

Shape dynamics of nearly spherical, multicomponent vesicles under shear flow

Anirudh Venkatesh¹ and Vivek Narsimhan¹ 

¹Davidson School of Chemical Engineering, Purdue University, West Lafayette, IN, 47906, USA

Corresponding author: Vivek Narsimhan, vnarsim@purdue.edu

(Received 22 September 2024; revised 8 July 2025; accepted 16 August 2025)

In biology, cells undergo deformations under the action of flow caused by the fluid surrounding them. These flows lead to shape changes and instabilities that have been explored in detail for single component vesicles. However, cell membranes are often multicomponent in nature, made up of multiple phospholipids and cholesterol mixtures that give rise to interesting thermodynamics and fluid mechanics. Our work analyses shear flow around a multicomponent vesicle using a small-deformation theory based on vector and scalar spherical harmonics. We set up the problem by laying out the governing momentum equations and the traction balance arising from the phase separation and bending. These equations are solved along with a Cahn–Hilliard equation that governs the coarsening dynamics of the phospholipid–cholesterol mixture. We provide a detailed analysis of the vesicle dynamics (e.g. tumbling, breathing, tank-treading and swinging/phase-treading) in two regimes – when flow is faster than coarsening dynamics (Péclet number $Pe \gg 1$) and when the two time scales are comparable ($Pe \sim O(1)$) – and provide a discussion on when these behaviours occur. The analysis aims to provide an experimentalist with important insights pertaining to the phase separation dynamics and their effect on the deformation dynamics of a vesicle.

Key words: membranes, capsule/cell dynamics, phase change

1. Introduction

Flow around biological membranes holds a great deal of importance in a multitude of systems (Seifert 1999; Herzenberg *et al.* 2006; Shi *et al.* 2018; Salmond *et al.* 2021). These biological membranes are modelled using surrogate structures known as vesicles. Vesicles are complex droplets having a lipid bilayer on the surface instead of simple fluid boundaries. The presence of this lipid bilayer imparts an elastic resistance to bending (Helfrich 1973). These lipid bilayer membranes act like two-dimensional (2-D) fluid

sheets that resist any stretching or compression (Campelo *et al.* 2014). Apart from being a marvellous surrogate system for understanding the biophysics of cellular membranes, lipid vesicles are often used as carriers for drug delivery processes (Needham 1999; Guo & Szoka 2003).

Lipid bilayers are made up for long chain compounds known as phospholipids that contain a hydrophilic head and a hydrophobic tail (Alberts 2017). When the bilayer consists of a single type of phospholipid, we call them ‘single component’ vesicles. These vesicles have been the centre of a plethora of studies over the past five decades. The physical properties and thermodynamics of such vesicles can be characterized by a lipid bilayer exhibiting a uniform bending resistance (Lipowsky & Sackmann 1995). Previous studies have explored the dynamics of such single component vesicles in great detail by studying multiple modes of vesicle motion in different systems: (i) shear flow – tank-treading (Keller & Skalak 1982; Seifert 1999; Abkarian, Lartigue & Viallat 2002), swinging (Noguchi & Gompper 2007), tumbling (Biben & Misbah 2003; Rioual, Biben & Misbah 2004; Noguchi & Gompper 2005; Kantsler & Steinberg 2006) and vacillated breathing (Misbah 2006); (ii) extensional flow (Zhao & Shaqfeh 2013; Narsimhan *et al.* 2014, 2015; Dahl *et al.* 2016; Kumar, Richter & Schroeder 2020); (iii) general linear flows (Vlahovska & Gracia 2007; Lin & Narsimhan 2019); (iv) oscillatory flows (Lin *et al.* 2021).

The deep understanding of single component vesicles has provided a base for further explorations into systems that are closer to reality. Often, these lipid bilayer membranes contain multiple phospholipids along with cholesterol molecules interspersed between them (John *et al.* 2002). The existence of multiple molecules in these bilayers makes for an incredible amalgamation of phase equilibrium thermodynamics and fluid mechanics (Safran 2018). Some of these phospholipids (e.g. dipalmitoylphosphatidylcholine (DPPC)) have a larger affinity towards cholesterol molecules than others (e.g. 1,2-Dioleoyl-sn-glycero-3-phosphocholine (DOPC)) (Veatch & Keller 2005a; Davis, Clair & Juhasz 2009; Uppamoochikkal, Tristram-Nagle & Nagle 2010). This preferential separation into phases leads to the formation of ordered and disordered liquid phases on the membrane surface (Veatch & Keller 2005b), a term known as ‘lipid rafts’ (Simons & Ikonen 1997). These rafts have a relevance in signal transduction (Simons & Toomre 2000) and protein transfer across membranes, thereby having implications for health and diseases (Michel & Bakovic 2007). These factors underscore the importance of studying such systems.

From a mechanical viewpoint, the existence of multiple phases imparts inhomogeneous properties like the bending rigidity to the vesicle due to the differences in bending stiffnesses of each constituent phospholipid (Baumgart, Hess & Webb 2003). Moreover, the resultant lateral phases fall prey to a tussle between convective motion due to the background fluid and surface diffusive motion due to the inherent molecular properties of the phases (Yanagisawa *et al.* 2007; Taniguchi, Yanagisawa & Imai 2011; Arnold, Gubbala & Takatori 2023).

While creating medical diagnostic devices, often, the measurement of mechanical properties of such vesicles is important (Kollmannsberger & Fabry 2011; Lei *et al.* 2021). These measurements help in improving the control and precision of devices. Previously, multiple numerical simulations have been performed to understand the motion of multicomponent vesicles under shear flow (Sohn *et al.* 2010; Liu *et al.* 2017; Gera & Salac 2018b). These studies have highlighted the influence of line tension and bending rigidity, along with the membrane tension of the vesicle, on the modes of motion that the vesicle undergoes – tank-treading, tumbling, swing/phase-treading, vertical banding, among others. More recently, under the limit of dominant advective forces, researchers came up with an analytical treatment of 2-D multicomponent vesicles and their swinging-to-tumbling transition (Gera, Salac & Spagnolie 2022). The transition

primarily depended on a ratio of the bending stiffness of the two phases and the capillary number of the vesicle. While informative and thoughtful, the study lacked an analytical treatment of the case when diffusive time scales are comparable to that of the convective time scales. Secondly, the study treated a multicomponent vesicle as a 2-D inextensible membrane, thus leaving room for out-of-plane deformations. To the authors' knowledge, such an analytical treatment has not been provided yet.

We aim to bridge this gap of knowledge through this study that focuses on the semianalytical prediction for a three-dimensional (3-D), nearly spherical, multicomponent vesicle placed under a background shear flow. We leverage the vector spherical harmonics-based techniques previously used for single component vesicles and drops (Vlahovska, Loewenberg & Blawdziewicz 2005; Vlahovska & Gracia 2007) to solve the underlying nonlinear dynamical equations up to leading order, while ensuring conservation of the composition and vesicle surface area. This helps us arrive at reduced-order equations governing the shape and composition of the vesicle and the phospholipid–cholesterol phases, respectively. We delineate multiple motions exhibited by the vesicle and discuss their dependence on material properties. The aim of this study is to equip an experimental researcher with a theory that could *a priori* predict the vesicle dynamics based on the material specifications and control variables.

We discuss problem set-up, solution strategy and numerical methods in §§ 2 and 3. We discuss results for multicomponent vesicles in the high Péclet number limit in § 4, where membrane coarsening effects are unimportant. We then discuss results in the intermediate Péclet number limit in § 5, where coarsening effects come into play. We discuss implications of asymmetric phase distributions in § 6. We then conclude the study with a discussion in § 7.

2. Problem formulation

We consider a lipid membrane vesicle containing a ternary mixture of phospholipids and cholesterol undergoing phase separation on the membrane surface (see figure 1). This vesicle is placed in an unbounded domain containing a background shear flow $\mathbf{u}^\infty = \dot{\gamma} y \hat{\mathbf{x}}$. This membrane encloses a Newtonian fluid of viscosity $(\lambda - 1)\eta$ with a surrounding fluid of viscosity η , where $\lambda - 1$ is the viscosity ratio between the inner and outer fluids. The bilayer has a bending stiffness of κ_c that is dependent on the phospholipid distribution characterized by an order parameter q . This dependence will be explained in § 2.1. Furthermore, the lipid molecules impose a membrane tension to preserve the membrane surface area A and there exists a line tension between the separated phases (indicated by black and white coloured lipid heads). The vesicle has a characteristic size R which is the radius of a sphere of the same volume.

2.1. Free energy of membrane

The lipid membrane energy landscape has been a problem of interest for the past few decades. Historically, a lipid bilayer membrane is treated as an elastic surface. Multiple models have been used to describe such materials – the Keller Skalak model (Keller & Skalak 1982), area difference model (Seifert 1997) and Helfrich model (Helfrich 1973; Campelo *et al.* 2014) to name a few. In our study, we use a Canham–Helfrich model.

The free energy of a multicomponent membrane is governed by three factors: bending, mixing and surface tension energies. The bending energy is given by

$$W_{\text{bend}} = \int \frac{1}{2} \kappa_c (2H - c_0)^2 dS, \quad (2.1)$$

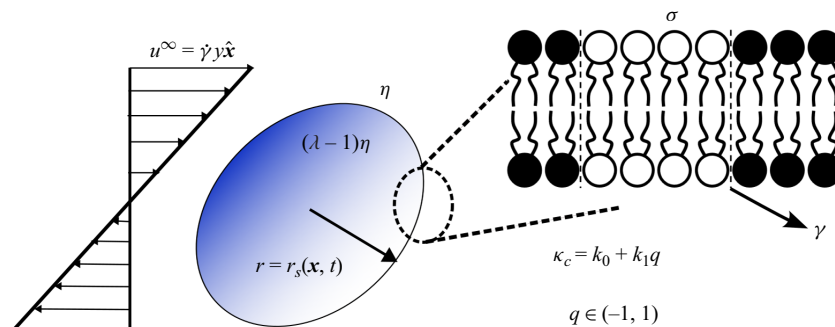


Figure 1. Schematic of system. The inset shows a zoomed-in version of the lipid bilayer and its properties (see § 2.1).

where $\kappa_c = k_0 + k_1 q$ is a bending modulus that depends on the lipid phase behaviour characterized by an order parameter q . The order parameter q is a variable whose values can range from -1 corresponding to pure L_d (cholesterol-lacking) and $+1$ corresponding to pure L_o (cholesterol-rich). Thermodynamically speaking, this is the local coordinate along a tie line in the two-phase coexistence region that is present in the ternary phase diagram for a system like DOPC, DPPC and cholesterol. A tie line represents the line that connects the compositions of the ordered and disordered phases at equilibrium, as commonly seen in liquid–liquid extraction systems (Geankoplis 2003). The values of k_0 and k_1 are $k_0 = 1/2(\kappa_{lo} + \kappa_{ld})$ and $k_1 = 1/2(\kappa_{lo} - \kappa_{ld})$, where κ_{lo} and κ_{ld} are the bending moduli of the L_o and L_d phases, respectively. Lastly, the parameters H and c_0 are the mean and spontaneous curvatures of the bilayer leaflet. We treat $c_0 = 0$ for a symmetric leaflet.

The mixing energy is given by the Landau–Ginzberg equation (Gompper & Klein 1992):

$$W_{mix} = \int \left(\frac{\tilde{a}}{2} q^2 + \frac{\tilde{b}}{4} q^4 + \frac{\gamma^2}{2} |\nabla_s q|^2 \right) dS. \quad (2.2)$$

The first two terms represent a double-well potential that gives rise to phase separation (for $\tilde{a} < 0$), and the last term is an energy penalty for creating phases that is related to line tension. This free energy has been used in many studies of lipid systems undergoing phase separation. These parameters are related to the experimentally measured phase split concentrations ($q^\pm$), line tension (ξ^{line}) and the interface width (ε^{width}) as follows:

$$q^\pm = \pm \sqrt{|\tilde{a}|/|\tilde{b}|}, \quad \xi^{line} = \frac{2\sqrt{2}}{3\tilde{b}} |\tilde{a}|^{3/2} \gamma, \quad \varepsilon^{width} = \sqrt{\frac{2\gamma^2}{|\tilde{a}|}}. \quad (2.3)$$

The Landau–Ginzberg equation has been used to qualitatively model bilayer membranes (Gera & Salac 2018b) – see the appendix of Camley & Brown (2014) for estimated values of $\tilde{a}$, $\tilde{b}$ and γ for a specific system ($R \sim O(nm)$). In their study, they mention that an approximate value for $q^+ \approx 0.3$ would be appropriate for phase-split concentrations.

Lastly, the surface tension energy is given by

$$W_\sigma = \int \sigma dS = \text{constant}. \quad (2.4)$$

The above contribution is constant because the lipid area per molecule is conserved and thus the membrane is incompressible. The surface tension σ thus acts as a Lagrange multiplier to ensure area conservation.

2.2. Fluid motion and membrane interface dynamics

We solve the Stokes equations inside and outside the vesicle that govern the velocity, stress and pressure distributions:

$$(\lambda - 1)\eta\nabla^2\mathbf{u}^{in} = \nabla p^{in}; \quad \nabla \cdot \mathbf{u}^{in} = 0; \quad (2.5a)$$

$$\eta\nabla^2\mathbf{u}^{out} = \nabla p^{out}; \quad \nabla \cdot \mathbf{u}^{out} = 0. \quad (2.5b)$$

These equations are subject to the following boundary conditions: (i) far-field, $\mathbf{u}^{out} \rightarrow \mathbf{u}^\infty$ as $|\mathbf{x}| \rightarrow \infty$; (ii) continuity of velocity, $\mathbf{u}^{in} = \mathbf{u}^{out}$ for $\mathbf{x} \in S$; (iii) membrane incompressibility, $\nabla_s \cdot \mathbf{u}^{in} = 0$ for $\mathbf{x} \in S$; (iv) traction balance, $\mathbf{f}^{hyd} = \mathbf{f}^{mem}$ for $\mathbf{x} \in S$.

In the traction balance, $\mathbf{f}^{hyd} = \mathbf{n} \cdot (\mathbf{T}^{out} - \mathbf{T}^{in})$ is the hydrodynamic traction from viscous stresses, where $\mathbf{n}$ is the outward-pointing normal vector and $\mathbf{T} = \mu(\nabla\mathbf{u} + \nabla\mathbf{u}^T) - p\mathbf{I}$ is the stress tensor for a fluid with viscosity μ ($\mu = \eta$ outside the vesicle, $\mu = (\lambda - 1)\eta$ inside the vesicle). The membrane traction $\mathbf{f}^{mem}$ is equal to the first variation of the membrane free energy with respect to position: $\mathbf{f}^{mem} = \delta W / \delta \mathbf{x}$. This can be broken into bending, mixing and surface tension contributions $\mathbf{f}^{mem} = \mathbf{f}^{bend} + \mathbf{f}^{mix} + \mathbf{f}^\sigma$, with expressions for each shown as follows:

$$\begin{aligned} \mathbf{f}^{bend} = \frac{\delta W_{bend}}{\delta \mathbf{x}} = & -\mathbf{n} \nabla_s^2 (\kappa_c(2H)) + \kappa_c(2H) \left[\nabla_s(2H) + (2K - 4H^2) \right] \mathbf{n} \\ & + 2H \left(\frac{1}{2} \kappa_c(2H)^2 \right) \mathbf{n} - \nabla_s \left(\frac{1}{2} \kappa_c(2H)^2 \right), \end{aligned} \quad (2.6a)$$

$$\begin{aligned} \mathbf{f}^{mix} = \frac{\delta W_{mix}}{\delta \mathbf{x}} = & \gamma^2 \nabla_s \cdot (\nabla_s q \otimes \nabla_s q) + 2H \left(\frac{1}{2} \gamma^2 |\nabla_s q|^2 + g(q) \right) \mathbf{n} \\ & - \nabla_s \left(\frac{1}{2} \gamma^2 |\nabla_s q|^2 + g(q) \right), \end{aligned} \quad (2.6b)$$

$$\mathbf{f}^\sigma = \frac{\delta W_\sigma}{\delta \mathbf{x}} = 2\sigma H \mathbf{n} - \nabla_s \sigma. \quad (2.6c)$$

The reader is directed to the following publications for details on how these equations are derived: Napoli & Vergori 2010 and Gera 2017. In the above expressions, $\otimes$ represents a dyadic product, $\nabla_s = (\mathbf{I} - \mathbf{nn}) \cdot \nabla$ is the surface gradient and $g = (\tilde{a}/2)q^2 + (\tilde{b}/4)q^4$ is the double well potential in the mixing free energy (2.2). The quantities $K = \det(\mathbf{L}) = C_1 C_2$ and $H = (1/2)\text{tr}(\mathbf{L}) = (C_1 + C_2)/2$ are the Gaussian and mean curvatures of the interface, respectively, where $\mathbf{L} = \nabla_s \mathbf{n}$ is the surface curvature tensor. The surface tension σ is a Lagrange multiplier to enforce membrane incompressibility ($\nabla_s \cdot \mathbf{u} = 0$ on the interface), and hence must be solved in addition to the velocity and pressure fields.

