

Reproducibility of energy intake, gastric emptying, blood glucose, plasma insulin and cholecystokinin responses in healthy young males

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Gastric emptying, as well as intragastric meal distribution, and gastrointestinal hormones, including cholecystokinin (CCK), play an important role in appetite regulation. The evaluation of gastrointestinal factors regulating food intake is commonly performed in healthy, lean, young male participants. It has, however, been suggested that there is a marked interindividual variability in the effects of nutrient ‘preloads’ on energy intake in this group. Whether there is significant intraindividual variation in acute energy intake after a nutrient preload, and, if so, how this relates to day-to-day differences in gastric emptying and gastrointestinal hormone release, is unclear. The purpose of the present paper is to evaluate the hypothesis that energy intake after a nutrient preload would be reproducible and associated with reproducible patterns of gastric emptying, intragastric distribution and gastrointestinal hormone release. Fifteen healthy men (age 25 (SEM 5) years) consumed a glucose preload (50 g glucose in 300 ml water; 815 kJ) on three occasions. Gastric emptying and intragastric meal distribution (using three-dimensional ultrasound), blood glucose, plasma insulin and CCK concentrations and appetite perceptions were evaluated over 90 min, and energy intake from a cold buffet-style meal was then quantified. Energy intake was highly reproducible within individuals between visits (intraclass correlation coefficient, $r_i = 0.9$). Gastric emptying, intragastric meal distribution, blood glucose, plasma insulin and CCK concentrations and appetite perceptions did not differ between visits ($r_i > 0.7$ for all). In healthy males, energy intake is highly reproducible, at least in the short term, and is associated with reproducible patterns of gastric emptying, glycaemia, insulinaemia and CCK release.

Day-to-day energy intake: Gastric emptying: Cholecystokinin: Three-dimensional ultrasound

Studies evaluating appetite and energy intake in a laboratory setting have frequently utilised ‘preload’ paradigms^(1–7), in which subjects receive a standardised amount of a test meal that can vary in macronutrient composition, physical state, volume and energy density, either orally or infused directly into the stomach or small intestine, and subsequent energy intake is assessed. Healthy, lean, young male subjects are frequently used in such studies, as they have been reported to have a greater capacity to adjust energy intake in response to a energy preload (i.e. by decreasing the amount of energy consumed at a subsequent meal), when compared with elderly men, healthy females or obese individuals^(8,9). Nevertheless, within this group, there appears to be substantial interindividual variability in both the total amount of food consumed and the magnitude of the reduction in their energy intake in response to energy manipulation⁽¹⁰⁾. For example, while a significant overall (mean) reduction in energy intake was evident in response to an intraduodenal lipid infusion, when compared with a control infusion of saline, the magnitude of the decrease

in energy intake in individual subjects was highly variable, and some subjects failed to compensate for the energy infused⁽¹⁰⁾.

Energy intake in a laboratory setting is commonly assessed using a standardised buffet-style meal, containing a range of food items, varying in macronutrient composition, and provided in excess of what subjects would be expected to consume^(1,11–13). The presentation of a meal in excess could potentially result in spontaneous overconsumption^(6,14), thereby confounding the results of the studies designed to detect subtle differences in energy intake in response to a treatment. To our knowledge, only two studies have hitherto addressed this issue^(13,15). Arvaniti *et al.*⁽¹³⁾ evaluated energy intake and macronutrient composition of food consumed from a standardised buffet-style meal, on two separate occasions, and reported that energy intake did not vary between the 2 d. Since such studies frequently consist of more than two study conditions^(12,16–18), we considered it important to evaluate intraindividual reproducibility of energy intake on three, rather than two, occasions.

Abbreviations: AUC, areas under the curve; CCK, cholecystokinin; 3D, three-dimensional; VAS, visual analogue scale.

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Gastrointestinal factors, including gastric distension (occurring as a result of slowing of gastric emptying)⁽¹⁹⁾, intragastric meal distribution⁽²⁰⁾ and the release of gastrointestinal hormones, including cholecystokinin (CCK)⁽²¹⁾, play an important role in the regulation of energy intake. It is presently unclear whether the consistency in energy intake, which has been described previously⁽¹³⁾, is associated with reproducible patterns of gastric emptying and gastrointestinal hormone release. The second aim of the present study was, therefore, to evaluate whether gastrointestinal changes in response to an orally ingested glucose 'preload' would also be reproducible when assessed repeatedly within an individual. We employed a glucose drink as the preload, as we have recently shown that its gastric emptying can be accurately assessed with a novel three-dimensional (3D) ultrasound technique⁽²²⁾. Finally, we reasoned that the determination of intraindividual variations in response to the same treatment would also allow us to calculate the minimum changes in energy intake and gastrointestinal function that would be required to detect statistically significant treatment effects, should a treatment be given, for which there is relatively little information; this formed the third aim of the present study.

