steered MD simulations presents a promising step forward in rapidly modeling at least the 2-state model of the catch bonds as another biophysical descriptor in computational immunology (Choi et al., 2023). A more rigorous consideration of catch bond formation has practical implications for enhancing T cell-based cancer immunotherapies. A recent study showed low-affinity TCRs can be optimized to acquire catch bonding characteristics, allowing for potent activation at relatively weak 3D binding affinities (Zhao et al., 2022). This has the ability to drive a strong antitumor immune response with a lower risk of potentially life-threatening cross-reactivity.

Molecular description

The translation from atomistic to molecular biophysical representation has become popular to allow algorithms to distinguish self versus non-self interactomes. The biophysical representations of glycans underpinning the pathogen entry path offer some stark examples. By employing tools like variational autoencoders, the so-called glycan shield of spike proteins was dissected to detect the role of specific glycan size, orientation, and chemistry (Casalino et al., 2021). A physical interpretation of the latent spaces was determined from protein-glycan contacts. Subsequently, we engineered the glycan shield based on their contact representation to reduce the infectivity of the NL63 coronavirus by nearly 50% (Chmielewski et al., 2023). This idea of monitoring contacts was also extrapolated to monitor inter-glycan interactions between the cell surface of the influenza virus and those of chicken and human cell surface glycocalyx (Lucas et al., 2021). Again, by translating fluorescence signals into a contact matrix representation, support vector machines were successful in identifying the critical density of glycans that make the H1N1 cells in mammalian cells show a greater binding than when grown in egg cells. Finally, the protein-protein contact matrices also found application in vector design for Astra-Zeneca and J&J's COVID vaccines, implicating platelet factor proteins in blood clotting side effects of the vaccine candidate (Baker et al., 2021). Altogether, contact matrices can offer a robust biophysical representation, wherein molecular interactions can be classified to be self or non-self.

Cellular description

Whole-cell models, though scarce, have found applications in computational immunology. A mechanistic, multiscale mathematical model of immunogenicity for therapeutic proteins was formulated by recapitulating key biological mechanisms, including antigen presentation, activation, proliferation, and differentiation of immune cells, secretion of antidrug antibodies, as well as in vivo disposition of antibodies and therapeutic proteins (Chen et al., 2014). The multiscale model structure can be represented by the subcellular, cellular, and whole-body levels. To represent the physiology of MHC-II, a key parameter used in these models involves the number of T-epitope-MHC, in silico T cell epitope prediction and experimental measurements of their MHC-binding affinities, which is scaffolded within a two-compartment drug pharmacokinetics model. Using adalimumab as an example therapeutic protein, the model is able to simulate immune responses against adalimumab in individual subjects and in a population and also provides estimations of immunogenicity incidence and drug exposure reduction that can be validated experimentally (Chen et al., 2014; Handel et al., 2020). Most of the cell models in immunology are agent-based that use the automaton algorithm with specific mechanistic logics or rules. Interestingly these rules show remarkable similarity with classical thermodynamic and kinetic principles, such as landscapes and equations of motion (Koopmans and Youk, 2021). Such models have found applications in CD4+ T cell responses to influenza infections, multiscale mechanistic modeling of human dendritic cells, and have potential applications in dendritic cell-based targeted cell therapies (Wertheim et al., 2021; Aghamiri et al., 2023).