In addition to the flow field, one must also solve for order parameter q on the membrane surface. The flow and coarsening behaviour of the order parameter satisfy a Cahn–Hilliard equation (Gera 2017),

$$\frac{\partial q}{\partial t} + \nabla_s \cdot (q\mathbf{u}) = \frac{\nu}{\zeta_0} \nabla_s^2 \zeta, \quad (2.7a)$$

$$\zeta = \frac{\delta W}{\delta q} = \left(\tilde{a}q + \tilde{b}q^3 - \gamma^2 \nabla_s^2 q + \frac{k_1}{2} (2H)^2 \right), \quad (2.7b)$$

where in the above expression, ν is the mobility of the phospholipids (units of metres squared per second) and $\zeta = \delta W / \delta q$ is the surface chemical potential (units of energy per area). The above expression states that lipids are either convected along the surface or move via gradients in chemical potential. The value reference chemical potential ζ_0 is given in Gera 2017.

Lastly, we impose the kinematic boundary condition to avoid any slip on the membrane surface. If we parameterize the vesicle radius as $r = r_s(\mathbf{x}, t)$, the kinematic boundary condition is below, where $D/Dt = (\partial/\partial t) + \mathbf{u} \cdot \nabla$ is the material derivative:

$$\frac{D}{Dt} (r - r_s) = 0 \quad \mathbf{x} \in S. \quad (2.8)$$

2.3. Time scales and dimensionless quantities

The vesicle dynamics occur over four time scales. The first one is a bending time scale $t_{bend} = R^3 \eta / k_0$ that denotes the time taken by a vesicle to restore its equilibrium configuration under the action of bending forces. The second time scale is that of the flow $t_{\dot{\gamma}} = \dot{\gamma}^{-1}$. The third represents the time scale for coarsening $t_q = R^2 / \nu$ and the fourth represents the rotational time scale $t_r = \lambda \dot{\gamma}^{-1}$. We pick the same characteristic scales for physical quantities as previously used in flow studies for single component vesicles (Vlahovska & Gracia 2007). All lengths are non-dimensionalized by the equivalent radius R , all times by the flow time scale $t_{\dot{\gamma}} = \dot{\gamma}^{-1}$, and all velocities by $U_{flow} = R / t_{\dot{\gamma}} = R \dot{\gamma}$. All pressures and viscous stresses are scaled by $\eta \dot{\gamma}$, whereas the membrane tractions $\mathbf{f}^{mem}$ are scaled by k_0 / R^2 .

Table 1 lists the set of physical parameters for this problem and their typical experimental values, while table 2 lists the dimensionless numbers for this problem. The first three dimensionless numbers are ones that are also found in the single component vesicle literature – the viscosity ratio parameter $\lambda = (\mu^{in} + \mu^{out}) / \mu^{out}$ between the inner and outer fluids, the capillary number $\chi = \eta R^3 \dot{\gamma} / k_0$ relating the viscous to bending forces on the membrane and the dimensionless excess area $\Delta = A / R^2 - 4\pi$ representing the floppiness of the vesicle. The remaining dimensionless numbers occur only in multicomponent vesicles. Of these, the most important ones are the dimensionless bending stiffness difference between the two phases $\beta = k_1 / k_0 = (\kappa_{lo} - \kappa_{ld}) / (\kappa_{lo} + \kappa_{ld})$, the Cahn number $Cn = \gamma / (R \sqrt{\zeta_0})$ (i.e. ratio of line tension energy to the energy scale of phase separation), the surface Péclet number $Pe = R^2 \dot{\gamma} / \nu$ (i.e. ratio of coarsening time from diffusion to flow time) and the line tension parameter $\alpha = k_0 / \gamma^2$ (ratio between bending energy to line tension energy). Note that the Cahn number can be re-expressed as the ratio between the interface thickness and vesicle size $Cn = \epsilon^{width} / (\sqrt{2} R)$. Furthermore, the average concentration q_0 affects the position along the local energy landscape. It also affects stresses arising from the phase energy (2.6b).

3. Solution for a nearly spherical vesicle

3.1. Perturbation expansion

We solve for the vesicle shape and composition as a function of time in the limits of small changes in deformation and concentration. We use $\epsilon_1 = \Delta^{1/2}$ and $\epsilon_2 = q^+$ as perturbation variables with $\epsilon_1 \sim \epsilon_2$, where Δ is the excess area and $q^+ = \sqrt{-a/b}$ is the phase split concentration (see table 2). This approximation (i.e. weak segregation approximation), has been used in multiple studies related to copolymer systems (Leibler 1980; Fredrickson & Helfand 1987; Seul & Andelman 1995), and has been applied to multicomponent vesicles in the past (Taniguchi *et al.* 1994; Kumar, Gompper & Lipowsky 1999). We solve the

Variable	Name	Order of magnitude	Reference
R	Equivalent radius of spherical vesicle	$O(1) \mu\text{m}$	Deschamps <i>et al.</i> (2009)
k_0	Average bending stiffness between two phases	$O(10^{-19} - 10^{-18}) \text{ J}$	Amazon <i>et al.</i> (2013)
k_1	Half of bending stiffness difference between two phases	$O(10^{-19}) \text{ J}$	Amazon <i>et al.</i> (2013)
ν	Mobility of phospholipids	$O(10^{-11}) \text{ m}^2 \text{ s}^{-1}$	Negishi <i>et al.</i> (2008)
γ	Line tension parameter	$O(10^{-9}) \text{ J}^{1/2}$	Luo & Maibaum (2020)
$\dot{\gamma}$	Shear rate	$O(0.1 - 1) \text{ s}^{-1}$	Deschamps <i>et al.</i> (2009)

Table 1. Physical parameter ranges and orders of magnitude.

Variable	Name	Value
$\lambda = (\mu^{in} + \mu^{out})/\mu^{out}$	Viscosity ratio parameter	$O(1 - 10)$
$\chi = \eta R^3 \dot{\gamma} / k_0$	Capillary number	$O(0.01 - 1)$
$\Delta = A/R^2 - 4\pi$	Excess area	$O(0.01 - 0.1)$
$a = \tilde{a}/\zeta_0$	Dimensionless double well potential term	$O(0.01 - 0.1)$
$b = \tilde{b}/\zeta_0$	Dimensionless double well potential term	$O(1)$
$q^+ = \sqrt{-a/b}$	Phase split concentration	$O(0.01 - 0.1)$
$\beta = k_1/k_0$	Ratio of bending stiffnesses	$O(0.1 - 1)$
$Cn = \gamma/(R\sqrt{\zeta_0})$	Cahn number	$O(0.1 - 1)$
$\alpha = k_0/\gamma^2$	Ratio of bending stiffness to line tension	$O(1)$
$Pe = R^2 \dot{\gamma} / \nu$	Péclet number (coarsening time scale/flow time scale)	$O(1 - 1000)$
q_0	Average order parameter	$O(q^+)$

Table 2. Dimensionless parameter ranges and orders of magnitude.

dynamical equations in § 2.2 to leading order, keeping volume ($V = 4\pi/3$) and surface area ($A = 4\pi + \Delta$) conserved to $O(\epsilon_1^2)$ and average order parameter ($q_0 = A^{-1} \int q dA$) conserved to $O(\epsilon_2^2)$.

Briefly, we expand the vesicle shape, surface tension and composition in terms of spherical harmonics. The non-dimensional vesicle quantities (radius r_s , membrane tension σ , and order parameter q) are written as

$$Z(\theta, \phi) = Z_0 + \sum_{l=1}^{\infty} \sum_{m=-l}^{+l} Z_{lm} Y_{lm}(\theta, \phi) \quad (3.1)$$

where $Z \equiv (r_s, \sigma, q)$. In the above equation, $Y_{lm}(\theta, \phi)$ are spherical harmonics (Appendix A), while $Z_{lm} = (f_{lm}, \sigma_{lm}, q_{lm})$ are coefficients for shape, surface tension and composition to be solved, with $f_{lm}, \sigma_{lm} \sim O(\epsilon_1)$ and $q_{lm} \sim O(\epsilon_2)$, with $\epsilon_1 \sim \epsilon_2$. The isotropic quantities $Z_0 = (r_0, \sigma_0, Q_0)$ are determined by applying the constraints of constant volume, constant area, and average order parameter. We obtain $r_0 = 1 - 1/4\pi \sum_{lm} f_{lm} f_{lm}^*$ and $Q_0 = q_0 - 1/2\pi \sum_{lm} q_{lm} f_{lm}^*$. The isotropic surface tension σ_0 is obtained implicitly from the expression $\sum_{lm} (l+2)(l-1) f_{lm} f_{lm}^* = \Delta$, which arises from conserving area to $O(\epsilon_1^2)$.

3.2. Solving Stokes equations

We solve the Stokes equations around the vesicle to determine how the shape and surface tension coefficients (f_{lm} , σ_{lm}) evolve over time. This section provides a high level overview of the steps, with algebraic details provided in [Appendix B](#).

We Taylor expand the membrane tractions $\mathbf{f}^{mem}$ in (2.6) to $O(\Delta^{1/2})$ on the surface of a sphere. We then solve Stokes equations with this traction boundary condition ($\chi \mathbf{f}^{hydro} = \mathbf{f}^{mem}$ at $r = 1$), along with boundary conditions of continuity of velocity across the interface ($\mathbf{u}^{out} = \mathbf{u}^{in}$ at $r = 1$), membrane incompressibility ($\nabla_s \cdot \mathbf{u} = 0$ at $r = 1$) and far-field velocity ($\mathbf{u}^{out} \rightarrow y\hat{\mathbf{x}}$ as $r \rightarrow \infty$). We note that this idea is commonplace in studies of single component vesicles (Misbah 2006; Vlahovska & Gracia 2007) – the difference here is that we are examining a situation where $\mathbf{f}^{mem}$ takes a more complicated form (see § 2.2). Stokes flow is solved using fundamental basis sets for Lamb’s solution – i.e. a series solution using vector spherical harmonics. We use the notation in Vlahovska *et al.* (2005) and Bławdziewicz *et al.* (2000), details are found in [Appendix B](#).

Once one performs this procedure, one applies the kinematic boundary condition (2.8). Doing so yields the differential equation for the shape mode f_{lm} of the vesicle:

$$\frac{df_{lm}}{dt} = \frac{im}{2} f_{lm} + c_{lm2}^+. \quad (3.2)$$

The first term on the right-hand side comes from the rigid body rotation from shear flow, while the next term (c_{lm2}^+) comes from the extensional deformation, which depends on the shape modes f_{lm} , concentration modes q_{lm} and isotropic surface tension σ_0 . The final results are presented here,

$$c_{lm2}^+ = \lambda^{-1} C_{lm} + \lambda^{-1} \chi^{-1} [(A_l + B_l \sigma_0) f_{lm} + D_l q_{lm}] \quad (3.3)$$

where the coefficients (A_l , B_l , C_{lm} , D_l) are listed in (B10)–(B11) in [Appendix B](#). These depend on the mode number (l , m) as well as the dimensionless quantities discussed in [table 2](#).

We note that the above expression (3.3) depends on the isotropic surface tension σ_0 , which one obtains by applying constraint of constant excess area given by $d\Delta/dt = 0$, where the excess area is given by $\Delta = \sum_{lm} (l+2)(l-1) f_{lm} f_{lm}^*$. This constraint couples all modes in the differential equation for the shape in (3.2).

3.3. Solving the Cahn–Hilliard equation

To determine how the concentration modes q_{lm} evolve over time, we solve the Cahn–Hilliard equation (2.7). In the perturbative limits $\Delta \ll 1$, $q^+ \ll 1$, we Taylor expand all quantities to $O(\Delta^{1/2})$ and $O(q^+)$ on the unit sphere except the terms coming from the double-well potential (i.e. $1/2 * a q^2 + 1/4 * b q^4$), where we keep all higher-order terms in the chemical potential. The reason why we perform this task is that such higher-order terms are needed to have a quartic free energy expression, which is necessary to have two-phase coexistence with a tie line. We note that similar approximations have been applied for equilibrium studies of multicomponent vesicles (Luo & Maibaum 2020). In the cited study, the free energies are Taylor expanded to quadratic order in the shape and concentration modes except the double well term, where modes are kept to quartic order. This yields tractions and chemical potentials to the same order of accuracy as the current study.

The Cahn–Hilliard equation becomes

$$\begin{aligned} \frac{dq_{lm}}{dt} = & \frac{im}{2} q_{lm} - \frac{1}{Pe} [A_{lm} l(l+1) + C n^2 l^2 (l+1)^2 q_{lm} \\ & + 2\alpha C n^2 \beta (l-1) l(l+1)(l+2) f_{lm}] , \end{aligned} \quad (3.4)$$

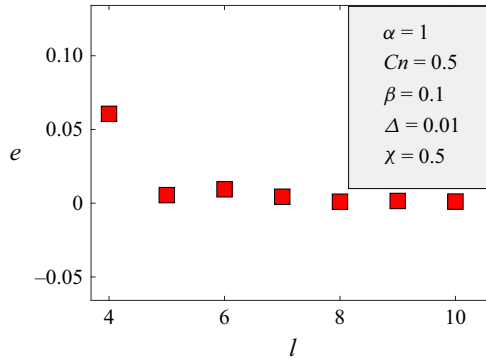


Figure 2. Error convergence plots for $Cn = 0.5$. The parameters for the simulations are $\alpha = 1$, $\beta = 0.1$, $\Delta = 0.01$, $\chi = 0.5$, $a = -0.1$, $b = 1$, $q_0 = 0$, $Pe = 5$, $\lambda = 2$. The y-axis represents the ratio of the difference in the order parameter over the vesicle surface divided by the average magnitude of the order parameter of the case with most modes $l_{max} = 11$.

where $\Lambda_{lm} = \int_{\Omega} (aq + bq^3) Y_{lm} d\Omega$ is the chemical potential from the double-well potential, projected onto spherical harmonics by integrating over a unit sphere. This term is weakly nonlinear as discussed above.

3.4. Numerical procedure

The final equations we solve are (3.2) for the vesicle shape along with conservation of area, as well as (3.4) for the concentration. These are coupled, nonlinear ordinary differential equations (ODEs).

When solving these equations, we decompose the shape and concentration modes into real and imaginary parts

$$f_{lm} = f'_{lm} + i f''_{lm}, \quad q_{lm} = q'_{lm} + i q''_{lm}, \quad (3.5)$$

and perform an ODE solve for the components $(f'_{lm}, f''_{lm}, q'_{lm}, q''_{lm})$. We only solve for components with $m \geq 0$ since for the radius to be real, $f'_{l-m} = (-1)^m f'_{lm}$ and $f''_{l-m} = (-1)^{m+1} f''_{lm}$, with similar relations holding for the concentration. Since we need to ensure that the constraint of area conservation is satisfied $A = 4\pi + \Delta$, utilizing regular solvers is not feasible. We have developed a predictor–corrector scheme to solve the ODEs as mentioned in Appendix C. After solving for the spectral coefficients, we reconstruct the vesicle shape and concentration fields using the spherical harmonic series in (3.1). A benchmark is provided in Appendix D for examining coarsening on a rigid sphere in the absence of flow. We have also benchmarked our results against those of previous studies pertaining to single component vesicles (Vlahovska & Gracia 2007).

For most of the simulations we perform, we choose a cutoff mode number between $l_{max} = 6$ and $l_{max} = 10$ when computing the order parameter. We find that beyond this number, the dominant modes $0 \leq l \leq 4$ for shape and order parameter change less than 2 %, and thus the trends discussed in the results section do not change appreciably. Figure 2 shows an example of a convergence test. Here, we computed the order parameter $q(\theta, \phi, t)$ using different cutoff modes, and compared the results with a simulation using a high cutoff mode. The relative error we computed was $e = \int_S (\bar{q}_{approx} - \bar{q}_{true}) dA / \int_S \bar{q}_{true} dA$, where $\bar{q}_{true}$ is the time-averaged value at (θ, ϕ) using the highest cutoff mode and $\bar{q}_{approx}$ is the time-averaged value at (θ, ϕ) using the lower cutoff mode. For small values of Cn ($Cn \ll 1$), we expect that a spectral method may lead to inaccuracies as one will encounter the sharp interface limit that is hard to resolve owing to the Gibbs phenomenon.