Materials and methods

Subjects

Fifteen healthy males, with a mean age of 25 (SEM 5) (range 18–30) years and normal body weight for their height (BMI of 22.5 (SEM 3.0) (range 19–25 kg/m²)), were recruited. The number of subjects included was based on power calculations derived from previous works^(4,22,23). The subjects were unrestrained eaters (score 3 (SEM 1) (range 0–12) on the eating restraint section (factor 1) of the Three-Factor Eating Questionnaire⁽²⁴⁾), had no significant gastrointestinal symptoms, disease or surgery, and were not taking any medication known to affect the gastrointestinal function or appetite. The subjects who regularly consumed >20 g alcohol or more than ten cigarettes per day were excluded. The study protocol was approved by the Royal Adelaide Hospital Research Ethics Committee, and all experiments were carried out in accordance with the Declaration of Helsinki. All subjects provided informed written consent before their enrolment. To distract from the primary aim of the study, the subjects were informed that the study was designed to evaluate the effects of a glucose drink on gastric emptying and gut hormone secretion.

Protocol

Each subject participated on three occasions, separated by 7–10 d. To standardise the study conditions further, the subjects were provided with a 'ready-to-eat' dinner (beef lasagne, 2472 kJ; McCain Foods Pty Ltd, Victoria, Australia) to be consumed at 20.00 hours on the evening before each study day, after which time they were required to fast. On the study days, the subjects attended the laboratory in the Discipline of Medicine at either 08.00 or 11.00 hours, i.e. two subjects could potentially be studied on 1 d; each subject attended at the same time of day on each visit. Upon arrival, an intravenous cannula was placed into a forearm vein for blood sampling, and the subject was seated comfortably in an upright position for the duration of the study. At $t = -15$ min, an image of the

fasted stomach was acquired using 3D ultrasound, a baseline blood sample collected and a visual analogue scale (VAS) questionnaire, assessing perceptions of appetite, completed. At $t = -2$ min, the subject consumed a preload consisting of 50 g glucose dissolved in 300 ml of water (815 kJ) within 2 min. At $t = 0$ min, immediately following the ingestion of the preload, another 3D image of the stomach was acquired, a blood sample collected and a VAS questionnaire completed. Subsequently, 3D ultrasound scans, blood samples and VAS scores were obtained at 15 min intervals until $t = 90$ min. At $t = 90$ min, the subject was presented with a standardised cold buffet-style meal, with food in excess of what they would be anticipated to consume, and invited to eat until comfortably full, for up to 30 min (i.e. $t = 90$ –120 min)⁽¹²⁾. The meal comprised four slices (125 g) of wholemeal bread, four slices (125 g) of white bread, 100 g sliced ham, 100 g sliced chicken, 85 g sliced cheddar cheese, 100 g lettuce, 100 g sliced tomato, 100 g sliced cucumber, 20 g mayonnaise, 20 g margarine, 170 g apple, 190 g banana, 200 g strawberry yogurt, 150 g chocolate custard, 140 g fruit salad, 600 ml iced coffee, 500 ml orange juice and 600 ml water. The total energy content of the buffet meal was 11 808 kJ. A final blood sample was collected and a VAS questionnaire completed following the meal ($t = 120$ min), after which the intravenous cannula was removed, and the subject was free to leave the laboratory.

Measurements

Gastric emptying and intragastric meal distribution. Gastric emptying was assessed using 3D ultrasonography (Logiq™ 9 ultrasound system; GE Medical Systems, Milwaukee, WI, USA), which we have validated against the 'gold standard' scintigraphy as an accurate measure of the gastric emptying of liquids in healthy subjects⁽²²⁾. This technique allows the evaluation of total, proximal and distal gastric volumes (i.e. gastric emptying and intragastric meal distribution)⁽²²⁾. For the 3D positioning and orientation measurement, a transmitter was placed next to the subject and a 3D sensor was attached to a 3-5C broad-spectrum 2.5–4 MHz convex transducer. All metal objects were removed from both the subject and the surrounding area to avoid the possibility of interference during acquisition. Three-dimensional sweeps of the total stomach were taken to evaluate the total gastric volume using EchoPAC-3D software® (GE Vingmed Sound, Horten, Norway). The raw data (original scan planes) were used for the 3D reconstructions of the stomach. The proximal and distal gastric segments were separated by vertically slicing the 3D stomach reconstruction from the incisura angularis at the lesser gastric curvature sagittally towards the greater curvature⁽²⁵⁾. Total, proximal and distal gastric volumes at each time point were derived and expressed as percentages of the volumes at $t = 0$ min (volume immediately following preload ingestion), with total gastric volume at $t = 0$ min defined as 100%. Gastric emptying profiles were then constructed, and the time at which 50% of the preload had emptied from the stomach (50% gastric emptying time (T_{50})) was derived⁽²²⁾.

Blood glucose, plasma insulin and plasma cholecystokinin concentrations

Venous blood glucose concentrations (mM) were determined immediately after the completion of each study by the glucose

oxidase method, using a portable glucometer (Medisense Precision QID; Abbott Laboratories, Bedford, MA, USA). This technique has a CV of 2.1–5.6%. The accuracy of this method has been confirmed in our laboratory using the hexokinase technique⁽²⁶⁾.

For the measurement of insulin and CCK concentrations, venous blood samples (10 ml) were collected in chilled EDTA-treated tubes containing 400 kIU aprotinin per ml of blood (Trasylol; Bayer Australia Ltd, Pymble, Australia). The samples were centrifuged at 3200 rpm for 15 min at 4°C, within 30 min of collection, and stored at –70°C until assayed.

Plasma insulin concentrations (mU/l) were measured by ELISA (Diagnostics Systems Laboratories Inc., Webster, TX, USA). The sensitivity of the assay was 0.26 mU/l, and the intra- and interassay CV were 2.6 and 6.2%, respectively.