Macro description

The integration of molecular immunology concepts into macrolevel analyses has already demonstrated significant potential in elucidating disease associations. A notable example is the use of patient-specific MHC genotypes to predict disease risk. For instance, large-scale analyses involving 9,176 cancer patients revealed that MHC-I genotypes were predictive of the tumor mutational landscape (Marty et al., 2017). This study found that oncogenic mutations were more likely to occur in regions not presented by the patient's MHC-I molecules, suggesting that gaps in antigen presentation contribute to tumor evolution. Similarly, patients

5

undergoing immune checkpoint blockade therapies have shown improved responses when their MHC-I genotype allows for the presentation of a more diverse array of potential peptides (Chowell et al., 2019). More recently, bio-physical approaches have been applied to link MHC-I genotypes with disease risk and progression (Wilson et al., 2024). Recently, we created a diverse protein ensemble of 5,281 MHC-I protein binding grooves, generating 211,240 structural models, which were subsequently translated into a simplified representation of electrostatic properties (5,281 averaged electrostatic maps). A subset of these maps, those with known MHC-I binding motifs, was used to train an Inception neural network capable of predicting MHC-I binding motifs from electrostatic maps alone. Beyond the ability to perform high-throughput proteome-scale binding predictions, the predicted binding motifs were utilized to construct interaction networks that accurately classified HIV disease progression and immune checkpoint therapy response. At the population level, applications of MHC-I genotype analysis have revealed further insights. A consensus MHC-I prediction model, ensem*bleMHC*, demonstrated that populations enriched for MHC-I alleles capable of strongly binding multiple peptides from SARS-CoV-2 structural proteins exhibited lower mortality rates during the prevaccination phase of the COVID-19 pandemic (Wilson et al., 2021). This suggests that MHC-I diversity and peptide-binding capacity at the population level may serve as predictors of disease outcomes in emerging viral threats. These findings highlight some of the promise of MHC genotype-based analysis in both disease risk assessment and therapeutic strategy development. MHC analysis can aid in predicting susceptibility to autoimmune diseases and cancer while also informing vaccine design by optimizing patient antigen selection.

Outlook: Future inspired by the past of functional representations

Most of the biophysics, including the powerful integrative models we know, is predicated upon the sequence \rightarrow structure \rightarrow function \rightarrow phenotype paradigm. With the maturation of machine learning techniques and the availability of data at various scales, researchers (particularly bioinformaticians) have been trying to bridge gaps between the different tiers of this process, starting from the age-old genotype-type modeling to CASP and AlphaFold's sequence structure up to recent attempts to go from sequence to ensemble. However, physical causality is often missing in the traditional bioinformatics models, thus far sidelining the role of AI-driven advances only to predictions of the forward direction. So, it is high time that we introduce physical ideas to conceive generative models that backmap phenotypes down to an ensemble of structures and sequences. Model representations play a central role in this mapping process. Although the traditional sequence of 3D coordinate structural representations requires an enormous amount of training data and is prone to overfitting, they nonetheless offer the most extensive models. In contrast, the thermodynamic or kinetic representations, using ideas of entropy or committor functions are quite generalizable across application domains but lack the physical interpretability (Mehdi et al., 2024). Loosely, they draw analogies to the plane wave basis set representations that find application in several areas of quantum mechanics (Nagy and Jensen, 2017). However, akin to how quantum mechanics was represented in the molecular systems using the Gaussian-like basis set representations, we posit that biophysical representations offer a segue for representing the deep learning models in the molecular space. To this end, we highlight a number of representations that are either

being used or hold the potential for multiscale applications in computational immunology. Similar to how Gaussian orbitals offer physical interpretation of highly resolved electronic structures (e.g. using the molecular orbital theory), biophysical functions offer interpretability. These functions, such as pocket fields, QM/MM charge density, binding affinity, catch bonding, contact matrices, and molecular electrostatics are deeply rooted in physical theories. These theories (thermodynamic integration, electronic structure theory, equilibrium and non-equilibrium statistical theories, linear response theories, polymer folding, and continuum mechanics) can be projected onto structure and function. Essentially, they offer a physical basis to the loss functions and the latent spaces that enable learning both the data and the context. So, we propose a sustained intellectual effort in this direction.

Open peer review. To view the open peer review materials for this article, please visit http://doi.org/10.1017/qrd.2025.7.

