Probability risk transmission matrix as a decision tool for assessing methods of transmission interruption of *Wuchereria bancrofti* infection

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SUMMARY

Lymphatic filariasis continues to cause severe morbidity and economic loss. The World Health Assembly (WHA) has passed a resolution to eliminate this disease by 2020. The major thrust of the elimination strategy is interrupting transmission by anti-parasitic treatment of entire communities. However, both vector density and community microfilaria load (CMFL) influence the intensity of transmission. Therefore, using a logistic regression approach a relationship has been established between the Risk of Infection Index (RII), vector density and CMFL. The present analysis indicates that there is no risk of transmission as long as the CMFL is maintained below 5 microfilaria (mf)/60 mm³ and the vector density per man-hour (MHD) is <25. However, transmission may continue when vector density is >25 and CMFL is <5 mf/60 mm³. In situations where CMFL is very high, parasitic control by mass administration may be cost effective in interrupting transmission. But at lower level of CMFL (<4 mf) and higher level of vector density it might be more cost effective to use vector control methods. A RII value <0.2 is considered to be the threshold for confirming interruption of transmission. Thus, the relationship has been depicted in the form of a probability matrix, which could be used for selecting an appropriate control strategy.

INTRODUCTION

Lymphatic filariasis continues to be a major source of chronic morbidity in tropical countries and an estimated 120 million people are infected either with *Wuchereria* or *Brugia* [1, 2]. In India economic loss due to filariasis was estimated to be around US\$ 842 million per year [3]. Considering the gravity of the problem, the World Health Assembly (WHA) passed a resolution in 1997 to eliminate this disease as a public health problem by 2020. The elimination strategy includes transmission control and morbidity management. Transmission could in principle be interrupted either by parasite control and/or vector control; present strategy exclusively relies on antiparasitic measures [4-6]. Past experience suggests that depending upon any one method is not wise; by either method initial success is very dramatic but subsequent progress in reducing either parasite load or vector density becomes increasingly costly. The Risk of Infection Index (RII), which indicates the intensity of transmission [7, 8] from indoor resting mosquitoes, is a useful index to measure the infection status in mosquitoes as well as humans. The RII is influenced by vector density and community microfilaria load (CMFL). In this paper we derived a probability matrix, which can be used to decide which control

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method would be more cost effective at the current level of endemicity.

METHODS

A study has been under way since 1993 by the Vector Control Research Centre, Pondicherry to evaluate the impact of mass chemotherapy with ivermectin and/ or diethylcarbamazine (DEC) on transmission and microfilaraemia in 20 rural sites of Villupuram District in Tamil Nadu state, India. The impact of chemotherapy on transmission parameters was evaluated by indoor resting collections of mosquitoes at monthly intervals in all the sites. The resting mosquitoes were collected from each site in 12 randomly selected huts by four well-trained insect collectors, spending 15 min per hut and a total of 3 man-hours per site. The total number of mosquitoes collected at each site was divided by the number of man-hours spent to obtain the vector density of the site in a particular month. The collected mosquitoes were brought to the laboratory and the mosquitoes of the vector species, Culex quinquefasciatus, the only incriminated natural vector of Wuchereria bancrofti in India were dissected to determine the vector infection and infectivity rates and parasite load. The number of developing filarial larvae of W. bancrofti were classified into microfilariae (mf), 'sausage stage' (L1), stage II (L2) and stage III or infective stage (L3) following the criteria proposed by Samarawickrema [9]. To assess the impact of chemotherapy on human infection, parasitological surveys were undertaken in all the sites immediately prior to the next round of chemotherapy. To determine sample size for assessing microfilaria prevalence in a site, a population of minimum 5000 was assumed. By allowing 20% error on the expected mf prevalence with 95% confidence the required sample size was estimated to be a minimum 7% of the population or 7% of households in the site. This 7% of households were selected randomly and all household members had blood sampled. The same proportionate sample was adopted in all the study sites. The details of the study design and results have been presented previously [10].

In addition data collected from 17 urban sites in the Pondicherry endemic area early in control operations in 1981 [11–14] were also used for calculating RII values and further statistical analyses. For validation and to estimate the level at which transmission does not occur, data were used from the 17 urban sites where resting mosquitoes were collected later in a control operation in 1986 [8, 11–14]. The RII for each site and month was calculated by a previous method [8] as follows:

$\mathbf{RII} = a \times b \times c,$

where, a = parous mosquitoes collected per manhour; b = average number of L3 larvae/parous mosquito; c = proportion of L3 larvae penetrating the host from one bite (i.e. 0.414×0.32) [15–17].