Plasma CCK concentrations (pM) were determined by RIA following ethanol extraction⁽¹¹⁾. The sensitivity of the assay was 2.5 pM, and the intra- and interassay CV were 9 and 27%, respectively.

Appetite perceptions and energy intake. Perceptions of fullness and desire to eat were assessed using a validated VAS questionnaire⁽²⁷⁾. Nausea and bloating were also assessed. Other perceptions, including happiness and drowsiness, were assessed to distract the subject from the main purpose of the questionnaire, but were not formally evaluated. Each VAS consisted of a 100 mm horizontal line, where 0 mm represented ‘sensation not felt at all’ and 100 mm ‘sensation felt the greatest’. The subject was required to place a vertical mark along the line to indicate the strength of each sensation.

The amount (g) of food consumed from the buffet meal was determined by weighing the meal before and after consumption. Energy intake (kJ) and macronutrient composition (percentage of energy from fat, carbohydrate and protein) were analysed using commercially available software (Foodworks 3.01; Xyris Software, Highgate Hill, Queensland, Australia)⁽¹²⁾.

Statistical analysis

Areas under the curve (AUC) for gastric emptying, blood glucose, plasma insulin and CCK concentrations were calculated using the trapezoidal rule. Peak concentrations for blood glucose, plasma insulin and CCK were defined as the greatest mean change from baseline in each subject at any given time point for each visit. Intrasubject reproducibility (i.e. the agreement within each individual’s data) between the three visits for T_{50} , AUC, peak concentrations and time-to-peak concentrations for blood glucose, plasma insulin and CCK, energy intake (kJ), weight of total food consumed (g), macronutrient distribution (i.e. percentage of energy derived from fat, carbohydrates and protein) and weight of individual food items consumed (g) from the buffet meal was evaluated by determining intraclass correlation coefficients, $r_i^{(28)}$. An $r_i \geq 0.8$ was considered to indicate excellent agreement, $0.8 > r_i \geq 0.7$ indicating good agreement and $0.7 > r_i \geq 0.6$ indicating moderate agreement⁽²⁹⁾. For variables measured over time (percentage of retention for total gastric volume, intragastric meal distribution, plasma hormone concentrations and VAS scores), repeated-measures ANOVA was used to evaluate any differences between visits, with visit and time as factors. The relationships between gastric emptying, T_{50} , peak concentrations and concentrations at $t = 90$ min of

blood glucose, plasma insulin and CCK with energy intake, and between gastric emptying and T_{50} with blood glucose and plasma insulin concentrations, were calculated using partial correlations⁽³⁰⁾. Statistical significance was accepted at $P < 0.05$. Data are presented as raw values and expressed as means with their standard errors. Based on the day-to-day variations observed in our sample, we calculated the minimum effect sizes required to detect a hypothetical treatment effect (with 80% power) for our measured parameters (i.e. gastric emptying, blood glucose, plasma insulin and CCK concentrations and mean energy intake). Since this was a repeated-measures study design, the correlation between these measures can be assumed, and for each of our measured parameters, the intraclass correlation coefficient was estimated. This value was then used in the SAS macro fpower (SAS version 9.1, Cary, NC, USA) to calculate the minimum difference between the data points of interest that would be detected by a sample of fifteen subjects.

Results

The study protocol was well tolerated, and all subjects completed all visits. Mean data for baseline values and in response to treatment are summarised in Tables 1 and 2, respectively. There were no differences in baseline values for any of the parameters, including desire to eat, either the 08.00 or 11.00 hour visits, nor did the timing of the commencement of the studies (i.e. 08.00 or 11.00 hours) affect any of the outcome measures (data not shown).

Gastric emptying

Total gastric emptying. Gastric emptying occurred in an overall linear pattern (Fig. 1 (a)), resulting in an average emptying rate of 8.8 ± 1.7 , 11.7 ± 0.4 and 7.1 ± 0.4 kJ/min for visits 1, 2 and 3 ($r_i = 0.90$), respectively. There was no difference in gastric emptying profiles between visits. Less than 30% (approximately 15 g glucose) of the drink remained in the stomach at the end of the study on all visits. There was significant intraindividual variation between visits in the T_{50} ($r_i = 0.12$) and the AUC of gastric emptying ($r_i = 0.23$).

Intragastric meal distribution. There was no difference in the amount of glucose retained in the proximal or distal stomach between visits (Fig. 1 (b)). The amount of glucose

Table 1. Baseline values for blood glucose, plasma insulin and cholecystokinin (CCK) concentrations and appetite and symptom ratings (Mean values with their standard errors)

Variable	Visit 1		Visit 2		Visit 3	
	Mean	SEM	Mean	SEM	Mean	SEM
Blood glucose (mm)	5.4	0.2	5.0	0.2	5.2	0.2
Plasma insulin (mU/l)	10.6	0.1	11.1	2.3	8.3	1.1
Plasma CCK (pM)	4.0	0.3	4.0	0.3	4.0	0.4
VAS ratings						
Desire to eat (mm)	54.0	8.0	62.0	8.0	63.0	8.0
Fullness (mm)	10.0	4.0	9.0	3.0	9.0	4.0
Nausea (mm)	6.0	2.0	5.0	3.0	3.0	1.0
Bloating (mm)	3.0	1.0	4.0	2.0	3.0	1.0

VAS, visual analogue scale.