Acknowledgments. A.S. also acknowledges start-up grants from Arizona State University School of Molecular Sciences and Biodesign Institute's Center for Applied Structural Discovery. A.S. acknowledges funding from the Division of Chemical Sciences, Geosciences, and Biosciences, Office of Basic Energy Sciences, of the U.S. Department of Energy through grants DESC0010575. A.S. acknowledges grant DE-SC0022956 for their support, also from the Department of Energy. This material is based on work supported by the National Defense Education Program (NDEP) for Science, Technology, Engineering, and Mathematics (STEM) Education.

Financial support. A.S. was supported by a CAREER award from the NSF (MCB-1942763) and an RO1 grant from the NIH (GM095583).

Competing interest. The authors declare no competing interests exist.

References

- Abella JR, Antunes DA, Clementi C and Kavraki LE (2019) APE-gen: A fast method for generating ensembles of bound peptide-MHC conformations. *Molecules* 24(5), 881.
- Aghamiri SS, Puniya BL, Amin R and Helikar T (2023) A multiscale mechanistic model of human dendritic cells for in-silico investigation of immune responses and novel therapeutics discovery. *Frontiers in Immunology* 14, 1112985.
- Andersen PH, Nielsen M and Lund OLE (2006) Prediction of residues in discontinuous B-cell epitopes using protein 3D structures. *Protein Science* 15(11), 2558–2567.
- Ansari HR and Raghava GPS (2010) Identification of conformational B-cell epitopes in an antigen from its primary sequence. *Immunome Research* 6, 1–9.
- Baker AT, Boyd RJ, Sarkar D, Teijeira-Crespo A, Chan CK, Bates E, Waraich K, Vant J, Wilson E, Truong CD, Lipka-Lloyd M, Fromme P, Vermaas J, Williams D, Machiesky L, Heurich M, Nagalo BM, Coughlan L, Umlauf S, Chiu PL, Rizkallah PJ, Cohen TS, Parker AL, Singharoy A and Borad MJ (2021) ChAdOx1 interacts with CAR and PF4 with implications for thrombosis with thrombocytopenia syndrome. *Science Advances* 7(49), eabl8213. https://doi.org/10.1126/sciadv.abl8213.
- Barker DJ, Maccari G, Georgiou X, Cooper MA, Flicek P, Robinson J and Marsh SGE (2023) The ipd-imgt/hla database. Nucleic Acids Research 51(D1), D1053–D1060.
- Bertoline LMF, Lima AN, Krieger JE and Teixeira SK (2023) Before and after AlphaFold2: An overview of protein structure prediction. *Frontiers in Bioinformatics* 3, 1120370.
- Bradley P (2023) Structure-based prediction of T cell receptor: Peptide-MHC interactions. *eLife* 12, e82813.
- Brauer F (2017) Mathematical epidemiology: Past, present, and future. Infectious Disease Modelling 2(2), 113–127.
- Bryant P, Pozzati G and Elofsson A (2022) Improved prediction of proteinprotein interactions using AlphaFold2. Nature Communications 13(1), 1265.