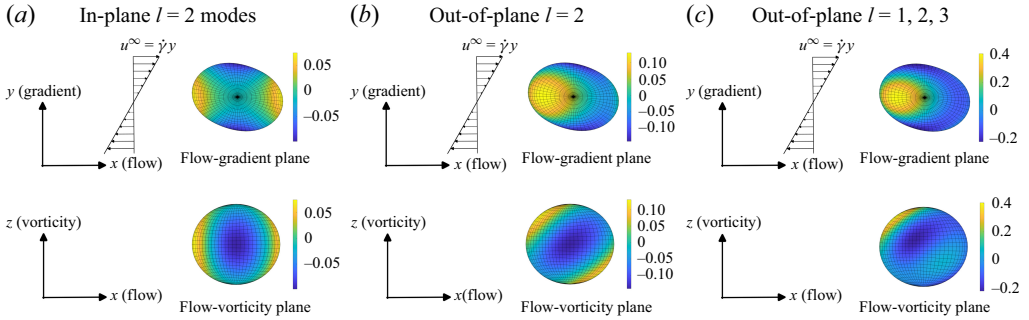


Figure 3. Initial conditions for $Pe \gg 1$ simulations. (a) In-plane $l = 2$ modes, $f'_{22} = f''_{22} = f_{20} = \sqrt{0.1\Delta}$, $q'_{22} = q''_{22} = q_{20} = \sqrt{\Delta/8}$; (b) out-of-plane $l = 2$ modes, $f'_{22} = f''_{22} = f_{20} = \sqrt{0.1\Delta}$; $q'_{22} = q''_{22} = q_{20} = \sqrt{\Delta/8}$, $q'_{21} = q''_{21} = q_{21} = \sqrt{\Delta/8}$; (c) One lobe initial condition with $l \neq 2$ concentration modes, $f'_{22}(0) = f''_{22}(0) = f_{20}(0) = \sqrt{\Delta/10}$, and $q'_{lm}(0) = \sqrt{\Delta/8}$, $q''_{lm}(0) = 0$; $l = 1, 2, 3, m = 0, \dots, l$.

4. Multicomponent vesicles, $Pe \gg 1$

4.1. Overview and initial conditions

This section will examine multicomponent vesicles in the limit of large Péclet number ($Pe \gg 1$), where the flow time scale is much faster than the coarsening time scale. In this regime, coarsening effects are unimportant, and the dynamics is determined by the inhomogeneous bending rigidity set by the initial condition. This regime has previously been studied for 2-D vesicles (Gera *et al.* 2022) where motions such as tumbling and swinging have been observed. However, we are unaware of similar studies for 3-D systems.

One question that naturally arises is whether the 3-D geometry will alter the dynamics significantly compared with the 2-D case. To answer this question, we will structure this section as follows. We will first examine initial conditions for concentration (q_{lm}) and shape (f_{lm}) where the $l = 2$ modes are excited, giving rise to a prolate-like shape with two lobes of a stiff (L_o) phase. These were argued to be the dominant modes in the 2-D study. We will first excite the $l = 2$ modes that are symmetric about the flow-shear plane ($l = 2, m = 0, \pm 2$), replicating conditions similar to a 2-D study, and then excite the $l = 2$ modes that do not obey this symmetry – coined out-of-plane modes (see figures 3a and 3b). We will conclude by examining initial conditions in which many $l \neq 2$ modes are excited for concentration (figure 3c).

Overall, when the initial shape and concentration profiles obey in-plane symmetry, one sees in-plane tumbling and swinging behaviour very similar to the 2-D studies. When out-of-plane modes are excited, one will also see tumbling and swinging, but the swinging can exhibit weak, out-of-plane oscillations, while the tumbling can occur out of plane. If the initial concentration profile is not symmetric with respect to the flow-gradient plane, the profile will remain non-symmetric and will retain the same number of lobes as the initial condition (thus, the concentration solution is not limited to the $l = 2$ subspace). Details are provided below.

4.2. Exciting $l = 2$ modes in-plane

This section explores the effects of exciting the $l = 2$ modes in-plane for shape and concentration. This initial condition corresponds to the situation mentioned in figure 3(a). In this case, an analytical theory for in-plane dynamics can be developed, similar to previously developed 2-D theories (Gera *et al.* 2022).

When $Pe \rightarrow \infty$, the diffusive coarsening effects are negligible and the phospholipids are convected by the background shear flow. Setting $Pe^{-1} = 0$ in (3.4) yields

$$\frac{dq_{lm}}{dt} = \frac{im}{2} q_{lm}, \quad (4.1)$$

and hence the solution is oscillatory: $q_{lm}(t) = q_{lm}(0) \exp(imt/2)$. The shape of the vesicle is then given by (3.2) along with conservation of area constraint, using the above expression for q_{lm} . We will solve for the shape and concentration modes for $l = 2, m = \pm 2$. We find that the $l = 2, m = 0$ shape mode (f_{20}) quickly decays to zero.

Since the dynamics are in plane, we will examine the equatorial plane of the vesicle ($\theta = \pi/2$) and decompose the shape and concentration as follows (using the notation of Gera *et al.* 2022):

$$r_s(\theta = \pi/2, \phi, t) = 1 + \epsilon_1 a_2(t) \cos(2\phi) + \epsilon_1 b_2(t) \sin(2\phi), \quad (4.2a)$$

$$q(\theta = \pi/2, \phi, t) = \epsilon_2 \cos(2\phi + t), \quad (4.2b)$$

where a_2 and b_2 are time dependent coefficients to be determined, and $\epsilon_1 = \Delta^{1/2}$ and $\epsilon_2 = q^+$ are the small parameters in this problem. In terms of the coefficients (f_{lm}, q_{lm}) discussed in the paper, $f_{2\pm 2} = 2\epsilon_1 \sqrt{2\pi/15} (a_2 \mp i b_2)$ and $q_{2\pm 2} = 2\epsilon_2 \sqrt{2\pi/15} \exp(\pm it)$. If we define a modified capillary number as $C = \chi/\epsilon_1$ and modified time $\tau = t/\epsilon_1$, we find the leading-order shape dynamics in (3.2) for $C \sim O(1)$, $\tau \sim O(1)$ and $\epsilon_2/\epsilon_1 \sim O(1)$ to be

$$\begin{aligned} \frac{da_2}{d\tau} &= -\Theta \left(a_2, b_2, \epsilon_1 \tau, \frac{\epsilon_2}{\epsilon_1}, C, \lambda, \beta \right) b_2, \\ \frac{db_2}{d\tau} &= \Theta \left(a_2, b_2, \epsilon_1 \tau, \frac{\epsilon_2}{\epsilon_1}, C, \lambda, \beta \right) a_2. \end{aligned} \quad (4.3)$$

These sets of ODEs take the same form as the 2-D case, except that the expression Θ is different due to the geometry of the problem. Appendix E shows the algebra to obtain Θ – here we state the final results. For the 3-D system, we obtain

$$\Theta_{3D} = \frac{512\pi}{5(9+23\lambda)} \left[\frac{5}{8} a_2 + \frac{\epsilon_2}{\epsilon_1} \beta C^{-1} (a_2 \sin(\epsilon_1 \tau) + b_2 \cos(\epsilon_1 \tau)) \right], \quad (4.4)$$

while for the 2-D system, we obtain

$$\Theta_{2D} = \frac{3\pi}{2\lambda} \left[a_2 + \frac{\epsilon_2}{\epsilon_1} \beta C^{-1} (a_2 \sin(\epsilon_1 \tau) + b_2 \cos(\epsilon_1 \tau)) \right], \quad (4.5)$$

where in the 2-D case the small parameter ϵ_1 is related to the excess length as $\epsilon_1 = \Delta_{2D}^{1/2}$ (see Appendix E). The expressions for Θ depend on the viscosity ratio λ , reduced area $\epsilon_1^2 = \Delta$ and a lumped bending stiffness parameter $S = (\epsilon_2/\epsilon_1) \beta C^{-1} = \beta q^+/\chi$. Results are discussed below.

The system of equations (4.3) admit a periodic solution for the shape modes $a_2(t)$ and $b_2(t)$. Two different behaviours are seen:

- (i) swinging – after an initial transient, the vesicle undergoes weak oscillations in its inclination angle ϕ_i about a fixed value, while the concentration field performs a full rotation along the membrane (for visualization, see figure 4);
- (ii) tumbling – the vesicle shape and concentration field undergo in-plane rigid body rotation (for visualization, see figure 5).

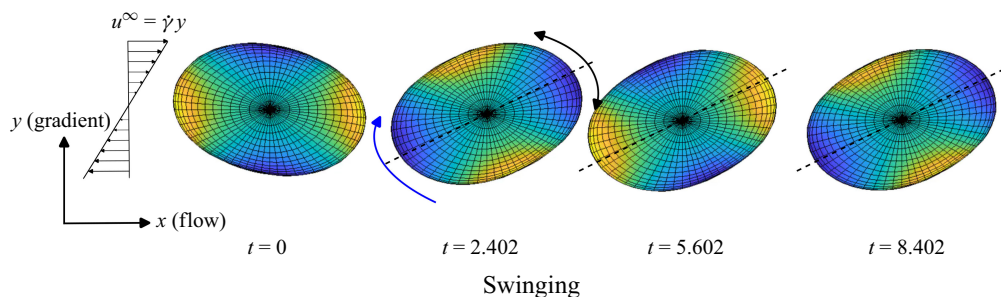


Figure 4. Swinging visualization in flow-gradient plane for $Pe \gg 1$. The initial condition is figure 3(a), and the parameters are $Pe = 10^4$, $a = -0.1$, $b = 1$, $Cn = 0.4$, $\alpha = 1$, $\beta = 0.5$, $\Delta = 0.1$, $\chi = 0.6$, $q_0 = 0$, $\lambda = 2$. The blue phase represents the softer phase and the yellow phase represents the stiffer phase. The black double arrow represents oscillations about a fixed axis (dashed line) after an initial transient. The blue arrow represents the motion of the phases around the vesicle. See Supplementary movie (Video1.mp4) for colourbar details.

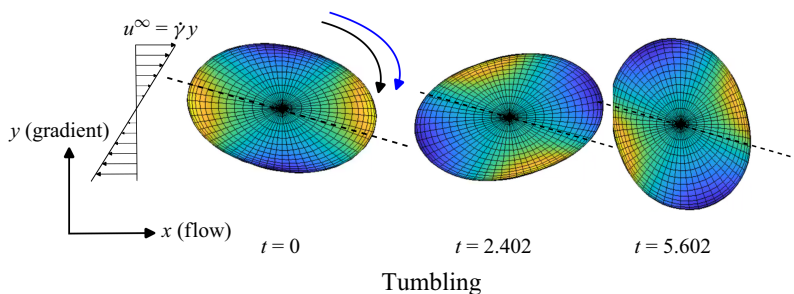


Figure 5. Tumbling visualization in the flow-gradient plane for $Pe \gg 1$. The initial condition is figure 3(a), and the parameters are $Pe = 10^4$, $a = -0.1$, $b = 1$, $Cn = 0.4$, $\alpha = 1$, $\beta = 0.5$, $\Delta = 0.1$, $\chi = 0.01$, $q_0 = 0$, $\lambda = 2$. The blue phase represents the softer phase and the yellow phase represents the stiffer phase. The black and blue arrows represent rigid body motion of the shape and concentration field, while the dashed line is the initial orientation. See Supplementary movie (Video2.mp4) for colourbar details.

For a fixed viscosity ratio λ and excess area Δ , the transition between the two behaviours is determined by the dimensionless quantity $S = (\epsilon_2/\epsilon_1)\beta C^{-1} = \beta q^+/\chi$. Below a critical value of this parameter, swinging occurs, while tumbling occurs above the critical value. Figure 6 illustrates a typical phase diagram. We see that the critical conditions exhibit similar trends for 3-D and 2-D vesicles, although their quantitative values might be different (Gera *et al.* 2022). The explanation for these trends is given by the authors as follows. (i) At low background shear rates, the vesicle elongates in the direction of the softer phase in a quasisteady manner. These regions have a higher curvature. In order to match the phase to the curvature, the vesicle undergoes rigid body tumbling. (ii) On the other hand, when the shear rates are high, the viscous stresses dominate over the elastic stresses thereby pushing the stiffer phase past the high curvature region. In order to account for this phase-shape mismatch, the vesicle undergoes a swinging motion.

Figure 7 shows a plot for $a_2(t)$ and $b_2(t)$ for swinging and tumbling vesicles, comparing the 3-D and 2-D theories in the limit $Pe \rightarrow \infty$. As mentioned, the agreement is not exact due to multiple prefactors entering the 3-D analysis compared with the 2-D case. However, the agreement for tumbling is nearly exact in both cases. The discrepancy primarily arises due to the differences in expressions for Θ_{2D} and Θ_{3D} .

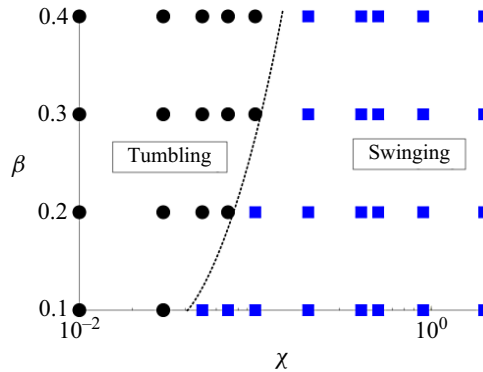


Figure 6. Phase diagram for $Pe \rightarrow \infty$ limit at $\Delta = 0.1$ and $\lambda = 2$, $q^+ = 0.1$.

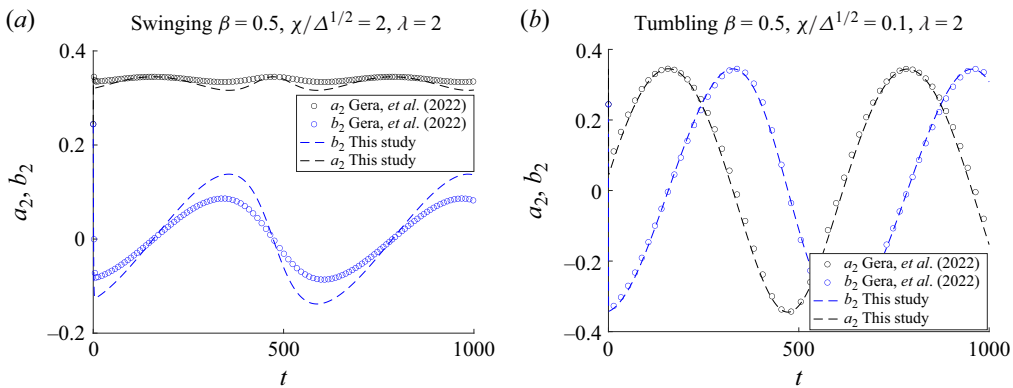


Figure 7. Here $Pe \rightarrow \infty$ results comparison with Gera *et al.* (2022) at $\Delta = 0.0001$ by solving (4.3) and assuming $\epsilon_2 = \epsilon_1 = \Delta^{1/2}$.

Lastly, in [Appendix F](#), we show a comparison between the $Pe \rightarrow \infty$ theory (4.3) and full numerical simulations (solving (3.2) and (3.4)). We see that the analytical theory matches well with numerics for $Pe \sim O(100)$ or above for both tumbling and swinging. We have also seen that shape deformations f_{20} in simulations rapidly decay to zero, indicating that deformations in this mode do not appear to affect the theory quantitatively.

4.3. Effects of higher modes being excited

In this section, we look at effects of exciting q_{lm} modes that are not constrained to $l = 2$. The initial condition considered in the following simulations is $f'_{22}(0) = f''_{22}(0) = f_{20}(0) = \sqrt{\Delta/10}$, and $q'_{lm}(0) = \sqrt{\Delta/8}$, $q''_{lm}(0) = 0$; $l = 1, 2, 3$, $m = 0, \dots, l$. On superimposing these modes, the resultant structure is one lobe containing the stiffer phase and the remaining being primarily the softer phase (see [figure 3c](#)).