Table 2. Mean values for gastric half-emptying time, blood glucose, plasma insulin and cholecystokinin (CCK) concentrations and food intake (Mean values with their standard errors)

Variable	Visit 1		Visit 2		Visit 3	
	Mean	SEM	Mean	SEM	Mean	SEM
Gastric emptying						
Half-emptying time (T_{50}) (min)	58.6	4.7	61.0	3.0	64.5	3.9
AUC (min% retention)	5228.0	271.0	5508.0	242.0	5574.0	263.0
Blood glucose						
Peak concentration (mM)	9.8	0.7	9.7	0.6	9.5	0.5
Time-to-peak concentration (min)	35.0	3.0	36.0	2.0	37.0	2.0
AUC (min mM)	697.0	49.0	698.0	40.0	689.0	34.0
Plasma insulin						
Peak concentration (mU/l)	69.0	10.0	77.0	13.0	63.0	9.0
Time-to-peak concentration (min)	47.0	4.0	49.0	5.0	51.0	6.0
AUC (min mU/l)	4077.0	580.0	4896.0	915.0	3874.0	563.0
Plasma CCK						
Peak concentration (pM)	6.8	0.4	7.6	0.5	7.2	0.6
Time-to-peak concentration (min)	23.0	4.0	22.0	4.0	23.0	4.0
AUC (min pM)	563.0	36.0	599.0	38.0	572.0	41.0
Food intake						
Energy intake (kJ)	3811.0	289.0	3632.0	338.0	3919.0	296.0
Weight of food (g)	915.0	70.0	896.0	92.0	937.0	97.0
Protein (%)	20.0	1.0	20.0	1.0	21.0	1.0
Fat (%)	29.0	2.0	30.0	2.0	30.0	1.0
Carbohydrate (%)	51.0	2.0	50.0	3.0	49.0	1.0

AUC, area under the curve.

in the proximal stomach decreased progressively over time (time effect: $P < 0.001$), reflecting total gastric emptying, while the amount in the distal stomach remained relatively constant.

Blood glucose and plasma hormone concentrations

Blood glucose. There was no difference in the overall blood glucose concentrations between visits (Fig. 2 (a)). The blood glucose rose within 15 min of glucose ingestion (time effect: $P < 0.01$) and decreased from $t = 60$ min. There was a good intraindividual agreement for AUC under the blood glucose profiles ($r_i = 0.72$) and for peak blood glucose concentrations ($r_i = 0.72$). By contrast, the time taken for blood glucose concentrations to peak varied between visits ($r_i = 0.33$). Blood glucose concentrations returned to baseline values following the buffet meal, with no differences in the values between visits.

Plasma insulin. There was no difference in the overall plasma insulin concentrations between visits (Fig. 2 (b)). Plasma insulin rose within 15 min of glucose ingestion (time effect: $P < 0.01$) and decreased from $t = 60$ min. There was an excellent intraindividual agreement for AUC under plasma insulin profiles ($r_i = 0.88$) and for peak insulin concentrations ($r_i = 0.81$). By contrast, the time taken for the plasma insulin concentrations to peak varied between visits ($r_i = 0.41$). Plasma insulin rose again in response to the buffet meal (time effect: $P < 0.01$), with no difference in the magnitude of the increase between visits.

Plasma cholecystokinin. There was no difference in the overall plasma CCK concentrations between visits (Fig. 2 (c)). Plasma CCK rose within 15 min of glucose ingestion (time effect: $P < 0.01$) and subsequently reached a plateau. There was an excellent intraindividual agreement for AUC under

plasma CCK profiles ($r_i = 0.84$) and for the time to peak of plasma CCK concentrations ($r_i = 0.89$), and a good agreement for peak plasma CCK concentrations ($r_i = 0.70$) between visits. Plasma CCK rose again in response to the meal (time effect: $P < 0.01$), with no difference in the magnitude of the increase between visits.

Appetite and energy intake

Appetite. There was no effect of treatment on scores for fullness or desire to eat between visits. Fullness increased (time effect: $P < 0.05$) in response to glucose ingestion until $t = 15$ min, after which time scores gradually returned to baseline. Desire to eat remained unchanged from the baseline (Fig. 3). Fullness rose, and desire to eat decreased, in response to the buffet meal (time effect: $P < 0.001$), with no differences between visits. No subject experienced nausea or bloating (data not shown).

Energy intake. Both energy intake ($r_i = 0.89$) and the amount of food consumed ($r_i = 0.80$) showed excellent agreement between visits. The agreement between visits for macronutrient composition was good for protein ($r_i = 0.77$), moderate for fat ($r_i = 0.68$) and low for carbohydrate ($r_i = 0.54$).

For approximately 50% of the foods contained in the buffet meal, there was either an excellent (for margarine, mayonnaise, custard, chicken and banana) or a good (for tomato, ham, cucumber and iced coffee) agreement, and for approximately 16% of the foods (cheese, fruit salad and fruit yoghurt), there was a moderate agreement, between visits. The agreement between visits was low for five buffet meal items (wholemeal and white breads, orange juice, lettuce and water; Table 3).