- Cai H, Zhang Z, Wang M, Zhong B, Li Q, Zhong Y, Wu Y, Ying T and Tang J (2024) Pretrainable geometric graph neural network for antibody affinity maturation. *Nature Communications* 15(1), 7785.
- Casalino L, Dommer AC, Gaieb Z, Barros EP, Sztain T, Ahn SH, Trifan A, Brace A, Bogetti AT, Clyde A, Ma H, Lee H, Turilli M, Khalid S, Chong LT, Simmerling C, Hardy DJ, Maia JD, Phillips JC, Kurth T, Stern AC, Huang L, McCalpin JD, Tatineni M, Gibbs T, Stone JE, Jha S, Ramanathan A and Amaro RE (2021) AI-driven multiscale simulations illuminate mechanisms of SARS-CoV-2 spike dynamics. *The International Journal of High Performance Computing Applications* 35(5), 432–451. https://doi.org/10.1177/ 10943420211006452.
- Ccoa WJP and Hocky GM (2022) Assessing models of force-dependent unbinding rates via infrequent metadynamics. *The Journal of Chemical Physics* 156(12), 125102. https://doi.org/10.1063/5.0081078.
- Chen X, Hickling T and Vicini P (2014) A mechanistic, multiscale mathematical model of immunogenicity for therapeutic proteins: Part 2—Model applications. CPT: Pharmacometrics & Systems Pharmacology 3(9), 1–10.
- Chmielewski D, Wilson EA, Pintilie G, Zhao P, Chen M, Schmid MF, Simmons G, Wells L, Jin J, Singharoy A and Chiu W (2023) Structural insights into the modulation of coronavirus spike tilting and infectivity by hinge glycans. *Nature Communications* 14(1), 7175. https://doi.org/10.1038/ s41467-023-42836-9.
- Choi H-K, Cong P, Ge C, Natarajan A, Liu B, Zhang Y, Li K, Rushdi MN, Chen W, Lou J, Krogsgaard M and Zhu C (2023) Catch bond models may explain how force amplifies TCR signaling and antigen discrimination. *Nature Communications* 14(1), 2616.
- Chowell D, Krishna C, Pierini F, Makarov V, Rizvi NA, Kuo F, Morris LGT, Riaz N, Lenz TL and Chan TA (2019) Evolutionary divergence of HLA class I genotype impacts efficacy of cancer immunotherapy. *Nature Medicine* 25(11), 1715–1720.
- Chowell D, Morris LGT, Grigg CM, Weber JK, Samstein RM, Makarov V, Kuo F, Kendall SM, Requena D, Riaz N, Greenbaum B, Carroll J, Garon E, Hyman DM, Zehir A, Solit D, Berger M, Zhou R, Rizvi NA and Chan TA (2018) Patient HLA class I genotype influences cancer response to checkpoint blockade immunotherapy. *Science* 359(6375), 582–587. https://doi.org/ 10.1126/science.aao4572.
- Cleves AE and Jain AN (2018) Quantitative surface field analysis: Learning causal models to predict ligand binding affinity and pose. *Journal of Computer-Aided Molecular Design* 32, 731–757.
- Demerdash ONA and Smith JC (2024) TCR-H: Explainable machine learning prediction of T-cell receptor epitope binding on unseen datasets. *Frontiers in Immunology* 15, 1426173.
- Deng L, Ly C, Abdollahi S, Zhao Y, Prinz I and Bonn S (2023) Performance comparison of TCR-pMHC prediction tools reveals a strong data dependency. *Frontiers in Immunology* 14, 1128326.
- Devlin JR, Alonso JA, Ayres CM, Keller GL, Bobisse S, Kooi CWV, Coukos G, Gfeller D, Harari A and Baker BM (2020) Structural dissimilarity from self drives neoepitope escape from immune tolerance. *Nature Chemical Biology* 16(11), 1269–1276.
- Ferdous S and Martin ACR (2018) AbDb: Antibody structure database—A database of PDB-derived antibody structures. *Database* 2018, bay040.
- Garrido F (2024) The discovery of the major histocompatibility complex (MHC): The H-2 in mice and the HLA in man. In *The Major Histocompatibility Complex (MHC/HLA) in Medicine: A Personal Recollection*. Springer, Cham, pp. 1–13. https://doi.org/10.1007/978-3-031-59866-1_1.
- Handel A, La Gruta NL and Thomas PG (2020) Simulation modelling for immunologists. *Nature Reviews Immunology* 20(3), 186–195.
- He L and Zhu J (2015) Computational tools for epitope vaccine design and evaluation. *Current Opinion in Virology* 11, 103–112.
- Hertig S and Vogel V (2012) Catch bonds. *Current Biology* 22, R823–R825, 10. Hull P (1970) Notes on dr snell's observations concerning the h-2 locus
- polymorphism. Hummer AM, Schneider C, Chinery L and Deane CM (2023) Investigating the
- volume and diversity of data needed for generalizable antibody-antigen G prediction. *bioRxiv*, 2023–2025.
- Janeway CA Jr, Travers P, Walport M and Shlomchik MJ (2001) The major histocompatibility complex and its functions. In *Immunobiology: The Immune*