Similar to the previous case, swinging is seen where the concentration performs a full rotation along the membrane while the inclination angles exhibit weak oscillations in the shear-flow and the flow–vorticity planes ([figure 8](#)). The modes are plotted in [figure 9](#) to understand the numerical details. We calculate the inclination angles (θ_i, ϕ_i) with respect to the shear-flow plane and shear-vorticity plane using the moment of inertia tensor (Ramanujan & Pozrikidis 1998). The vesicle maintains its one lobe concentration profile

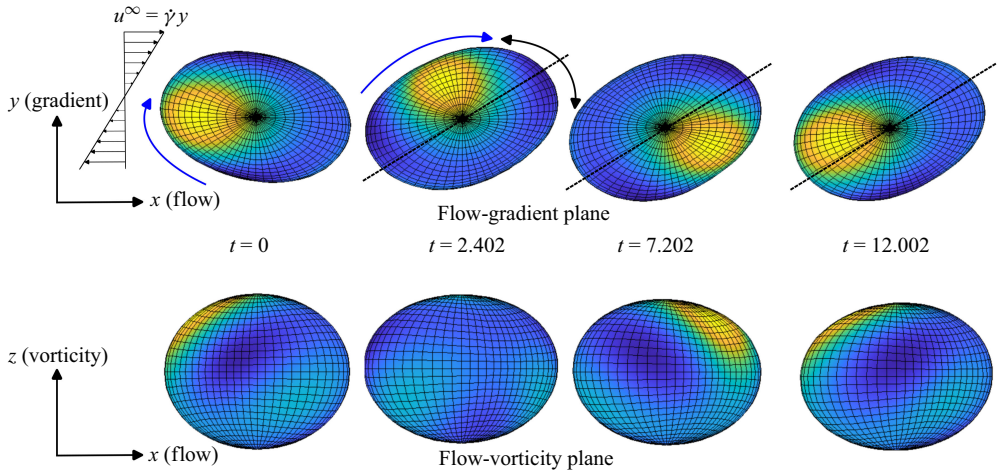


Figure 8. Swinging visualization when higher-order q_{lm} modes are excited for $Pe \gg 1$. The initial condition is [figure 3\(c\)](#), and the parameters for simulations are $\beta = 0.5$, $\alpha = 1$, $a = -0.1$, $b = 1$, $Pe = 10^4$, $\Delta = 0.1$, $\chi = 0.6$, $Cn = 0.4$, $\lambda = 2$, $q_0 = 0$. The blue phase represents the softer phase and the yellow phase represents the stiffer phase. The black double arrow represents oscillations about a fixed axis (dashed line) after an initial transient. The blue arrow represents the motion of the phases around the vesicle. See Supplementary movie ([Video7.mp4](#), [Video8.mp4](#)) for colourbar details.

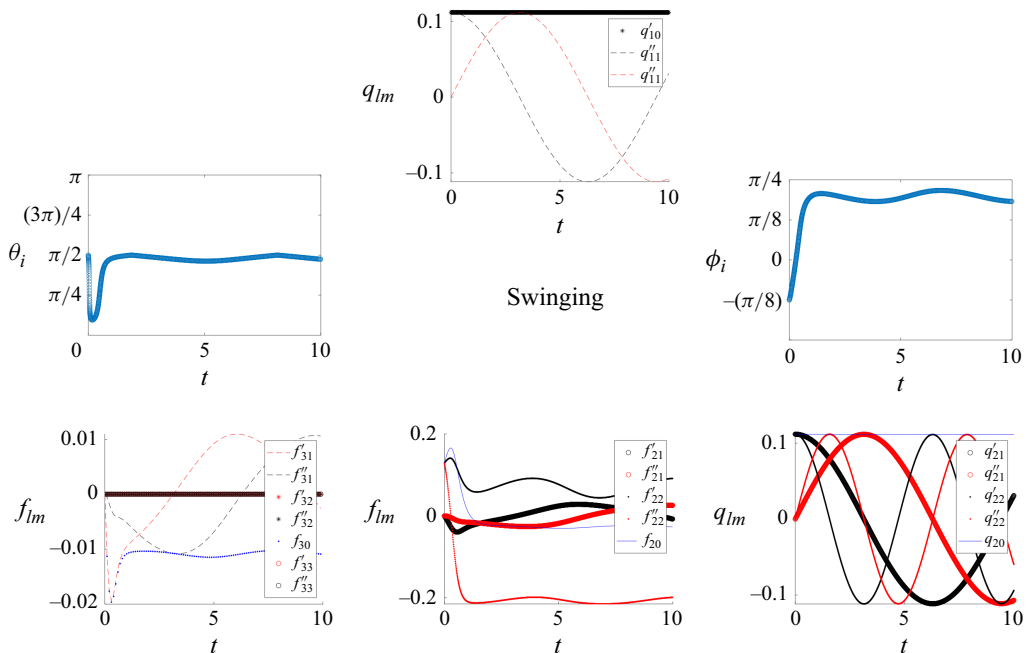


Figure 9. Modes and inclination angles for [figure 8](#) (swinging for $Pe \gg 1$).

during its motion, indicating that the spatial concentration pattern is set by the initial condition.

In the second type of motion (tumbling), as seen in [figure 10](#), the oscillations in the shear-flow plane are significant (rigid-body rotations), whereas the oscillations in the

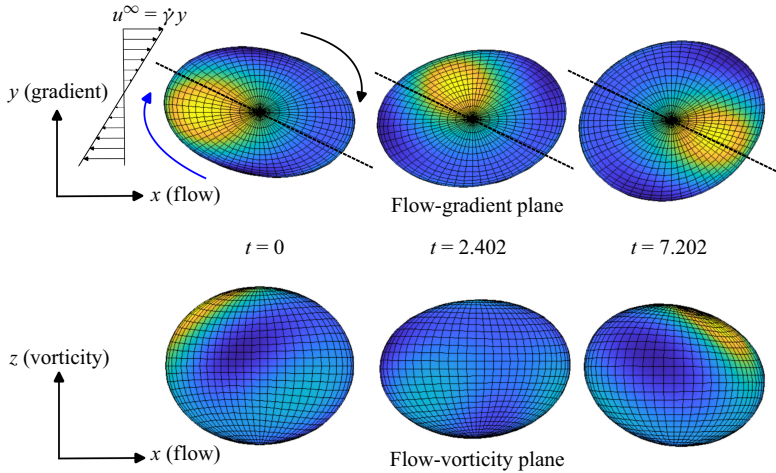


Figure 10. Tumbling visualization when higher-order q_{lm} modes are excited for $Pe \gg 1$. The initial condition is figure 3(c), and the parameters for simulations are $\beta = 0.5$, $\alpha = 1$, $a = -0.1$, $b = 1$, $Pe = 10^4$, $\Delta = 0.1$, $\chi = 0.01$, $Cn = 0.4$, $q_0 = 0$, $\lambda = 2$. The blue phase represents the softer phase and the yellow phase represents the stiffer phase. The black and blue arrows represent rigid body motion of the shape and concentration field, and the dashed line is the initial orientation. See Supplementary movies (Video9.mp4, Video10.mp4) for colourbar details.

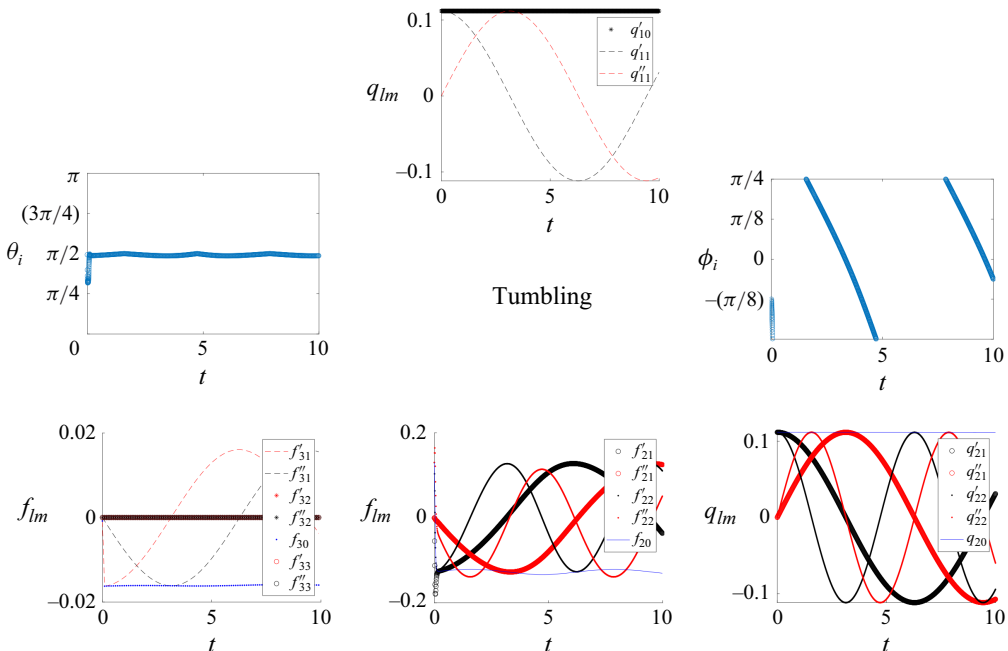


Figure 11. Modes and inclination angles for figure 10 (tumbling for $Pe \gg 1$).

vorticity-flow plane are small. We plot the modes in figure 11. We can observe that the f_{3m} deformations are an order of magnitude smaller than f_{2m} modes which is expected due to the nature of the background flow. This vesicle behaves almost like an oblate-spheroid rotating in flow.

We also performed multiple simulations with initial conditions same as those in figure 3(b). The primary observation is that the nature of motion remains similar to figures 8 and 10, albeit with a different concentration topology. In these simulations, the order parameter fields look like figure 3(b). The movies for such motions can be seen in Video3.mp4 (swinging), Video4.mp4 (swinging), Video5.mp4 (tumbling) and Video6.mp4 (tumbling).

5. Multicomponent vesicles, intermediate Pe

In this section, we focus on vesicles undergoing motion for a $Pe \sim O(1-10)$. Under this regime, the flow and coarsening time scales are comparable, which give rise to a wider range of vesicle shape behaviours compared with the previous section. Additional parameters to consider (beyond the ones discussed previously) are (i) line tension parameter $\alpha = k_0/\gamma^2$, which indicates the relative bending to line tension energy, and (ii) Cahn number $Cn = \epsilon^{width}/(\sqrt{2}R)$, which is related to the interface width compared with the vesicle size. This is also related to line tension. Just like in the previous section, we will divide our analysis based on the initial conditions of the simulations – (i) in plane dynamics (flow-shear), and (ii) out-of-plane dynamics.

5.1. In-plane dynamics

Below summarizes the behaviour observed in the long time limit when one excites the $l = 2$ shape and concentration modes in-plane. The initial condition is given in figure 3(a).

- (i) Tank-treading. Here, both the vesicle's shape and concentration field are stationary. The vesicle is ellipsoidal and fixed at a steady inclination angle ϕ_i , and the concentration field is frozen (figure 12). This behaviour can only be found when coarsening effects are present, since the concentration field in the $Pe \gg 1$ limit only admits periodic solutions (see (4.1)).
- (ii) Tumbling. The vesicle's shape and concentration field exhibit rigid body rotation (figure 13). One can observe that the f_{20} mode decays rapidly, thereby leaving purely in-plane rotations.
- (iii) Vacillated – breathing/trembling. The vesicle performs rotations in the flow-gradient plane while there are significant deformations in the flow–vorticity plane represented by the f_{20} mode (figure 14). A detailed understanding of this motion is given in Misbah (2006).

Figures 12–14 show snapshots of these behaviours, as well as typical plots for the shape modes f_{lm} , concentration modes q_{lm} and in-plane inclination angle ϕ_i . Movies are also provided in the Supplementary material.

Occasionally, we have observed situations where the vesicle undergoes tumbling motion before the motion dampens out at very long times to tank-treading behaviour. This has been noted in figure 15. We believe this is not a numerical instability but arises from the nonlinear mode-mixing term bq^3 in the Cahn–Hilliard equation. It is important therefore to understand what is the time scale at which these motions are observed experimentally. In figure 15, we see that the tumbling motion persists up to $t \sim 10$, which translates to a physical time of $10\dot{\gamma}^{-1}$ that could range from $O(1) - O(10)$ s. Shear flow experiments are usually limited by issues like photobleaching or the escape of the vesicle from the field-of-view. Hence, we believe in order to characterize the motion, the time scale of observation should be reported for a phase diagram. For the other dynamical regimes listed above, we do not observe a change of behaviour at very long times ($t \sim O(100)$).

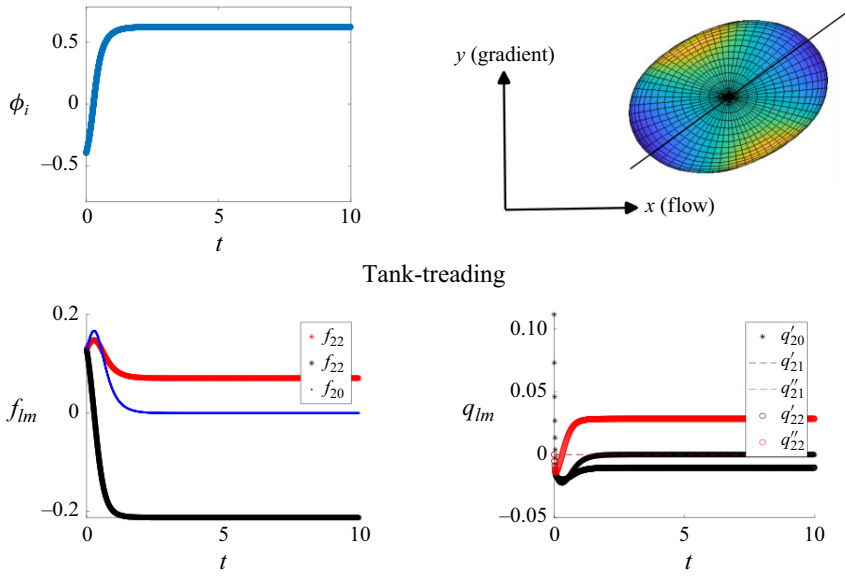


Figure 12. Tank-treading for $Pe \sim O(1)$. The initial condition is [figure 3\(a\)](#), and the parameters are $\chi = 0.1$, $\alpha = 1$, $Cn = 1$, $\beta = 0.1$, $\lambda = 2$, $Pe = 5$, $\Delta = 0.1$, $a = -0.1$, $b = 1$, $q_0 = 0$. The blue phase represents the softer phase, and the yellow phase represents the stiffer phase. The inset snapshot represents the frozen state and inclination angle of the vesicle. See the Supplementary movie ([Video11.mp4](#)) for colourbar information.

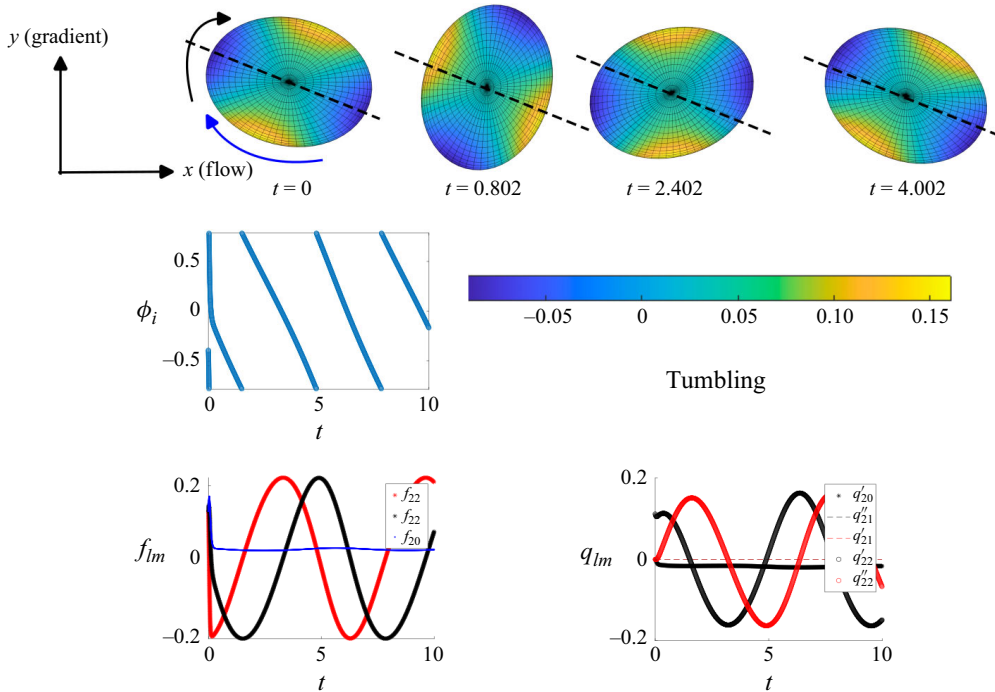


Figure 13. Tumbling for $Pe \sim O(1)$. The initial condition is [figure 3\(a\)](#), and the parameters are $\alpha = 1$, $\beta = 0.5$, $Cn = 0.4$, $\chi = 0.01$, $Pe = 5$, $\lambda = 2$, $\Delta = 0.1$, $a = -0.1$, $b = 1$, $q_0 = 0$. The colour bar represents the order parameter q . The blue phase represents the softer phase and the yellow phase represents the stiffer phase. The blue and black arrows represent rigid-body motion of lipids and shape, and the dashed line is the initial orientation. See Supplementary movie ([Video12.mp4](#)) for visualization.