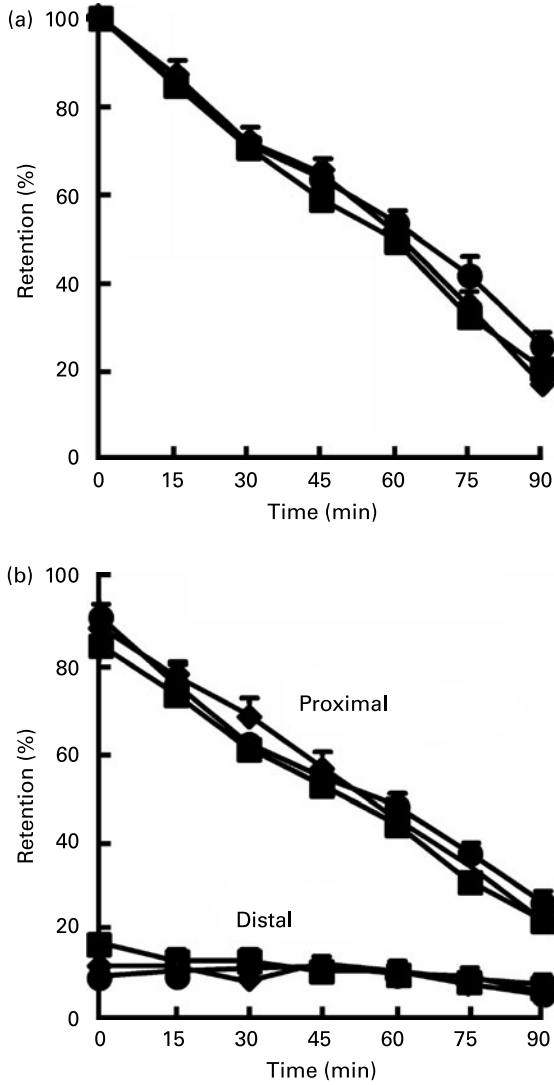


Fig. 1. (a) Total and (b) proximal and distal gastric emptying (percentage of retention) of a 'preload' containing 50 g glucose in 300 ml water on three different days. Data are mean values with their standard errors (n 15). Visit 1, \blacksquare ; visit 2, \blacklozenge ; visit 3, \bullet .

Relationships between gastric emptying, blood glucose, plasma insulin and cholecystokinin with energy intake

There were no significant relationships between energy intake with the amount of glucose remaining in the stomach at $t = 90$ min (i.e. immediately before the buffet meal), T_{50} , peak concentrations of blood glucose, plasma insulin and CCK, blood glucose, plasma insulin or CCK concentrations at $t = 90$ min (data not shown).

Relationships between blood glucose and plasma insulin concentrations with gastric emptying

There were inverse relationships between blood glucose concentrations at $t = 30$ min ($r = -0.33$, $P < 0.05$) and $t = 45$ min ($r = -0.29$, $P = 0.05$) with the amount of glucose remaining in the stomach at these times, and direct relationships between blood glucose concentrations between $t = 15$ and 90 min ($r > 0.38$, $P < 0.01$) with T_{50} , and the change in

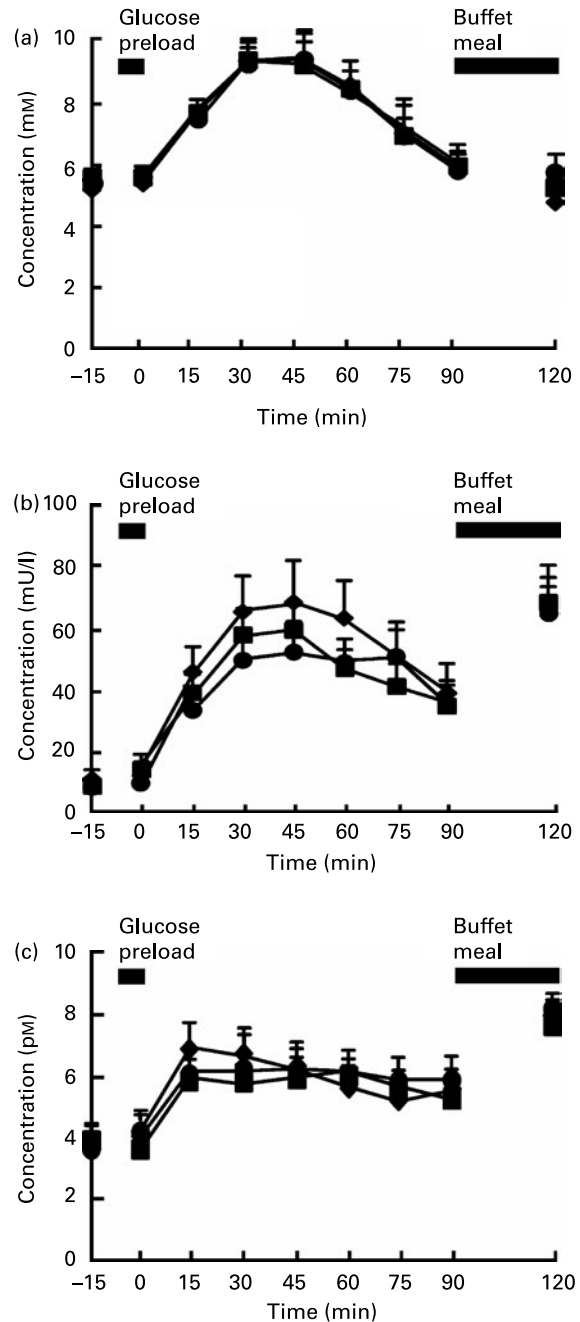


Fig. 2. (a) Blood glucose, (b) plasma insulin and (c) plasma CCK concentrations following ingestion of a 'preload' containing 50 g glucose in 300 ml water on three different days. Data are mean values with their standard errors (n 15). Visit 1, \blacksquare ; visit 2, \blacklozenge ; visit 3, \bullet .

blood glucose from baseline at $t = 30$ min ($r = 0.32$, $P < 0.05$) and $t = 45$ min ($r = 0.32$, $P < 0.05$) with the rate of gastric emptying (kcal/min).