System in Health and Disease, 5th edn. New York: Garland Science. Available from: https://www.ncbi.nlm.nih.gov/books/NBK27156/

- Jespersen MC, Peters B, Nielsen M and Marcatili P (2017) BepiPred-2.0: Improving sequence-based B-cell epitope prediction using conformational epitopes. *Nucleic Acids Research* **45**(W1), W24–W29.
- Jumper J, Evans R, Pritzel A, Green T, Figurnov M, Ronneberger O, Tunyasuvunakool K, Bates R, Žídek A, Potapenko A, Bridgland A, Meyer C, Kohl SAA, Ballard AJ, Cowie A, Romera-Paredes B, Nikolov S, Jain R, Adler J, Back T, Petersen S, Reiman D, Clancy E, Zielinski M, Steinegger M, Pacholska M, Berghammer T, Bodenstein S, Silver D, Vinyals O, Senior AW, Kavukcuoglu K, Kohli P and Hassabis D (2021) Highly accurate protein structure prediction with AlphaFold. Nature 596(7873), 583–589. https://doi.org/10.1038/s41586-021-03819-2.
- Keller GLJ, Weiss LI and Baker BM (2022) Physicochemical heuristics for identifying high fidelity, near-native structural models of peptide/MHC complexes. Frontiers in Immunology 13, 887759.
- Koopmans L and Youk H (2021) Predictive landscapes hidden beneath biological cellular automata. *Journal of Biological Physics* 47(4), 355–369.
- Kyeong H-H, Choi Y and Kim H-S (2018) GradDock: Rapid simulation and tailored ranking functions for peptide-MHC class I docking. *Bioinformatics* 34(3), 469–476.
- Lucas TM, Gupta C, Altman MO, Sanchez E, Naticchia MR, Gagneux P, Singharoy A and Godula K (2021) Mucin-mimetic glycan arrays integrating machine learning for analyzing receptor pattern recognition by influenza a viruses. *Chem* 7(12), 3393–3411.
- Marshall BT, Long M, Piper JW, Yago T, McEver RP and Zhu C (2003) Direct observation of catch bonds involving cell-adhesion molecules. *Nature* 423, 190–193, 5.
- Marty R, Kaabinejadian S, Rossell D, Slifker MJ, van de Haar J, Engin HB, de Prisco N, Ideker T, Hildebrand WH, Font-Burgada J and Carter H (2017) MHC-I genotype restricts the oncogenic mutational landscape. *Cell* 171(6), 1272–1283. https://doi.org/10.1016/j.cell.2017.09.050.
- Mehdi S, Smith Z, Herron L, Zou Z and Tiwary P (2024) Enhanced sampling with machine learning. *Annual Review of Physical Chemistry* **75**, 347–370.
- Młokosiewicz J, Deszyński P, Wilman W, Jaszczyszyn I, Ganesan R, Kovaltsuk A, Leem J, Galson JD and Krawczyk K (2022) AbDiver: A tool to explore the natural antibody landscape to aid therapeutic design. *Bioinformatics* 38(9), 2628–2630.
- Mora T and Walczak AM (2018) Quantifying lymphocyte receptor diversity. In Systems Immunology: An Introduction to Modeling Methods for Scientists. Das J, ed. CRC Press, Taylor and Francis, pp. 183–198.
- Nagy B and Jensen F (2017) Basis sets in quantum chemistry. Reviews in Computational Chemistry 30, 93–149.
- Parizi FM, Marzella DF, Ramakrishnan G, t Hoen PAC, Karimi-Jafari MH and Xue LC (2023) PANDORA v2. 0: Benchmarking peptide-MHC II models and software improvements. *Frontiers in Immunology* 14, 1285899.
- Park S and Schulten K (2004) Calculating potentials of mean force from steered molecular dynamics simulations. *The Journal of Chemical Physics* 120(13), 5946–5961.