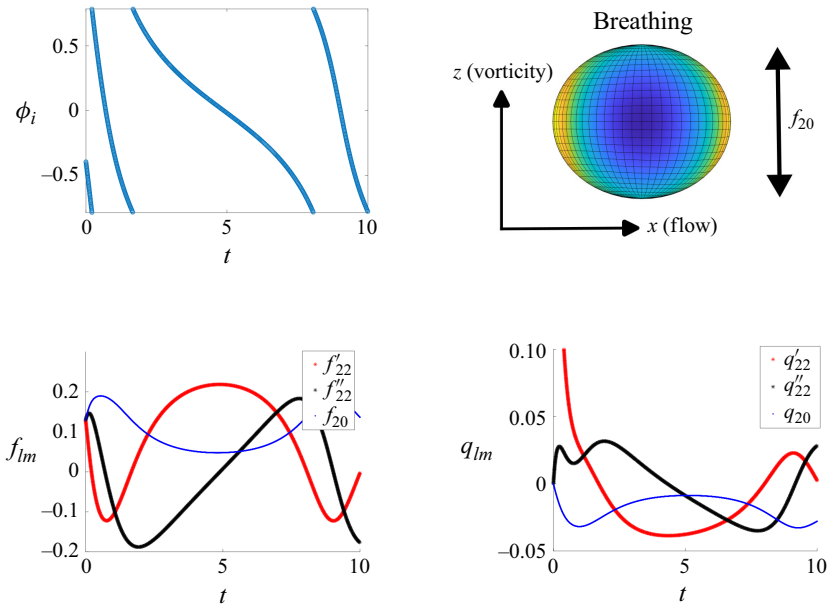


Figure 14. Breathing for $Pe \sim O(1)$. The initial condition is figure 3(a), and the parameters are $\chi = 0.01$, $\alpha = 1$, $Cn = 0.8$, $\beta = 0.1$, $\lambda = 10$, $Pe = 5$, $\Delta = 0.1$, $a = -0.1$, $q_0 = 0$, $b = 1$. The inset snapshot represents the visualization of the vesicle undergoing breathing motion. The blue phase represents the softer phase and the yellow phase represents the stiffer phase. See Supplementary movies (Video13.mp4, Video14.mp4) for visualization and colourbar information.

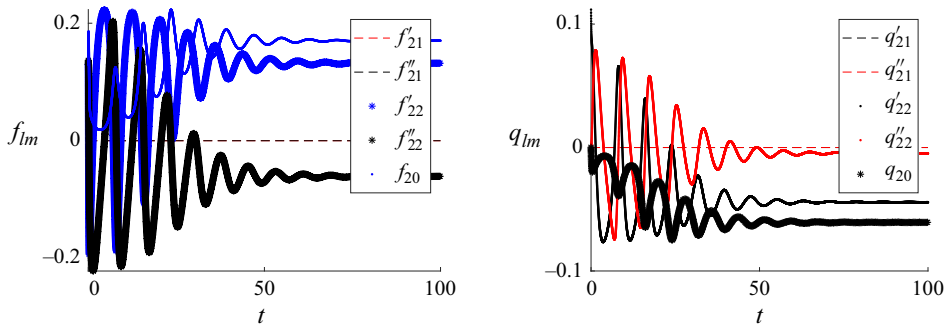


Figure 15. An example of tumbling motion dampening to give rise to tank-treading. The parameters are $\lambda = 2$, $\beta = 0.25$, $Cn = 0.5$, $Pe = 5$, $\Delta = 0.1$, $\chi = 0.01$, $a = -0.1$, $b = 1$, $q_0 = 0$, $\alpha = 1$.

5.2. Out-of-plane dynamics

In this section, we inspect the effects of exciting out-of-plane, higher-order modes not constrained to the $l = 2$ space. The initial condition is in figure 3(c). In the following simulations, unless specified, we used a cutoff mode $l_{max} = 8$ when solving the shape and order parameter. The following behaviours were observed.

5.2.1. Tank-treading (stationary)

In some simulations, we see the vesicle is frozen with a steady shape and concentration profile. This typically occurs when the interface width between phases is diffuse ($Cn \sim 1$), where the vesicle behaves similarly to a single component vesicle with matched viscosity

$\lambda = 2$ – stationary tank-treading motion. The steady inclination in-plane angle ϕ_i depends on the excess area and has been discussed previously (Vlahovska & Gracia 2007). The out-of-plane inclination angle θ_i is $\pi/2$, which indicates that the vesicle remains in the flow-shear (gradient) plane. We also note that the initial condition that primarily consisted of one yellow (stiff) lobe develops two lobes and stays stationary (indicated by q'_{22}, q''_{22}). Essentially, this motion is very similar to the in-plane counterpart discussed in § 5.1. See Supplementary movie [Video15.mp4](#) for more details.

5.2.2. Tumbling

When the capillary number χ is low, the vesicle is stiffer and tumbles under the influence of the background shear flow. The vesicle preserves its one-yellow-lobe configuration. The f_{2m} modes exhibit oscillatory motion. This motion is similar to the high Pe limit tumbling (see [figure 10](#)). See Supplementary movies [Video16.mp4](#) and [Video17.mp4](#) for a visualization.

5.2.3. Swinging/phase-treading

When the capillary number χ is higher, the vesicle undergoes swinging motion where the concentration rotates along the membrane and the shape exhibits weak oscillations about a fixed orientation. Similar to the high Pe limit, the shape oscillations are weakly out of plane. However, unlike the high Pe limit where the topology of the concentration profile is set by the initial condition, here the topology of the profile can change due to coarsening effects. For example, for the initial condition discussed in [figure 3\(c\)](#) where the vesicle has one single stiff (L_o) domain, coarsening during swinging gives rise to two stiff phases treading along the surface ([figures 16](#) and [17](#)). However, this effect is highly dependent on flow strength, bending stiffness and line tension. If for example we keep the same conditions as [figure 16](#), except we increase the capillary number (increase χ) and increase line tension (decrease α), one can see a situation where the one lobe initial condition persists and gives rise to $l = 1$ mode swinging in the subsequent results ([figures 18](#) and [19](#)). In this motion, the q_{11}, q_{1-1} modes persist while all other higher modes q_{lm} decay. The vesicle behaves like a Janus particle with divided phases, similar to what has been noted in previous studies (Gera & Salac 2018a).

5.2.4. Phase diagrams

This section provides phase diagrams to quantify general trends for vesicle dynamics. Since there is a large region of phase space to explore, we will isolate the effect of a few variables mentioned in § 2.3.

The first set of variables we will explore are the capillary number (χ) and line tension parameter (α). We will choose a moderately deflated vesicle ($\Delta = 0.1$) with matched interior and exterior fluid viscosity ($\lambda = 2$) and zero average order parameter ($q_0 = 0$), and choose $O(1)$ values for the other parameters: $Pe = 5$, $\beta = 0.5$. [Figure 20](#) shows the types of motion observed by the vesicle for different values of capillary number χ and line tension parameter α , when the initial condition is the same as that of [figure 3\(c\)](#). Two different interface widths are examined: $Cn = 0.2$ (more sharp) and $Cn = 0.55$ (less sharp). The regimes are reported over time window $t \approx 50$.

We will now explain the reasons behind these phase diagrams. In the first case where we see the relatively sharper interface $Cn = 0.2$ ([figure 20a](#)), we see that at small capillary numbers (χ), the vesicle exhibits tumbling motion. The rationale for this is that a low value of χ increases f_{lm}/χ in the shape equations (3.2)–(3.3). This effect causes the vesicle to elongate in the softer phase direction, after which it is rotated by the vorticity of the

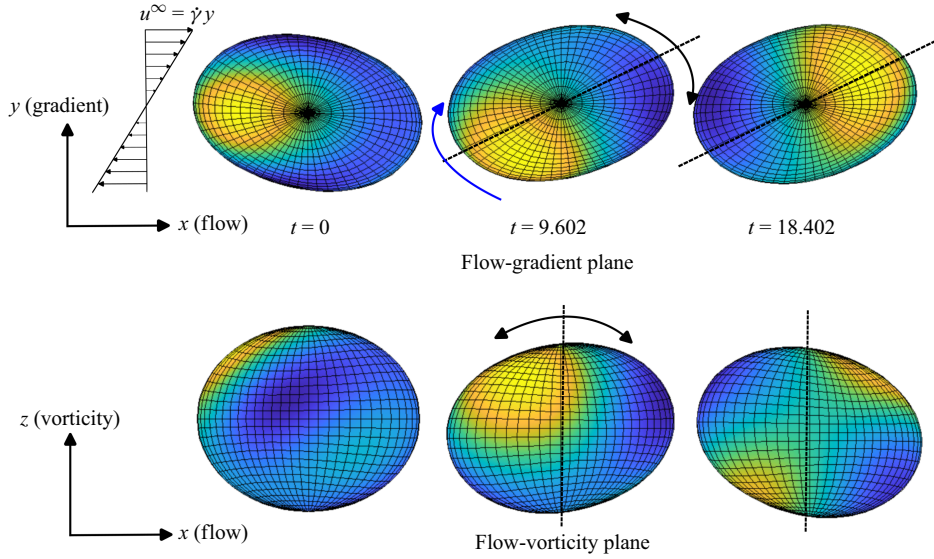


Figure 16. Phase-treading/swinging snapshots when higher-order q_{lm} modes are excited at $Pe \sim O(1)$. The initial condition is figure 3(c), and parameters are $\beta = 0.5$, $\alpha = 3$, $a = -0.1$, $b = 1$, $Pe = 5$, $\Delta = 0.1$, $\chi = 0.3$, $Cn = 0.2$, $\lambda = 2$, $q_0 = 0$. The blue arrow represents the lipid motion whereas the double-headed black arrow represents the vesicle shape oscillations about a fixed angle (black dashed line). The concentration profile coarsens from one lobe to two lobes. See Supplementary movies (Video18.mp4, Video19.mp4) for visualization and colourbar information.

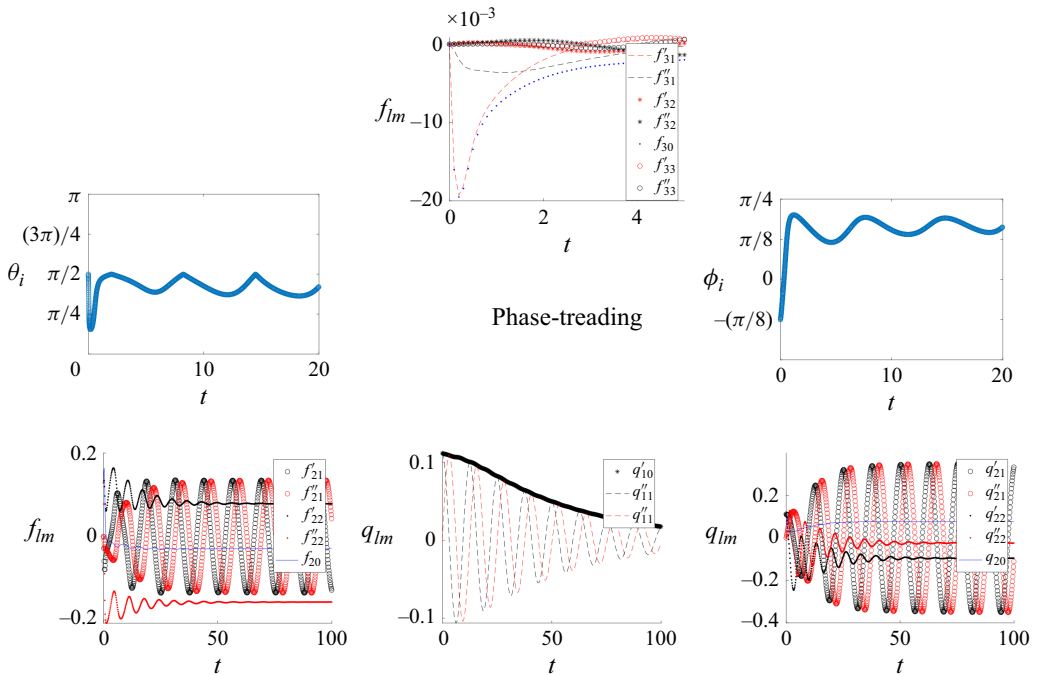


Figure 17. Modes and inclination angles for figure 16 (phase-treading at $Pe \sim O(1)$ with topology change).

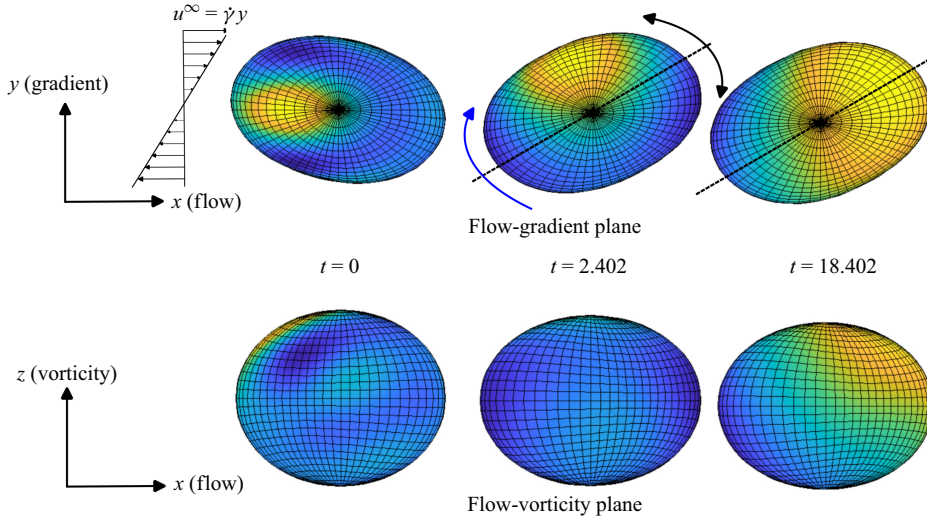


Figure 18. Phase-treading/swinging snapshots at $Pe \sim O(1)$ when higher-order q_{lm} modes are excited with $l=1$ modes driving the lipid motion. The initial condition is figure 3(c), and the parameters are $\beta = 0.5$, $\alpha = 1$, $a = -0.1$, $b = 1$, $Pe = 5$, $\Delta = 0.1$, $\chi = 0.6$, $Cn = 0.2$, $\lambda = 2$ and $q_0 = 0$. The blue arrow represents the lipid motion whereas the double-headed black arrow represents oscillations about a fixed angle (black dashed line). See Supplementary movies (Video20.mp4, Video21.mp4) for visualization and colourbar information.

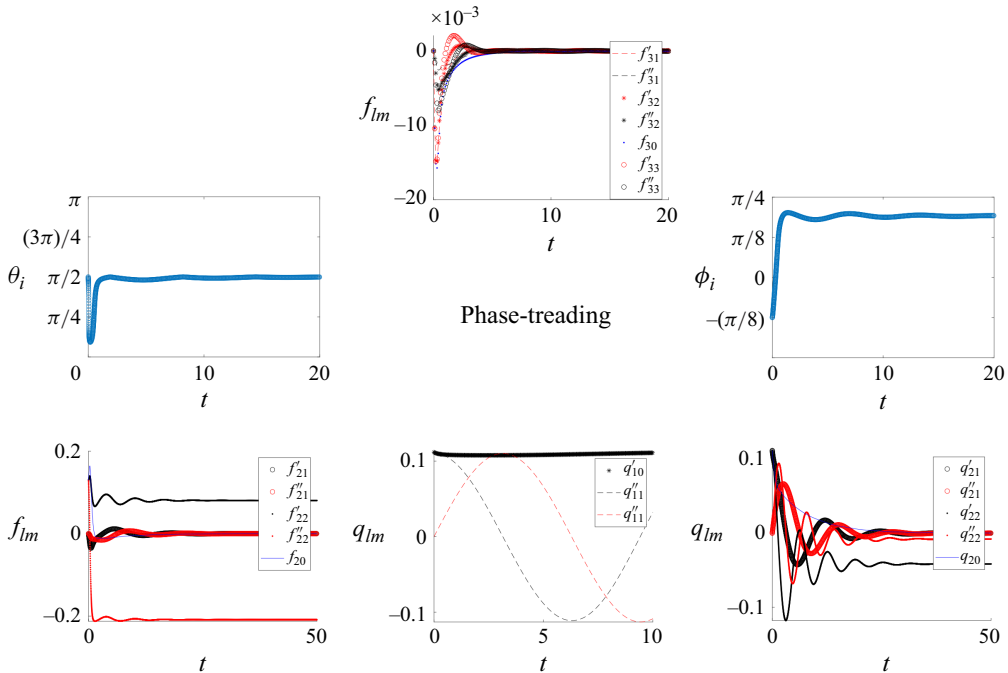


Figure 19. Modes and inclination angles for figure 18 (phase-treading at $Pe \sim O(1)$ without topology change).