Based on the parasitological survey, mf prevalence at each site (percentage of individuals with filarial infection) was calculated and the CMFL was calculated as follows:

CMFL = prevalence of mf

 $\times \left(\begin{array}{c} \text{geometric mean count of} \\ \text{microfilaraemia per individual} \end{array} \right).$

Statistical analysis

Multiple linear regression analysis

Considering the RII values at each site as dependent values and the resting vectors collected per man-hour (MHD) and CMFL as explanatory variables, multiple regression analysis was carried out.

Logistic regression analysis conceptual framework

Considering RII as an indicator of transmission, logistic regression is an appropriate model to estimate the probability of transmission. The form of the logistic model is

$$\Pr(y=1|x) = \frac{\exp(\alpha + \Sigma\beta_i x_i)}{1 + \exp(\alpha + \Sigma\beta_i x_i)},$$

where y=1 or y=0 indicates whether a community is at risk of transmission in relation to regression variables (x_i) such as vector resting density and CMFL. The constant term of the logistic regression equation is indicated by α and the logistic regression coefficients are denoted by β_i from which odds ratios are calculated.

RESULTS

Initially multiple regression analysis was carried out using the RII as the dependent variable and CMFL and MHD as independent variables. It was observed that only 21% of variation in RII was explained by the CMFL and MHD. Further, the coefficient of

Variable	Coefficient value	S.E.	P value	Odds ratio	95% CI
CMFL	0.0488	0.0245	0.0467	1.05	1.00-1.10
MHD	0.0550	0.0057	0.0000	1.06	1.04 - 1.07
Constant	-1.703	0.1786	0.0000		

Table 1. Parameter estimation of logistic regression equation bymaximum-likelihood estimation method

CMFL, Community microfilaria load; MHD, vector density per man-hour. Null model $\chi^2 = 788.44$; model $\chi^2 = 132.50$.

CMFL was not statistically significant indicating that the multiple regression equation did not adequately fit the data. Therefore, a logistic regression equation was fitted to the data.

In the logistic regression analysis the RII was forced to be a binary response variable. Therefore, all the RII values greater than zero were recoded as '1' and all zero values of RII were left as '0'. The vector density (MHD) and CMFL were considered as covariates. For each variable the estimated slope coefficients, standard error (s.E.), odds ratio, log-likelihood for the model and the likelihood ratio test statistics (deviance) for the hypothesis that the slope coefficient is zero are presented in Table 1. The deviance is obtained as minus twice the difference between the loglikelihood for the constant-only model and the model containing the respective variables. On the null hypothesis that the deviance will follow the χ^2 distribution at 1 D.F., the significance of each variable was tested. The deviance of intercept-only model was 788.4 and the significance of the model was assessed by the likelihood-ratio test statistics (deviance) between the model of intercept-only and the model with independent variables. The model χ^2 (132.5) with 2 D.F. was highly significant (P < 0.001), i.e. one can reject the null hypothesis that the coefficients for all the terms in the model were zero. The significance of individual coefficients was also tested based on the Wald statistic (the square of the ratio of the coefficient to its standard error), which has a χ^2 distribution. The result showed that while the variable MHD was highly significant (P < 0.001), CMFL was of only borderline significance (P = 0.0467) in the model. The probability of RII for a given value of CMFL and MHD is calculated as follows

$\operatorname{prob} = I/(I + \exp(-z)),$

where $z = -1.703 + 0.049 \times \text{CMFL} + 0.055 \times \text{MHD}$. For different values of CMFL and MHD the probability matrix of RII was calculated and they are presented in Table 2.

In order to estimate the probability risk level at which transmission ceases to occur, risk was estimated for the 17 urban sites in the Pondicherry endemic area where resting mosquitoes were collected in the later stage of a control operation [8, 11-14]. For all of these 17 sites the incidence rate (%) of filarial infection among the children (0-5 years) born during the control period was calculated. The risk and the corresponding incidence rate are depicted in the Figure. It was observed that while in four out of six sites where the filarial incidence was recorded the risk was ≥ 0.5 , but in two sites it was around 0.2. Simple linear regression analysis was carried out using the incidence rate as the dependent variable and risk as the independent variable. The fitted regression equation (incidence = $-0.76 + 4.10 \times risk$) was statistically significant (F = 7.88, P = 0.013) indicating that linear regression adequately fitted the data. From the regression equation the predicted risk was 0.2 when the incidence rate was taken as zero. Therefore, a minimum probability of risk of 0.2 is considered to be safe value for interruption of transmission.