There were inverse relationships between the plasma insulin concentrations at $t = 30$ min ($r = -0.35$, $P < 0.05$) and $t = 45$ min ($r = -0.38$, $P < 0.01$) with the amount of glucose drink retained in the stomach at these times, and direct relationships between the change in the plasma insulin from the baseline at $t = 30$ min ($r = 0.31$, $P < 0.05$), $t = 45$ min ($r = 0.30$, $P < 0.05$) and $t = 90$ min ($r = 0.38$, $P = 0.01$) with the rate of gastric emptying (kcal/min).

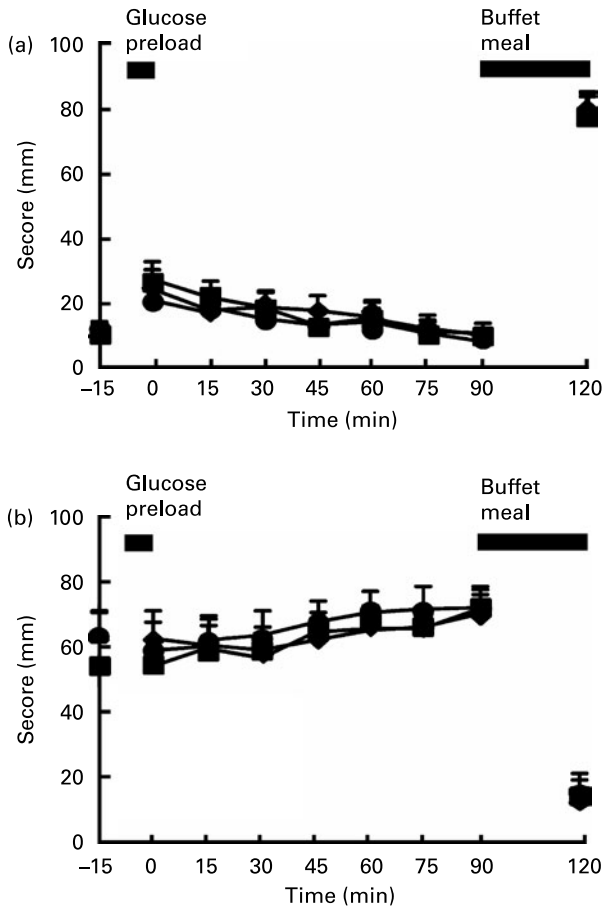


Fig. 3. Scores for (a) fullness and (b) desire to eat following the ingestion of a 'preload' containing 50 g glucose in 300 ml water on three different days. Data are mean values with their standard errors ($n = 15$). Visit 1, \blacksquare ; visit 2, \blacklozenge ; visit 3, \bullet .

Table 3. Day-to-day reproducibility of food items consumed at the buffet meal

Food item	r_i
Margarine	0.93
Mayonnaise	0.88
Custard	0.86
Chicken	0.84
Banana	0.82
Tomato	0.77
Ham	0.74
Cucumber	0.72
Iced coffee	0.71
Cheese	0.66
Fruit salad	0.66
Fruit yoghurt	0.66
Wholemeal bread	0.56
Orange juice	0.50
White bread	0.41
Lettuce	0.36
Water	0.30
Apple	Not consumed

r_i , intraclass correlation coefficient, defined as follows: $r_i \geq 0.8$ excellent agreement; $0.8 > r_i \geq 0.7$ good agreement; $0.7 > r_i \geq 0.6$ moderate agreement between visits.

There was a direct relationship between the blood glucose and the plasma insulin concentrations at $t = 15$ min ($r = 0.53$, $P = 0.001$).

Calculation of minimum effect size for gastric emptying, blood glucose, plasma hormones and energy intake, based on observed intraindividual variations

Based on the day-to-day variations observed in our sample, in order to detect a treatment effect, we calculated that minimum mean effect sizes for gastric emptying AUC would have to be ≥ 866 min%, gastric emptying $T_{50} \geq 13.2$ min, blood glucose AUC ≥ 130 min mm, time-to-peak blood glucose ≥ 6.8 min, peak blood glucose concentration ≥ 1.97 mM, plasma insulin AUC ≥ 2230 min mU/l, time-to-peak plasma insulin ≥ 20.3 min, peak plasma insulin concentration ≥ 33.5 mU/l, plasma CCK AUC ≥ 162 min pM, time-to-peak plasma CCK ≥ 11.9 min, peak plasma CCK concentration ≥ 2.20 pM and energy intake ≥ 916 kJ.

Discussion

The present observations indicate that, in a laboratory setting: (1) appetite perceptions and energy intake in response to a nutrient preload in healthy lean men are highly reproducible; and (2) this consistency in energy intake is associated with reproducible patterns of gastric emptying, plasma insulin and CCK secretion. In addition, to our knowledge, the present study is the first to provide information about minimum effect sizes for our measured parameters required to detect a hypothetical treatment effect based on our data on intraindividual variations in response to the same treatment.