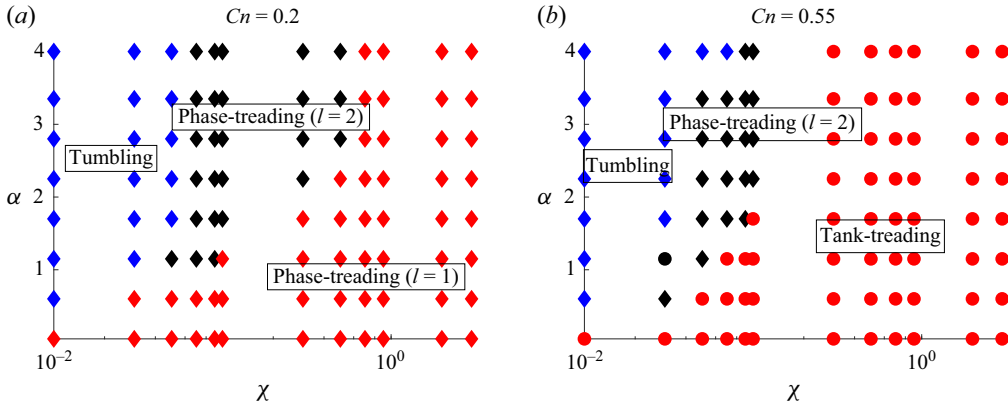


Figure 20. Phase diagrams for α versus χ at $\Delta = 0.1$, $Pe = 5$, $\beta = 0.5$, $\lambda = 2$, $a = -0.1$, $b = 1$, $q_0 = 0$. In these simulations, the initial condition is figure 3(c).

background flow, leading to tumbling motion, very similar to what is seen in the high Pe limit (see § 4). When we increase the χ value, depending on how stiff the vesicle is (given by α) we can get swinging/phase-treading motion. The $(im/2)q_{lm}$ term in the Cahn–Hilliard equation (3.4) drives the oscillatory behaviour of q_{lm} . This translates to the shape equation (3.2)–(3.3) showing oscillatory motion due to the q_{lm} term, but the χ^{-1} prefactor in (3.3) dampens these oscillations leading to swinging/phase-treading motion. We have further subdivided the phase-treading/swinging motion into $l = 1$ and $l = 2$ driven, akin to figures 16 and 18.

When the vesicle interface becomes more diffuse (Cn increases), we see that the vesicle shows behaviour more similar to single-component vesicles. This is seen in figure 20(b) where the $Cn = 0.55$. At small χ values, the term $\chi/\Delta^{1/2}$ is small thereby driving tumbling motion as stated before. As χ increases, the f_{lm} oscillations are dampened out in the shape equations (3.2)–(3.3) and the q_{lm} oscillations in the Cahn–Hilliard equation (3.4) are dampened due to the line tension penalty term $Cn^2 l^2 (l + 1)^2 q_{lm}$ coming into play. As this Cn increases further, the vesicle will largely show tank-treading behaviour, which is expected for single component vesicles with a matched viscosity ($\lambda = 2$).

6. Effect of asymmetric distributions

In the previous sections, we presented results based on $q_0 = 0$ which indicates an effective 50 : 50 phase distribution of the two separated phases. However, it is possible that we might encounter cases where $q_0 \neq 0$. In such cases, our theory can help predict the dynamics in limits where the q_0 value is not large enough to lie outside the phase-split region. On inspecting the shape equations (B10), we see that the shape-concentration coupling term D_l could make a drift caused by q_0 . One can observe from (B10) that for $l = 1$ modes, the shape coefficients take the following form:

$$\frac{df_{1m}}{dt} = \frac{imf_{1m}}{2} - \chi^{-1} \left[\frac{4\mu_0}{9\alpha Cn^2} \right] q_{1m}, \quad (6.1)$$

where $\mu_0 = aq_0 + bq_0^3$. This indicates that any non-zero initial condition in q_{1m} would induce a drift in f_{1m} , which would lead to translational motion – i.e. centre of mass tracking.

This phenomenon can be observed in figure 21. When one performs simulations at the same initial conditions but examines two different average concentrations $q_0 = 0.01$

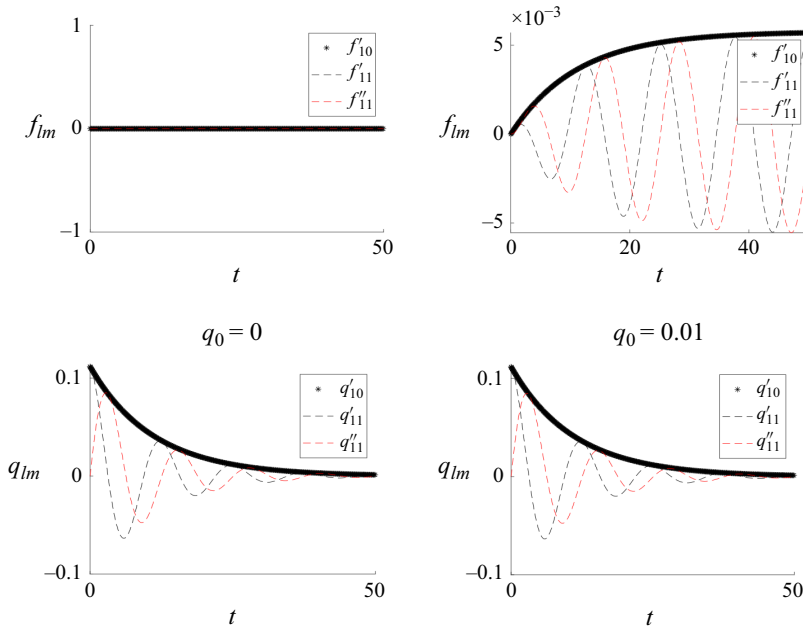


Figure 21. Lateral drift caused by asymmetric phase split ($q \neq 0$). In both cases, the initial condition is $f'_{22}(0) = f''_{22}(0) = f_{20}(0) = \sqrt{\Delta/10}$, and $q'_{10}(0) = q'_{11}(0) = q'_{22} = \sqrt{\Delta/8}$. The parameters for the simulations are $Pe = 5$, $\lambda = 2$, $a = -0.1$, $b = 1$, $\Delta = 0.1$, $\beta = 0.5$, $\chi = 0.6$, $\alpha = 1$, $Cn = 0.4$.

and $q_0 = 0$, one observes that a non-zero q_0 generates an oscillatory drift in the f'_{11} , f''_{11} modes and a non-zero drift in f_{10} . This indicates that the centre of mass of the vesicle performs sinusoidal oscillations about a fixed point in space. This lateral drift has been observed in previous studies as well (Gera & Salac 2018b). Lastly, we note that although the mechanisms are different, a similar oscillatory translational motion has been observed in single component vesicles in quadratic flows with confinement, and has been coined ‘snaking’ (Lyu *et al.* 2023).

7. Conclusion

In this study, we explored the effect of shear flow around a multicomponent vesicle containing a ternary mixture of phospholipids and cholesterol molecules within the bilayer. We analysed the small-deformation, weakly segregated limit and found that multicomponent vesicles exhibit very different dynamics compared with their single component counterparts. Results were examined in two distinct limits: one where the coarsening dynamics on the surface are unimportant ($Pe \gg 1$), and another where the coarsening time scale is comparable to the flow time scale ($Pe \sim O(1)$).

When $Pe \gg 1$, the phases are advected by the shear flow over the surface. The strength of the coupling with the shape deformations, given by $S = \beta q^+ / \chi$, determines the nature of this motion – swinging or tumbling. The initial conditions heavily dictate the dynamics in this system. In cases where purely the $l = 2$ in-plane modes are excited, the results show qualitative agreement with previous 2-D studies on such vesicles at the $Pe \rightarrow \infty$ limit (Gera *et al.* 2022). However, when out-of-plane modes are excited, it is possible to observe weakly out-of-plane tumbling and swinging motion. The concentration profile will perform periodic dynamics and retain its initial topology over time. Thus, if it starts asymmetric with respect to the flow-gradient plane, it will remain asymmetric over time.

In the intermediate Pe limit ($Pe \sim O(1) - O(10)$), we observe variety of motions based on the initial conditions and physical parameters such as tumbling, breathing, swinging/phase-treading and tank-treading. The biggest difference between $Pe \sim O(1)$ and $Pe \gg 1$ dynamics is that coarsening can occur on the membrane, which can give rise to stationary shapes and concentration profiles like tank-treading (which is not admissible in the $Pe \rightarrow \infty$ limit), and can alter the topology (i.e. number of lobes) in the concentration profile during swinging or tank-treading. We provided a general overview of the dynamics and illustrated phase diagrams for the different dynamical regimes. Generally, tumbling is favoured at small capillary numbers. At higher capillary numbers, phase-treading or tank-treading can occur, with the latter favoured at lower line tensions (e.g. more diffuse interface between phases). Lastly, we discussed the role of asymmetric phase split on the dynamics ($q \neq 0$), which can give rise to effects like oscillatory translational motion.

While the weak-segregation, small-deformation limit eases up the mathematical rigour largely due to the exclusion of nonlinear stresses, we believe it provides a great starting point for further semianalytical treatments of these systems using higher-order theories.

Supplementary movies. Supplementary movies are available at <https://doi.org/10.1017/jfm.2025.10632>.

Funding. The authors would like to acknowledge support from National Science Foundation (grant 2147559-CBET).

Declaration of interests The authors report no conflicts of interest.

Appendix A. Spherical harmonics

We define spherical harmonics Y_{lm} using the following convention (Jones 1985):

$$Y_{lm}(\theta, \phi) = \sqrt{\frac{(2l+1)(l-m)!}{4\pi(l+m)!}} (-1)^m P_l^m(\cos \theta) e^{im\phi}, \quad (\text{A1})$$

where (θ, ϕ) are the polar and azimuthal angles, and P_l^m are associated Legendre polynomials. The inner products of these spherical harmonics are orthonormal – i.e. $\int Y_{lm} Y_{l'm'}^* d\Omega = \Delta_{ll'} \Delta_{mm'}$, where the integral is over the unit sphere and $*$ represents complex conjugate. Furthermore, the following relationship holds: $Y_{l-m} = (-1)^m Y_{lm}^*$.

We can also define vector spherical harmonics as follows:

$$\mathbf{y}_{lm0} = [l(l+1)]^{-1/2} r \nabla Y_{lm}, \quad \mathbf{y}_{lm1} = i \hat{\mathbf{r}} \times \mathbf{y}_{lm0}, \quad \mathbf{y}_{lm2} = \hat{\mathbf{r}} Y_{lm}. \quad (\text{A2})$$

These vector spherical harmonics follow the properties of orthogonality by dot product and inner product.

Appendix B. Algebraic details for Stokes flow solution

B.1. Lamb's solution

Using vector spherical harmonics in (A2), one can define the velocity basis sets $\mathbf{u}^{\pm}_{lmq}$ ($q = 0, 1, 2$) where $\pm$ represents growing or decaying harmonics, respectively,

$$\mathbf{u}^{-}_{lm0} = \frac{1}{2} r^{-l} (2-l+lr^{-2}) \mathbf{y}_{lm0} + \frac{1}{2} r^{-l} [l(l+1)]^{1/2} (1-r^{-2}) \mathbf{y}_{lm2}, \quad (\text{B1a})$$

$$\mathbf{u}^{-}_{lm1} = r^{-l-1} \mathbf{y}_{lm1}, \quad (\text{B1b})$$

$$\mathbf{u}^{-}_{lm2} = \frac{1}{2} r^{-l} (2-l) \left(\frac{l}{l+1} \right)^{1/2} (1-r^{-2}) \mathbf{y}_{lm0} + \frac{1}{2} r^{-l} (l+(2-l)r^{-2}) \mathbf{y}_{lm2}, \quad (\text{B1c})$$

$$\mathbf{u}^+_{lm0} = \frac{1}{2}r^{l-1}(-l+1) + (l+3)r^2 \mathbf{y}_{lm0} - \frac{1}{2}r^{l-1}[l(l+1)]^{1/2}(1-r^2)\mathbf{y}_{lm2}, \quad (\text{B2a})$$

$$\mathbf{u}^+_{lm1} = r^l \mathbf{y}_{lm1}, \quad (\text{B2b})$$

$$\mathbf{u}^+_{lm2} = \frac{1}{2}r^{l-1}(3+l) \left(\frac{l+1}{l} \right)^{1/2} (1-r^2)\mathbf{y}_{lm0} + \frac{1}{2}r^{l-1}((l+3) - (l+1)r^2)\mathbf{y}_{lm2}. \quad (\text{B2c})$$

In terms of these basis sets, the velocity fields inside and outside the vesicle are

$$\mathbf{u}^{out} = \sum_{lmq} c_{lmq}^{\infty} \mathbf{u}^+_{lmq}(\mathbf{x}) + \sum_{lmq} (c_{lmq}^- - c_{lmq}^{\infty}) \mathbf{u}^-_{lmq}(\mathbf{x}), \quad (\text{B3a})$$

$$\mathbf{u}^{in} = \sum_{lmq} c_{lmq}^+ \mathbf{u}^+_{lmq}(\mathbf{x}). \quad (\text{B3b})$$

In the above equation, c_{lmq}^{∞} are the coefficients associated with the far field $\mathbf{u}^{\infty} = \mathbf{y} \hat{\mathbf{x}}$, which correspond to $c_{2\pm 20}^{\infty} = \mp i\sqrt{\pi/5}$, $c_{2\pm 22}^{\infty} = \mp i\sqrt{2\pi/15}$, $c_{101}^{\infty} = i\sqrt{2\pi/3}$. The coefficients c_{lmq}^- and c_{lmq}^+ are associated with the disturbance fields outside and inside vesicle, which one must solve.

From the velocity fields in (B3), one can compute the hydrodynamic stresses on the unit sphere $r = 1$. These are written as follows:

$$\mathbf{n} \cdot \mathbf{T}^{in} = (\lambda - 1) \sum_{lmq} \tau_{lmq}^{in} \mathbf{y}_{lmq}, \quad \mathbf{n} \cdot \mathbf{T}^{out} = \sum_{lmq} \tau_{lmq}^{out} \mathbf{y}_{lmq}. \quad (\text{B4})$$

Using the notation and results from Bławdziewicz *et al.* (2000), the coefficients τ_{lmq} are related to the velocity coefficients (c_{lmq}^{∞} , c_{lmq}^+ , c_{lmq}^-) as follows (details for $\Theta_{q'q}$ are given in Vlahovska & Gracia 2007):

$$\tau_{lmq}^{out} = \sum_{q'=0}^2 c_{lmq'}^{\infty} (\Theta_{q'q}^+ - \Theta_{q'q}^-) + \sum_{q'=0}^2 c_{lmq'}^- \Theta_{q'q}^-, \quad \tau_{lmq}^{in} = \sum_{q'=0}^2 c_{lmq'}^+ \Theta_{q'q}^+. \quad (\text{B5})$$

B.2. Solving equations

Solving the Stokes equations reduces to solving the unknown coefficients (c_{lmq}^+ , c_{lmq}^-) associated with the disturbance velocity fields, as well as the surface tension σ_{lm} on the interface. For each mode (l, m), there are seven unknowns to solve:

$$(c_{lm0}^+, c_{lm1}^+, c_{lm2}^+, c_{lm0}^-, c_{lm1}^-, c_{lm2}^-, \sigma_{lm}). \quad (\text{B6})$$

These are found by applying continuity of velocity across the surface $\mathbf{u}^{out} = \mathbf{u}^{in}$ at $r = 1$ (three equations), membrane incompressibility $\nabla_s \cdot \mathbf{u}^{in} = 0$ at $r = 1$ (one equation), and traction balance $\chi \mathbf{f}^{hydro} = \mathbf{f}^{mem}$ at $r = 1$ (three equations), where $\mathbf{f}^{mem}$ is Taylor expanded to $O(\Delta^{1/2})$ on the unit sphere.

Applying the boundary conditions is straightforward except the traction balance. Below, we write the membrane traction,

$$\mathbf{f}^{mem} = t_{lm0}^{mem} \mathbf{y}_{lm0} + t_{lm2}^{mem} \mathbf{y}_{lm2} \quad (\text{B7})$$

where

$$t_{lm0}^{mem} = \left(\left[-2\beta - \frac{\mu_0}{\alpha Cn^2} \right] q_{lm} - \sigma_{lm} \right) \sqrt{l(l+1)}, \quad (B8a)$$

$$t_{lm2}^{mem} = 2\beta l(l+1)q_{lm} + \left(\frac{(l+2)(l-1)g_0}{\alpha Cn^2} + (1+\beta q_0)(l-1)(l+1)(l+2) + \sigma_0(l-1)(l+2) \right) f_{lm} + 2\sigma_{lm}. \quad (B8b)$$

In the above equations, the quantities

$$g_0 = \frac{1}{2}aq_0^2 + \frac{1}{4}bq_0^4; \quad \mu_0 = aq_0 + bq_0^3 \quad (B9)$$

represent the double well potential and chemical potential evaluated at the base state q_0 . The dimensionless quantities α , Cn , β are defined in [table 2](#), which correspond to the line tension parameter, Cahn number and dimensionless bending stiffness difference.