DISCUSSION

The present analysis was carried out to find the relationship between the risk of infection, CMFL and resting vector density. Using the established logistic regression equation a probability matrix was constructed for different levels of CMFL and vector density. The negative intercept in the equation indicates that when the values of CMFL and MHD approach zero the risk of transmission is almost nil. Moreover, for both the variables CMFL and MHD the lower limit of odds ratio is 1. Since incidence of

51	0.811	0.819	0.826	0.833	0.840	0.846	0.852	0.859	0.864	0.870	0.875	0.881	0.886	0.891	0.895
48	0.780	0.789	0.797	0.804	0.812	0.819	0.827	0.834	0.840	0.847	0.853	0.859	0.865	0.871	0.87
45	0.746	0.755	0.764	0.772	0.781	0.789	0.797	0.805	0.813	0.820	0.827	0.834	0.841	0.847	0.853
42	0.707	0.717	0.727	0.737	0.746	0.755	0.764	0.773	0.782	0.790	0.798	0.806	0.813	0.821	0.828
39	0.666	0.677	0.687	0.698	0.708	0.718	0.728	0.738	0.747	0.756	0.765	0.774	0.782	0.791	0.799
36	0.622	0.633	0.645	0.656	0.667	0.678	0.688	0.699	0.709	0.719	0.729	0.738	0.748	0.757	0.766
33	0.576	0.588	0.600	0.611	0.623	0.634	0.646	0.657	0.668	0.679	0.689	0.700	0.710	0.720	0.730
30	0.528	0.541	0.553	0.565	0.577	0.589	0.601	0.612	0.624	0.635	0.647	0.658	0.669	0.680	0.690
27	0.480	0.493	0.505	0.517	0.529	0.542	0.554	0.566	0.578	0.590	0.602	0.613	0.625	0.636	0.648
24	0.433	0.445	0.457	0.469	0.481	0.494	0.506	0.518	0.530	0.543	0.555	0.567	0.579	0.591	0.603
21	0.386	0.398	0.410	0.422	0.434	0.446	0.458	0.470	0.482	0.495	0.507	0.519	0.531	0.544	0.556
18	0.342	0.353	0.364	0.376	0.387	0.399	0.411	0.423	0.435	0.447	0.459	0.471	0.483	0.496	0.508
15	0.300	0.310	0.321	0.332	0.343	0.354	0.365	0.376	0.388	0.400	0.412	0.424	0.436	0.448	0.460
12	0.261	0.271	0.280	0.290	0.301	0.311	0.322	0.333	0.344	0.355	0.366	0.377	0.389	0.401	0.413
9	0.226	0.234	0.243	0.252	0.262	0.271	0.281	0.291	0.302	0.312	0.323	0.333	0.344	0.356	0.367
6	0.194	0.202	0.210	0.218	0.226	0.235	0.244	0.253	0.263	0.272	0.282	0.292	0.302	0.313	0.324
3	0.166	0.172	0.180	0.187	0.195	0.202	0.210	0.219	0.227	0.236	0.245	0.254	0.263	0.273	0.283
	1.0	2.0	3.0	4.0	5.0	6.0	7.0	8.0	9.0	10.0	11.0	12.0	13.0	14.0	15.0

 Table 2. Implication of probability matrix of risk of infection index on control strategy

Community microfilaria load (mf/60 mm³)

(Probability of ≤ 0.5 considered to be indicator of no transmission).

Key to cells: Vector control (bold face italic); combination of vector and parasite control (light grey cells); monitoring and surveillance until infection dies out (dark grey cells); parasite control (white cells).



Fig. Incidence of filarial infection in the 0–5 years age group by risk probability. #, Observed; —, predicted. Incidence = $-0.76 + 4.1 \times \text{risk}$, $R^2 = 0.59$, P = 0.013.