In the present study, both energy intake (kJ) and the amount of food consumed (g) from a test meal in response to a glucose preload showed very good reproducibility between the three visits, supporting the hypothesis that, at least in a laboratory setting, acute energy intake does not markedly change on a day-to-day basis in healthy lean men. This observation is in agreement with previous studies that demonstrated good reproducibility of energy intake using a standardised buffet meal^(13,15). It had been suggested that subjects tend to over-consume energy when allowed *ad libitum* access to a variety of sandwiches⁽⁶⁾, and that this could potentially relate to the sense of novelty that is associated with a selection of appetising foods, as in the case of a buffet-style meal. Conversely, the subjects may experience a sense of boredom when presented with the same meal over a number of occasions, resulting in a reduction in energy intake at later visits. Hence, a limitation of the study by Arvaniti *et al.*⁽¹³⁾ is that energy intake was assessed only on two occasions. In the present study, we were able to demonstrate that energy intake is reproducible when assessed on three occasions; thus, neither the variety of food presented with the buffet meal nor providing the subjects with the same foods on multiple occasions appeared to influence energy intake. The excellent agreement in energy intakes between the days in both the present and the previous^(13,15) studies, therefore, indicates that the use of a buffet-style meal produces a reliable measure of energy intake. Taken together, the data suggest that the magnitude of the effects on energy intake observed previously in response to either an oral nutrient preload or a duodenal

nutrient infusion^(4,31) is significant, and that the number of subjects required to observe these effects is sufficient.

While energy intake and the amount of food consumed were both highly reproducible between study days, the macronutrient composition of the food consumed, although still showing moderate to good agreement, varied more between the three visits. The greater variation in the macronutrient composition, despite very little variation in energy intakes between visits, may reflect the selection of foods available, i.e. individuals tended to select different items with varying macronutrient contents from the meal on each occasion to achieve their overall energy intake. Hence, the present data suggest that buffet-style meals, as used in the present study, are highly suitable for the evaluation of total energy intake, but less effective in evaluating the macronutrient distribution, or food choice. How isoenergetic buffet-style meals varying in their food composition may affect energy intake and macronutrient composition has not been evaluated, but warrants the investigation.

The consistency in energy intake was associated with reproducible patterns of gastric emptying, intragastric meal distribution, glycaemia, insulinaemia and CCK secretion, suggesting that these factors may have contributed to reproducible energy intakes in response to the nutrient preload. By contrast, the statistical analysis indicated a lack of agreement between visits for the time taken for 50% of the meal to empty from the stomach. Gastric half-emptying time (T_{50}) has been used widely as a measure of gastric emptying, particularly in scintigraphic studies^(32–35); however, it is not as well established for 3D ultrasound studies. For example, while an agreement has been demonstrated between scintigraphy and 3D ultrasound, the limits of agreement for the T_{50} of 300 ml dextrose solution (25%), as measured by 3D ultrasound for gastric emptying profiles, were -35.3 min to $+47.6$ min, which, while statistically not different from the data obtained scintigraphically, were highly variable⁽²²⁾. It is important to recognise that 3D ultrasonography is associated with some limitations. The presence of intragastric air, particularly in the fundus⁽³⁶⁾, has the potential to compromise visualisation of the gastric outline, and this may have contributed to the lack of agreement in T_{50} between visits.

In the present study, there were direct relationships between the blood glucose and the plasma insulin responses with the rate of gastric emptying, consistent with previous observations^(37,38). It is well established that CCK mediates, at least in part, the effects of nutrients, particularly fat, on gastrointestinal motility and energy intake^(39,40). Thus, while the present finding that both gastric emptying and energy intake in response to the glucose preload were unrelated to plasma CCK concentrations may appear surprising, it is likely to reflect relatively modest stimulation of CCK by glucose. Dietary lipid and protein are known to be much more potent stimuli for CCK release than glucose⁽⁴¹⁾.

The present study has calculated effect sizes based on intraindividual variations in response to a standardised treatment on repeated occasions. The present data, from a relatively small sample of fifteen subjects, suggest that quite large effect sizes in response to a treatment are required to detect a significant treatment effect, indicating that, while there was a very good statistical agreement for most parameters between the study days, variability is still substantial. Thus, in circumstances where small differences between the

treatments need to be detected, the sample size may need to be quite large, particularly in the studies evaluating energy intake.

Some limitations of the present study warrant discussion. First, we evaluated only healthy lean males, hence the present observations may not be applicable to females and other subject groups, i.e. under- or overweight and obese. While the number of subjects included was based on power calculations derived from our previous studies, it is nonetheless possible that small differences in some of the outcome measures may not have been detected. Finally, potential changes in individual food preferences were not evaluated over the three visits, although it is clear that, in general, subjects selected the majority of food items included in the buffet meal.

In conclusion, we have demonstrated that, when measured repeatedly in a laboratory setting, energy intake in response to a nutrient preload is reproducible in healthy lean males, and this consistency is associated with reproducible patterns of gastric emptying and gastrointestinal hormone secretion.

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References

1. Cook CG, Andrews JM, Jones KL, Wittert GA, Chapman IM, Morley JE & Horowitz M (1997) Effects of small intestinal nutrient infusion on appetite and pyloric motility are modified by age. *Am J Physiol* **273**, R755–R761.