In the dynamical equation for f_{lm} in the main text (3.2), one needs an expression for c_{lm2}^+ . After considerable algebra, one obtains expression (3.3), with coefficients C_{lm} , A_l , B_l and D_l given as follows:

$$C_{lm} = \frac{[c_{lm0}^\infty \sqrt{l(l+1)}(2l+1) + c_{lm2}^\infty(4l^3 + 6l^2 - 4l - 3)]}{d(\lambda, l)}, \quad (B10a)$$

$$A_l = -\frac{(l-1)l(l+1)(l+2)}{d(\lambda, l)} \left[(1+\beta q_0)l(l+1) + \frac{g_0}{\alpha Cn^2} \right], \quad (B10b)$$

$$B_l = -\frac{(l-1)l(l+1)(l+2)}{d(\lambda, l)}, \quad (B10c)$$

$$D_l = -\frac{2(l-1)l(l+1)(l+2)\beta}{d(\lambda, l)} - \frac{2l(l+1)\mu_0}{\alpha Cn^2 d(\lambda, l)}, \quad (B10d)$$

where

$$d(\lambda, l) = \frac{9}{\lambda} + (2l^3 + 3l^2 - 5). \quad (B11)$$

Appendix C. Numerical scheme to solve ODEs

We start with the following ODEs for the deformation and order parameter dynamics (for more details of the equations, see Venkatesh [2025](#)):

$$\frac{df_{lm}}{dt} = \frac{im}{2} f_{lm} + \lambda^{-1} C_{lm} + \lambda^{-1} \chi^{-1} [(A_l + B_l \sigma_0) f_{lm} + D_l q_{lm}], \quad (C1)$$

$$\frac{dq_{lm}}{dt} = \frac{im}{2} q_{lm} - \frac{1}{Pe} [\Lambda_{lm} l(l+1) + Cn^2 l^2 (l+1)^2 q_{lm} + 2\alpha Cn^2 \beta (l-1)l(l+1)(l+2) f_{lm}]. \quad (C2)$$

In the above equations, f_{lm} , q_{lm} and σ_0 are unknown. We start by splitting f_{lm} , q_{lm} into their real and imaginary parts – e.g. $f_{lm} = f'_{lm} + i f''_{lm}$ – while keeping in mind that $f_{l-m} = (-1)^{|m|} f_{l|m|}^*$. For the deformation, this gives us equations of the form ($m \geq 0$),

$$\frac{df'_{lm}}{dt} = g(f'_{lm}, f''_{lm}, q'_{lm}, q''_{lm}, \lambda, l, \chi) + \sigma_0 \mathcal{M}(f'_{lm}, \lambda, l, \chi), \quad (C3a)$$

$$\frac{df''_{lm}}{dt} = h(f'_{lm}, f''_{lm}, q'_{lm}, q''_{lm}, \lambda, l, \chi) + \sigma_0 \mathcal{P}(f'_{lm}, \lambda, l, \chi), \quad (C3b)$$

where g , h , $\mathcal{M}$ and $\mathcal{P}$ are functions of shape and deformation.

At time $t = t_i$, we perform a first-order Euler predictor–corrector scheme to march the vesicle shape (C3a)–(C3b). We first take a predictor step while neglecting the σ_0 terms

$$\frac{\tilde{f}'_{lm} - f'^i_{lm}}{\Delta t} = g(f'^i_{lm}, f''^i_{lm}, q^i_{lm}, q''^i_{lm}, \lambda, l, \chi), \quad (C4a)$$

$$\frac{\tilde{f}''_{lm} - f''^i_{lm}}{\Delta t} = h(f'^i_{lm}, f''^i_{lm}, q^i_{lm}, q''^i_{lm}, \lambda, l, \chi), \quad (C4b)$$

where $\tilde{f}'_{lm}$, $\tilde{f}''_{lm}$ are predictor values. We now proceed to take a corrector step to time $t = t_{i+1}$, now considering only the σ_0 portion of (C3a)–(C3b),

$$\frac{f'^{i+1}_{lm} - \tilde{f}'_{lm}}{\Delta t} = \sigma_0 \mathcal{M}(\tilde{f}'_{lm}, \lambda, l, \chi), \quad (C5a)$$

$$\frac{f''^{i+1}_{lm} - \tilde{f}''_{lm}}{\Delta t} = \sigma_0 \mathcal{P}(\tilde{f}''_{lm}, \lambda, l, \chi). \quad (C5b)$$

We choose σ_0 such that the shape at $t = t_{i+1}$ satisfies the area constraint $A = 4\pi + \Delta$, in other words

$$\sum_{l,m \geq 0} \frac{(l-1)(l+2)}{2} \left((f'^{i+1}_{lm})^2 + (f''^{i+1}_{lm})^2 \right) = \Delta. \quad (C6)$$

The latter step is done with root finding (using `fzero` in MATLAB with tolerance 10^{-8}). The reason for performing this step is to ensure that the nonlinear equations do not drift the differential–algebraic equation system away from the constraint (Ascher, Chin & Reich 1994). This method seems to perform better than simply imposing $\sum_{lm} (l-1)(l+2) df_{lm}/dt f_{lm}^* = 0$ to calculate σ_0 analytically.

After solving for shape, we perform time stepping for the Cahn–Hilliard equation in (C2). We perform the procedure from (Yoon *et al.* 2020). We treat the equation implicitly except the nonlinear term $aq + bq^3$, which we split into an explicit term $(a-2)q + bq^3$ and an implicit term $2q$. The procedure is shown below. After converting q_{lm} into real and imaginary components, we obtain

$$\begin{aligned} \frac{q'^{i+1}_{lm} - q'^i_{lm}}{\Delta t} &= \frac{-m}{2} q''^{i+1}_{lm} \\ &\quad - \frac{1}{Pe} \left[\text{Re} \left(\Lambda'^i_{lm} \right) l(l+1) + 2q'^{i+1}_{lm} l(l+1) + Cn^2 l^2 (l+1)^2 q'^{i+1}_{lm} \right. \\ &\quad \left. + 2\alpha Cn^2 \beta (l-1)l(l+1)(l+2) f'^{i+1}_{lm} \right], \end{aligned} \quad (C7a)$$

$$\begin{aligned} \frac{q''^{i+1}_{lm} - q''^i_{lm}}{\Delta t} &= \frac{m}{2} q'^{i+1}_{lm} \\ &\quad - \frac{1}{Pe} \left[\text{Im} \left(\Lambda'^i_{lm} \right) l(l+1) + 2q''^{i+1}_{lm} l(l+1) + Cn^2 l^2 (l+1)^2 q''^{i+1}_{lm} \right. \\ &\quad \left. + 2\alpha Cn^2 \beta (l-1)l(l+1)(l+2) f''^{i+1}_{lm} \right], \end{aligned} \quad (C7b)$$

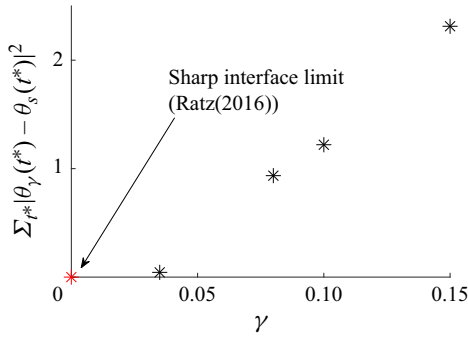


Figure 22. Sum of squared errors compared with analytical solution of surface Cahn–Hilliard equation Rätz (2016). The values of γ are $\gamma = 0.15, 0.1, 0.08, 0.035$. In the simulations, $D = 1$. The sum is calculated over all points in time. The time is simulated until $t = 0.1$.

where $\Lambda'_{lm} = \int_{\Omega} ((a - 2)q + bq^3) Y_{lm} d\Omega$, and $\text{Re}(\cdot)$ and $\text{Im}(\cdot)$ represent real and imaginary parts of an expression.

Appendix D. Convergence of the spectral method in the absence of flow

For the Cahn–Hilliard equation without flow, we benchmarked our simulation against a sharp-interface limit theory developed by Rätz 2016, where the authors developed an axisymmetric solution to Cahn–Hilliard dynamics on a sphere. The authors solve the following equation:

$$\frac{\partial q}{\partial t} = D \nabla_s^2 (aq + bq^3 - \gamma^2 \nabla_s^2 q) \quad (\text{D1})$$

where $t = \gamma t^*$, with t^* being a modified time. They solve this equation for an initial condition being a striped domain where $q = 1$ from $\theta = 0$ to $\theta = \theta_1 = 0.43$, $q = -1$ from $\theta = \theta_1 = 0.43$ to $\theta = \theta_2 = 0.92$, and $q = 1$ from $\theta = \theta_2 = 0.92$ to $\theta = \pi$. In the sharp interface limit as $\gamma \rightarrow 0$, the three domains eventually merge into two domains, with the steady-state solution being $\theta_1 = 0$ and $\theta_2 = 0.812$ for $a = 1$, $b = -1$ and $D = 1$.

For different values of γ , we plot θ_1 versus t^* , where θ_1 represents the first point at which the order parameter changes from a positive to a negative value. We then calculate the sum of squared errors between the numerical solution θ_γ and the semianalytical solution θ_s (valid at $\gamma = 0$) for θ_1 . The semianalytical solution is given by the following differential equation for θ_1 :

$$\frac{d\theta_i}{dt^*} = -\frac{c^+}{\sin \theta_i} \quad (\text{D2})$$

where $i = 1, 2$ and $c^+ = -(\sqrt{2}/3)((\cot \theta_1 + \cot \theta_2)/(\log |\tan(\theta_1/2)|) + (\log |\tan(\theta_2/2)|))$. Figure 22 shows the error for different values of γ and shows that the numerical solution converges to the analytical solution in the sharp interface limit $\gamma \rightarrow 0$. Note that we mainly deal with the diffuse interface limit in our study and do not need the γ values to be extremely small.

Appendix E. Algebra, $Pe \rightarrow \infty$ theory ($l = 2$ modes excited in-plane)

We examine (3.2)–(3.3) and use the expressions $f_{22} = 2\epsilon_1 \sqrt{2\pi/15}(a_2 - ib_2)$ and $q_{22} = 2\epsilon_2 \sqrt{2\pi/15} \exp(it)$. If we define a modified capillary number as $C = \chi/\epsilon_1$ and modified

time $\tau = t/\epsilon_1$, the system of equations at leading order for $C \sim O(1)$, $\tau \sim O(1)$, and $\epsilon_2/\epsilon_1 \sim O(1)$ are

$$\begin{aligned}\frac{da_2}{d\tau} &= \lambda^{-1} C^{-1} \left[(A_2 + B_2 \sigma_0) a_2 + \frac{\epsilon_2}{\epsilon_1} D_2 \cos(\epsilon_1 \tau) \right], \\ \frac{db_2}{d\tau} &= \lambda^{-1} C^{-1} \left[(A_2 + B_2 \sigma_0) b_2 - \frac{\epsilon_2}{\epsilon_1} D_2 \sin(\epsilon_1 \tau) \right] - \lambda^{-1} \frac{1}{2i} \sqrt{\frac{15}{2\pi}} C_{22}. \quad (\text{E1})\end{aligned}$$

In the above equation, A_2 , B_2 , C_{22} and D_2 are coefficients defined in (B10)–(B11) for $l = 2, m = 2$. The surface tension σ_0 is a Lagrange multiplier to enforce the surface area constraint – i.e. $a_2^2 + b_2^2 = 15/32\pi$. We solve for σ_0 using the expression $a_2(da_2/d\tau) + b_2(db_2/d\tau) = 0$, and plug it back into the above ODE (E1) to yield

$$\begin{aligned}\frac{da_2}{d\tau} &= -\Theta b_2, \\ \frac{db_2}{d\tau} &= \Theta a_2, \quad (\text{E2})\end{aligned}$$

where Θ is

$$\Theta = -\frac{1}{a_2^2 + b_2^2} \left[\frac{1}{2i} \sqrt{\frac{15}{2\pi}} \lambda^{-1} C_{22} a_2 + \frac{\epsilon_2}{\epsilon_1} \lambda^{-1} C^{-1} D_2 (a_2 \sin(\epsilon_1 \tau) + b_2 \cos(\epsilon_1 \tau)) \right]. \quad (\text{E3})$$

Substituting the expressions for C_{22} and D_2 (i.e. $C_{22} = -60\lambda i \sqrt{2\pi/15} (9 + 23\lambda)^{-1}$, $D_2 = -48\lambda\beta (9 + 23\lambda)^{-1}$ for $q_0 = 0$), as well as the constraint $a_2^2 + b_2^2 = 15/32\pi$ yields (4.4).

In Gera *et al.* 2022, the authors obtained the same form of the differential equation as (E2) for a_2 and b_2 for a 2-D vesicle, when the radius is written as $r = 1 + \epsilon a_2(t) \cos(2\phi) + \epsilon b_2 \sin(2\phi)$. However, the expression for Θ is different due the fact the vesicle is 2-D rather than 3-D. Their expression is (using the notation in our paper)

$$\Theta_{2D} = \frac{1}{\lambda} \frac{1}{Q^2} \left[a_2 + \frac{\epsilon_2}{\epsilon_1} \beta C^{-1} (a_2 \sin(\epsilon_1 \tau) + b_2 \cos(\epsilon_1 \tau)) \right] \quad (\text{E4})$$

where Q is a constant equal to $Q^2 = 1/3 \sum_{n=1}^{\infty} (n^2 - 1)(a_n^2 + b_n^2)$. If one defines a 2-D excess length parameter as $\Delta_{2D} = L/R - 2\pi$, where L is the membrane length and area $A = \pi R^2$, one obtains

$$\Delta_{2D} = \frac{3\pi}{2} \epsilon_1^2 Q^2. \quad (\text{E5})$$

Keeping consistent with the notation used in our paper, if we define the small parameter $\epsilon = \Delta_{2D}^{1/2}$, this yields $Q^2 = 2/3\pi$, which gives the final expression written in the main text (4.5).

Appendix F. Comparison between theory and simulation, $Pe \rightarrow \infty$, in-plane deformations and concentrations

This section performs full numerical simulations to check the validity of $Pe \rightarrow \infty$ theory discussed in § 4.2. Figures 23 and 24 show cases where swinging and tumbling occur at $Pe = 1000$. The circles indicate the theory (4.3) for the q_{lm} and f_{lm} modes, while the dashed lines are the full numerical simulations (solving (3.2) and (3.4)). In both simulations, we start with an initial condition $f'_{22}(0) = \sqrt{\Delta/10}$, $f''_{22}(0) = -\sqrt{\Delta/10}$, $q'_{22}(0) = 2q^+ \sqrt{2\pi/15}$. We observe that simulations closely match the theory,

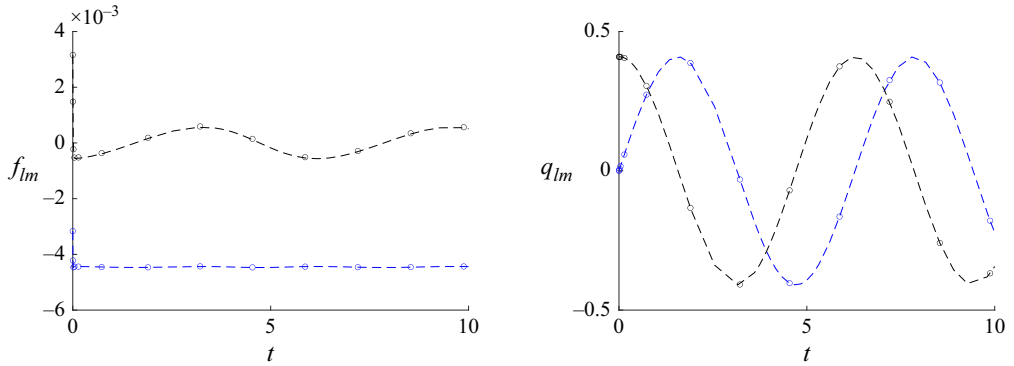


Figure 23. High Pe swinging motion for $\alpha = 0.1$, $Cn = 0.1$, $\beta = 0.5$, $Pe = 1000$, $\chi = 2$, $q_0 = 0$, $a = -0.1$, $b = 1$, $\Delta = 0.0001$, $\lambda = 2$. The dashed lines represent the full numerical solution of (3.2), (3.4) whereas the circles represent f'_{22} , f''_{22} , q'_{22} , q''_{22} for the $Pe \rightarrow \infty$ theory equations (4.2), (4.3). The blue circles/dashed curves represent the f''_{22} , q''_{22} modes. The black circles/dashed curves represent the f'_{22} , q'_{22} modes.