filarial infection was recorded generally in sites where the probability of risk is >0.5, it is presumed that the probability of ≥ 0.5 is a positive indicator of transmission. Therefore, it can be inferred from Table 2 that there is no risk of transmission as long as the CMFL is maintained below 5 mf/60 mm³ and MHD is <25. While the MHD is <15 there is no risk of transmission even for higher CMFL (15 mf/60 mm³). This implies that in an endemic area, parasite control should be initiated to bring down the CMFL to ≤ 2 and maintain the resting density at <30 per manhour by introducing vector control in order to facilitate interruption of infection. It also indicates that in the initial stages where CMFL is very high antiparasitic tools may be more cost effective than vector control, the impact of which is only seen after many years of control operations. An earlier study [18] indicated that inclusion of a vector control programme along with chemotherapy was not cost effective, i.e. as we become closer to elimination, the cost effectiveness will decline and we may reach a point where any additional input will not produce the desired results. However, earlier studies [18, 19] have shown that after mass drug administration there was sustained reduction in microfilaraemia when vector control measures alone were used in subsequent years. Therefore, in areas with low level of CMFL and higher vector density it might be more economical to bring down vector density by vector control measures rather than mass drug administration. The probability risk matrix can serve as a decision-making tool for judging the most cost-effective control programme to achieve interruption of lymphatic filariasis transmission.

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DECLARATION OF INTEREST

None.

REFERENCES

- Michael E, Bundy DAP, Grenfell BT. Re-assessing the global prevalence and distribution of lymphatic filariasis. *Parasitology* 1996; 112: 409–428.
- Michael E, Bundy DAP. Global mapping of lymphatic filariasis. *Parasitology Today* 1997; 13: 472–476.
- Ramaiah KD, et al. The economic burden of lymphatic filariasis in India. Parasitology Today 2000; 16: 251– 253.
- Ottesen EA, Campbell WC. Ivermectin in human medicine. *Journal of Antimicrobial Chemotherapy* 1994; 34: 195–203.
- Ottesen EA, et al. Strategies and tools for the control/ elimination of lymphatic filariasis. Bulletin of the World Health Organization 1997; 75: 491–503.
- Ottesen EA, Ismail MM, Horton J. The role of albendazole in programmes to eliminate lymphatic filariasis. *Parasitology Today* 1999; 15: 382–386.
- Das PK, et al. Cumulative exposure and its relationship with chronic filarial disease in bancroftian filariasis. Southeast Asian Journal of Tropical Medicine and Public Health 1994; 25: 516–521.
- Vanamail P, Ramaiah KD, Das PK. Risk of infection of *Wuchereria bancrofti* to humans by *Culex quinquefasciatus* in Pondicherry and its relationship with microfilaria prevalence *Acta Tropica* 1993; 55: 231–237.
- 9. Samarawickrema WA. A study of the age composition of natural population of *Culex pipiens fatigans* Wiedemann in relation to the transmission of filariasis

due to *Wuchereria bancrofti* (Cobbold) in Cylon. *Bulletin of the World Health Organization* 1967; **37**: 117–137.

- Das PK, et al. Placebo-controlled community trial of four cycles of single dose diethylcarbamazine or ivermectin against Wuchereria bancrofti infection and transmission in India. Transactions of the Royal Society of Tropical Medicine and Hygiene 2001; 95: 336–341.
- Rajagopalan PK, et al. Evaluation of integrated vector control measures on filariasis transmission in Pondicherry. *Indian Journal of Medical Research* 1988; 87: 434–439.
- Rajagopalan PK, et al. Bancroftian filariasis in Pondicherry, south India: 1. Pre-control epidemiological observations. *Epidemiology and Infection* 1989; 103: 685–692.
- Subramanian S, et al. Bancroftian filariasis in Pondicherry, south India: 2. Epidemiological evaluation of the effect of vector control. *Epidemiology and Infection* 1989; 103: 693–702.
- Das PK, et al. Bancroftian filariasis in Pondicherry, south India – epidemiological impact of recovery of the vector population. *Epidemiology and Infection* 1992; 108: 483–493.
- De Meillon B, Grab B, Sebastian A. Evaluation of Wuchereria bancrofti infection in Culex pipiensfatigans in Rangoon, Burma. Bulletin of the World Health Organization 1967; 36: 91–100.
- Ewert A, Ho BC. The fate of Brugia pahangi larvae immediately after feeding by infective vector mosquito. *Transactions of the Royal Society of Tropical Medicine* and Hygiene 1967; 61: 659–662.
- Ho BC, Ewert A. Experimental transmission of filarial larvae in relation to feeding behaviour of the mosquito vectors. *Transactions of the Royal Society of Tropical Medicine and Hygiene* 1967; 61: 663–666.
- Reuben R, et al. Annual single-dose diethylcarbamazine plus ivermectin for control of bancroftian filariasis: comparative efficacy with and without vector control. *Annals of Tropical Medicine and Parasitology* 2001; 95: 361–378.
- 19. Maxwell CA, et al. Can vector control play a useful supplementary role against bancroftian filariasis? Bulletin of the World Health Organization 1999; 77: 138–143.