- Peters B, Nielsen M and Sette A (2020) T cell epitope predictions. Annual Review of Immunology 38(1), 123–145.
- Petrovsky N and Brusic V (2002) Computational immunology: The coming of age. Immunology and Cell Biology 80(3), 248–254.
- Ponomarenko JV and Bourne PE (2007) Antibody-protein interactions: Benchmark datasets and prediction tools evaluation. BMC Structural Biology 7, 1–19.
- Raha R, Ding Y, Liu Q and Wu F-X (2022) Unseen epitope-TCR interaction prediction based on amino acid physicochemical properties. In 2022 IEEE International Conference on Bioinformatics and Biomedicine (BIBM). Las Vegas, NV: IEEE, pp. 3122–3129, doi: 10.1109/BIBM55620.2022.9995066.
- Rigden DJ and Fernández XM (2023) The 2024 nucleic acids research database issue and the online molecular biology database collection. *Nucleic Acids Research* 52, D1–D9, 10.
- Rigo MM, Antunes DA, de Freitas M, de Almeida Mendes M, Meira L, gaglia MS and Vieira GF (2015) DockTope: A web-based tool for automated pMHC-I modelling. *Scientific Reports* 5(1), 18413.
- Ross R (1911) The Prevention of Malaria. E.P. Dutton and company, New York.
- Schneider C, Raybould MIJ and Deane CM (2022) SAbDab in the age of biotherapeutics: Updates including SAbDab-nano, the nanobody structure tracker. Nucleic Acids Research 50(D1), D1368–D1372.
- Schoeler C, Malinowska KH, Bernardi RC, Milles LF, Jobst MA, Durner E, Ott W, Fried DB, Bayer EA, Schulten K and Gaub HE (2014) Ultrastable cellulosome-adhesion complex tightens under load. *Nature Communications* 5(1), 5635.
- Sokurenko EV, Vogel V and Thomas WE (2008) Catch bond mechanism of force-enhanced adhesion: Counter-intuitive, elusive but ... Widespread?. *Cell Host & Microbe* **4**, 314, 10.
- Wertheim KY, Puniya BL, La Fleur A, Shah AR, Barberis M and Helikar T (2021) A multi-approach and multi-scale platform to model cd4+ t cells responding to infections. *PLoS Computational Biology* **17**(8), e1009209.
- Wieczorek M, Abualrous ET, Sticht J, Álvaro-Benito M, Stolzenberg S, Noé F and Freund C (2017) Major histocompatibility complex (mhc) class i and mhc class ii proteins: Conformational plasticity in antigen presentation. *Frontiers in Immunology* 8, 292.
- Wilson E, Cava JK, Chowell D, Raja R, Mangalaparthi KK, Pandey A, Curtis M, Anderson KS and Singharoy A (2024) The electrostatic landscape of MHC-peptide binding revealed using inception networks. *Cell Systems* 15(4), 362–373.
- Wilson EA, Hirneise G, Singharoy A and Anderson KS (2021) Total predicted MHC-I epitope load is inversely associated with population mortality from SARS-CoV-2. Cell Reports Medicine 2(3).
- Zhang C, Anderson A and DeLisi C (1998) Structural principles that govern the peptide-binding motifs of class I MHC molecules. *Journal of Molecular Biology* 281(5), 929–947.
- Zhao X, Kolawole EM, Chan W, Feng Y, Yang X, Gee MH, Jude KM, Sibener LV, Fordyce PM, Germain RN, Evavold BD and Garcia KC (2022) Tuning t cell receptor sensitivity through catch bond engineering. *Science* 376(6589), eabl5282. https://doi.org/10.1126/science.abl5282.