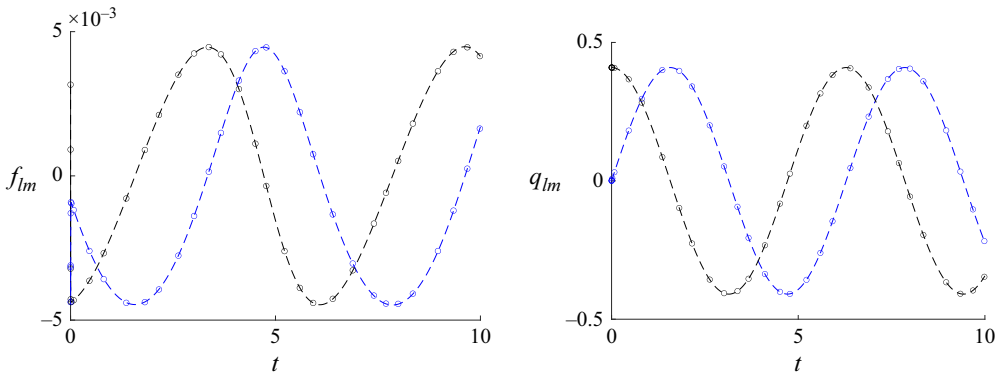


Figure 24. High Pe tumbling motion for $\alpha = 0.1$, $Cn = 0.1$, $\beta = 0.5$, $Pe = 1000$, $\chi = 0.05$, $q_0 = 0$, $a = -0.1$, $b = 1$, $\Delta = 0.0001$, $\lambda = 2$. We use $l_{max} = 8$ in these simulations. The dashed lines represent the full numerical solution of (3.2), (3.4) whereas the circles represent f'_{22} , f''_{22} , q'_{22} , q''_{22} for the $Pe \rightarrow \infty$ theory equations (4.2), (4.3). The blue circles/dashed curves represent the f''_{22} , q''_{22} modes. The black circles/dashed curves represent the f'_{22} , q'_{22} modes.

and this behaviour generally holds for $Pe \sim O(100)$ and above. Another interesting observation is the validity of the expressions for the $Pe \rightarrow \infty$ (4.3). These expressions hold only for $\Delta \ll 1$. As Δ increases, the rotational term in (3.2) becomes significant and starts influencing the dynamics as well. Hence, the approximation given by expressions (4.3) are not accurate as Δ increases. Lastly, we generally observe in our simulations that the out-of-plane f_{20} deformations rapidly decay to zero, and thus this mode does not seem to affect the theory.

REFERENCES

- ABKARIAN, M., LARTIGUE, C. & VIALLAT, A. 2002 Tank treading and unbinding of deformable vesicles in shear flow: determination of the lift force. *Phys. Rev. Lett.* **88** (6), 068103.
ALBERTS, B. 2017 *Molecular Biology of the Cell*. Garland Science.

- AMAZON, J.J., GOH, S.L. & FEIGENSON, G.W. 2013 Competition between line tension and curvature stabilizes modulated phase patterns on the surface of giant unilamellar vesicles: a simulation study. *Phys. Rev. E* **87**, 022708.
- ARNOLD, D.P., GUBBALA, A. & TAKATORI, S.C. 2023 Active surface flows accelerate the coarsening of lipid membrane domains. *Phys. Rev. Lett.* **131** (12), 128402.
- ASCHER, U.M., CHIN, H. & REICH, S. 1994 Stabilization of DAEs and invariant manifolds. *Numer. Math.* **67**, 131–149.
- BAUMGART, T., HESS, S.T. & WEBB, W.W. 2003 Imaging coexisting fluid domains in biomembrane models coupling curvature and line tension. *Nature* **425** (6960), 821–824.
- BIBEN, T. & MISBAH, C. 2003 Tumbling of vesicles under shear flow within an advected-field approach. *Phys. Rev. E* **67** (3), 031908.
- BŁAWDZIEWICZ, J., VLAHOVSKA, P. & LOEWENBERG, M. 2000 Rheology of a dilute emulsion of surfactant-covered spherical drops. *Physica A: Stat. Mech. Appl.* **276**, 50–85.
- CAMLEY, B.A. & BROWN, F.L. 2014 Fluctuating hydrodynamics of multicomponent membranes with embedded proteins. *J. Chem. Phys.* **141** (7), 075103.
- CAMPELO, F., ARNAEZ, C., MARRINK, S.J. & KOZLOV, M.M. 2014 Helfrich model of membrane bending: from Gibbs theory of liquid interfaces to membranes as thick anisotropic elastic layers. *Adv. Colloid Interface Sci.* **208**, 25–33.
- DAHL, J.B., NARSIMHAN, V., GOUVEIA, B., KUMAR, S., SHAQFEH, E.S.G. & MULLER, S.J. 2016 Experimental observation of the asymmetric instability of intermediate-reduced-volume vesicles in extensional flow. *Soft Matt.* **12** (16), 3787–3796.
- DAVIS, J.H., CLAIR, J.J. & JUHASZ, J. 2009 Phase equilibria in DOPC/DPPC-d62/cholesterol mixtures. *Biophys. J.* **96** (2), 521–539.
- DESCHAMPS, J., KANTSLE, V., SEGRE, E. & STEINBERG, V. 2009 Dynamics of a vesicle in general flow. *Proc. Natl Acad. Sci. USA* **106** (28), 11444–11447.
- FREDRICKSON, G.H. & HELFAND, E. 1987 Fluctuation effects in the theory of microphase separation in block copolymers. *J. Chem. Phys.* **87** (1), 697–705.
- GEANKOPLIS, C. 2003 *Transport Processes and Separation Process Principles (Includes Unit Operations)*. Prentice Hall Press.
- GERA, P. 2017 *Hydrodynamics of multicomponent vesicles*. PhD thesis, State University of New York at Buffalo.
- GERA, P. & SALAC, D. 2018a Modeling of multicomponent three-dimensional vesicles. *Comput. Fluids* **172**, 362–383.
- GERA, P. & SALAC, D. 2018b Three-dimensional multicomponent vesicles: dynamics and influence of material properties. *Soft Matt.* **14** (37), 7690–7705.
- GERA, P., SALAC, D. & SPAGNOLIE, S.E. 2022 Swinging and tumbling of multicomponent vesicles in flow. *J. Fluid Mech.* **935**, A39.
- GOMPPER, G. & KLEIN, S. 1992 Ginzburg–Landau theory of aqueous surfactant solutions. *J. Phys. II (Paris)* **2** (9), 1725–1744.
- GUO, X. & SZOKA, F.C. 2003 Chemical approaches to triggerable lipid vesicles for drug and gene delivery. *Accounts Chem. Res.* **36** (5), 335–341.
- HELFRI, W. 1973 Elastic properties of lipid bilayers: theory and possible experiments. *Z. Naturforsch. C* **28** (11–12), 693–703.
- HERZENBERG, L., TUNG, J.W.M., MOORE, W., HERZENBERG, L.A. & PARKS, D. 2006 Interpreting flow cytometry data: a guide for the perplexed. *Nat. Immunol.* **7**, 681–685.
- JOHN, K., SCHREIBER, S., KUBELT, J., HERRMANN, A. & MÜLLER, P. 2002 Transbilayer movement of phospholipids at the main phase transition of lipid membranes: implications for rapid flip-flop in biological membranes. *Biophys. J.* **83** (6), 3315–3323.
- JONES, M.N. 1985 *Spherical Harmonics and Tensors for Classical Field Theory*. Research Studies Press; Wiley.
- KANTSLE, V. & STEINBERG, V. 2006 Transition to tumbling and two regimes of tumbling motion of a vesicle in shear flow. *Phys. Rev. Lett.* **96** (3), 036001.
- KELLER, S.R. & SKALAK, R. 1982 Motion of a tank-treading ellipsoidal particle in a shear flow. *J. Fluid Mech.* **120**, 27–47.
- KOLLMANNBERGER, P. & FABRY, B. 2011 Linear and nonlinear rheology of living cells. *Annu. Rev. Mater. Res.* **41**, 75–97.
- KUMAR, D., RICHTER, C.M. & SCHROEDER, C.M. 2020 Conformational dynamics and phase behavior of lipid vesicles in a precisely controlled extensional flow. *Soft Matt.* **16** (2), 337–347.

- KUMAR, P.B.S., GOMPPER, G. & LIPOWSKY, R. 1999 Modulated phases in multicomponent fluid membranes. *Phys. Rev. E* **60** (4), 4610.
- LEI, K. *et al.* 2021 Cancer-cell stiffening via cholesterol depletion enhances adoptive T-cell immunotherapy. *Nat. Biomed. Engng* **5** (12), 1411–1425.
- LEIBLER, L. 1980 Theory of microphase separation in block copolymers. *Macromolecules* **13** (6), 1602–1617.
- LIN, C., KUMAR, D., RICHTER, C.M., WANG, S., SCHROEDER, C.M. & NARSIMHAN, V. 2021 Vesicle dynamics in large amplitude oscillatory extensional flow. *J. Fluid Mech.* **929**, A43.
- LIN, C. & NARSIMHAN, V. 2019 Shape stability of deflated vesicles in general linear flows. *Phys. Rev. Fluids* **4**, 123606.
- LIPOWSKY, R. & SACKMANN, E. 1995 *Structure and Dynamics of Membranes—from Cells to Vesicles (handbook of Biological Physics)*, Vol. 1. Elsevier.
- LIU, K., MARPLE, G.R., ALLARD, J., LI, S., VEERAPANENI, S. & LOWENGRUB, J. 2017 Dynamics of a multicomponent vesicle in shear flow. *Soft Matt.* **13** (19), 3521–3531.
- LUO, Y. & MAIBAUM, L. 2020 Modulated and spiral surface patterns on deformable lipid vesicles. *J. Chem. Phys.* **153** (14), 144901.
- LYU, J., CHEN, P.G., FARUTIN, A., JAEGER, M., MISBAH, C. & LEONETTI, M. 2023 Swirling of vesicles: shapes and dynamics in Poiseuille flow as a model of RBC microcirculation. *Phys. Rev. Fluids* **8**, L021602.
- MICHEL, V. & BAKOVIC, M. 2007 Lipid rafts in health and disease. *Biol. Cell* **99** (3), 129–140.
- MISBAH, C. 2006 Vacillating breathing and tumbling of vesicles under shear flow. *Phys. Rev. Lett.* **96** (2), 028104.
- NAPOLI, G. & VERGORI, L. 2010 Equilibrium of nematic vesicles. *J. Phys. A: Math. Theor.* **43** (44), 445207.
- NARSIMHAN, V., SPANN, A.P. & SHAQFEH, E.S.G. 2014 The mechanism of shape instability for a vesicle in extensional flow. *J. Fluid Mech.* **750**, 144–190.
- NARSIMHAN, V., SPANN, A.P. & SHAQFEH, E.S.G. 2015 Pearling, wrinkling, and buckling of vesicles in elongational flows. *J. Fluid Mech.* **777**, 1–26.
- NEEDHAM, D. 1999 Materials engineering of lipid bilayers for drug carrier performance. *MRS Bull.* **24** (10), 32–41.
- NEGISHI, M., SETO, H., HASE, M. & YOSHIKAWA, K. 2008 How does the mobility of phospholipid molecules at a water/oil interface reflect the viscosity of the surrounding oil? *Langmuir* **24** (16), 8431–8434.
- NOGUCHI, H. & GOMPPER, G. 2005 Dynamics of fluid vesicles in shear flow: effect of membrane viscosity and thermal fluctuations. *Phys. Rev. E* **72** (1), 011901.
- NOGUCHI, H. & GOMPPER, G. 2007 Swinging and tumbling of fluid vesicles in shear flow. *Phys. Rev. Lett.* **98**, 128103.
- RAMANUJAN, S. & POZRIKIDIS, C. 1998 Deformation of liquid capsules enclosed by elastic membranes in simple shear flow: large deformations and the effect of fluid viscosities. *J. Fluid Mech.* **361**, 117–143.
- RIOUAL, F., BIBEN, T. & MISBAH, C. 2004 Analytical analysis of a vesicle tumbling under a shear flow. *Phys. Rev. E* **69** (6), 061914.
- RÄTZ, A. 2016 A benchmark for the surface Cahn–Hilliard equation. *Appl. Math. Lett.* **56**, 65–71.
- SAFRAN, S. 2018 *Statistical Thermodynamics of Surfaces, Interfaces, and Membranes*. CRC Press.
- SALMOND, N., KHANNA, K., OWEN, G.R. & WILLIAMS, K.C. 2021 Nanoscale flow cytometry for immunophenotyping and quantitating extracellular vesicles in blood plasma. *Nanoscale* **13**, 2012–2025.
- SEIFERT, U. 1997 Configurations of fluid membranes and vesicles. *Adv. Phys.* **46** (1), 13–137.
- SEIFERT, U. 1999 Fluid membranes in hydrodynamic flow fields: formalism and an application to fluctuating quasispherical vesicles in shear flow. *European Phys. J. B-Condens. Matt. Complex Syst.* **8**, 405–415.
- SEUL, M. & ANDELMAN, D. 1995 Domain shapes and patterns: the phenomenology of modulated phases. *Science* **267** (5197), 476–483.
- SHI, Z., GRABER, Z.T., BAUMGART, T., STONE, H.A. & COHEN, A.E. 2018 Cell membranes resist flow. *Cell* **175** (7), 1769–1779.e13.
- SIMONS, K. & IKONEN, E. 1997 Functional rafts in cell membranes. *Nature* **387** (6633), 569–572.
- SIMONS, K. & TOOMRE, D. 2000 Lipid rafts and signal transduction. *Nat. Rev. Mol. Cell Biol.* **1** (1), 31–39.
- SOHN, J.S., TSENG, Y.-H., LI, S., VOIGT, A. & LOWENGRUB, J.S. 2010 Dynamics of multicomponent vesicles in a viscous fluid. *J. Comput. Phys.* **229** (1), 119–144.
- TANIGUCHI, T., KAWASAKI, K., ANDELMAN, D. & KAWAKATSU, T. 1994 Phase transitions and shapes of two component membranes and vesicles II : weak segregation limit. *J. Phys. II (Paris)* **4** (8), 1333–1362.
- TANIGUCHI, T., YANAGISAWA, M. & IMAI, M. 2011 Numerical investigations of the dynamics of two-component vesicles. *J. Phys.: Condens. Matt.* **23** (28), 284103.
- UPPAMOOSHIKKAL, P., TRISTRAM-NAGLE, S. & NAGLE, J.F. 2010 Orientation of tie-lines in the phase diagram of DOPC/DPPE/cholesterol model biomembranes. *Langmuir* **26** (22), 17363–17368.

- VEATCH, S.L. & KELLER, S.L. 2005*a* Miscibility phase diagrams of giant vesicles containing sphingomyelin. *Phys. Rev. Lett.* **94** (14), 148101.
- VEATCH, S.L. & KELLER, S.L. 2005*b* Seeing spots: complex phase behavior in simple membranes. *Biochim. Biophys. Acta* **1746** (3), 172–185.
- VENKATESH, A. 2025 Dynamics, stability, and thermodynamics of deformable biological membranes interacting with Newtonian and viscoelastic fluids. PhD thesis, Purdue University Graduate School, Indiana, USA.
- VLAHOVSKA, P.M. & GRACIA, R.S. 2007 Dynamics of a viscous vesicle in linear flows. *Phys. Rev. E* **75**, 016313.
- VLAHOVSKA, P.M., LOEWENBERG, M. & BLAWZDZIEWICZ, J. 2005 Deformation of a surfactant-covered drop in a linear flow. *Phys. Fluids* **17** (10), 103103.
- YANAGISAWA, M., IMAI, M., MASUI, T., KOMURA, S. & OHTA, T. 2007 Growth dynamics of domains in ternary fluid vesicles. *Biophys. J.* **92** (1), 115–125.
- YOON, S., JEONG, D., LEE, C., KIM, H., KIM, S., LEE, H.G. & KIM, J. 2020 Fourier-spectral method for the phase-field equations. *Mathematics* **8** (8), 1385.
- ZHAO, H. & SHAQFEH, E.S.G. 2013 The shape stability of a lipid vesicle in a uniaxial extensional flow. *J. Fluid Mech.* **719**, 345–361.