2. Cecil JE, Francis J & Read NW (1998) Relative contributions of intestinal, gastric, oro-sensory influences and information to changes in appetite induced by the same liquid meal. *Appetite* **31**, 377–390.
3. Cecil JE, Francis J & Read NW (1999) Comparison of the effects of a high-fat and high-carbohydrate soup delivered orally and intragastrically on gastric emptying, appetite, and eating behaviour. *Physiol Behav* **67**, 299–306.
4. Feinle C, O'Donovan D, Doran S, Andrews JM, Wishart J, Chapman I & Horowitz M (2003) Effects of fat digestion on appetite, APD motility, and gut hormones in response to duodenal fat infusion in humans. *Am J Physiol Gastrointest Liver Physiol* **284**, G798–G807.
5. O'Donovan D, Feinle-Bisset C, Wishart J & Horowitz M (2003) Acute effects of lipase inhibition on subsequent food intake in humans. *Br J Nutr* **90**, 849–852.
6. Norton GN, Anderson AS & Hetherington MM (2006) Volume and variety: relative effects on food intake. *Physiol Behav* **87**, 714–722.
7. Sturm K, MacIntosh CG, Parker BA, Wishart J, Horowitz M & Chapman IM (2003) Appetite, food intake, and plasma concentrations of cholecystokinin, ghrelin, and other gastrointestinal hormones in undernourished older women and well-nourished young and older women. *J Clin Endocrinol Metab* **88**, 3747–3755.
8. Rolls BJ, Kim-Harris S, Fischman MW, Foltin RW, Moran TH & Stoner SA (1994) Satiety after preloads with different amounts of fat and carbohydrate: implications for obesity. *Am J Clin Nutr* **60**, 476–487.
9. Shide DJ, Caballero B, Reidelberger R & Rolls BJ (1995) Accurate energy compensation for intragastric and oral nutrients in lean males. *Am J Clin Nutr* **61**, 754–764.
10. Feinle-Bisset C & Horowitz M (2006) Dietary factors in functional dyspepsia. *Neurogastroenterol Motil* **18**, 608–618.
11. MacIntosh CG, Morley JE, Wishart J, Morris H, Jansen JB, Horowitz M & Chapman IM (2001) Effect of exogenous cholecystokinin (CCK)-8 on food intake and plasma CCK, leptin, and insulin concentrations in older and young adults: evidence for increased CCK activity as a cause of the anorexia of aging. *J Clin Endocrinol Metab* **86**, 5830–5837.
12. Feltrin KL, Little TJ, Meyer JH, *et al.* (2004) Effects of intraduodenal fatty acids on appetite, antropyloroduodenal motility, and plasma CCK and GLP-1 in humans vary with their chain length. *Am J Physiol Regul Integr Comp Physiol* **287**, R524–R533.
13. Arvaniti K, Richard D & Tremblay A (2000) Reproducibility of energy and macronutrient intake and related substrate oxidation rates in a buffet-type meal. *Br J Nutr* **83**, 489–495.
14. Kral TV, Roe LS & Rolls BJ (2004) Combined effects of energy density and portion size on energy intake in women. *Am J Clin Nutr* **79**, 962–968.
15. Gregersen NT, Flint A, Bitz C, Blundell JE, Raben A & Astrup A (2008) Reproducibility and power of *ad libitum* energy intake assessed by repeated single meals. *Am J Clin Nutr* **87**, 1277–1281.
16. Castiglione KE, Read NW & French SJ (1998) Food intake responses to upper gastrointestinal lipid infusions in humans. *Physiol Behav* **64**, 141–145.
17. French SJ, Conlon CA, Mutuma ST, Arnold M, Read NW, Meijer G & Francis J (2000) The effects of intestinal infusion of long-chain fatty acids on food intake in humans. *Gastroenterology* **119**, 943–948.
18. MacIntosh CG, Horowitz M, Verhagen MA, Smout AJ, Wishart J, Morris H, Goble E, Morley JE & Chapman IM (2001) Effect of small intestinal nutrient infusion on appetite, gastrointestinal hormone release, and gastric myoelectrical activity in young and older men. *Am J Gastroenterol* **96**, 997–1007.
19. Sepple CP & Read NW (1989) Gastrointestinal correlates of the development of hunger in man. *Appetite* **13**, 183–191.
20. Jones KL, Doran SM, Hveem K, Bartholomeusz FD, Morley JE, Sun WM, Chatterton BE & Horowitz M (1997) Relation between postprandial satiation and antral area in normal subjects. *Am J Clin Nutr* **66**, 127–132.
21. Kissileff HR, Pi-Sunyer FX, Thornton J & Smith GP (1981) C-terminal octapeptide of cholecystokinin decreases food intake in man. *Am J Clin Nutr* **34**, 154–160.
22. Gentilcore D, Hausken T, Horowitz M & Jones KL (2006) Measurements of gastric emptying of low- and high-nutrient liquids using 3D ultrasonography and scintigraphy in healthy subjects. *Neurogastroenterol Motil* **18**, 1062–1068.
23. MacIntosh CG, Andrews JM, Jones KL, Wishart JM, Morris HA, Jansen JB, Morley JE, Horowitz M & Chapman IM (1999) Effects of age on concentrations of plasma cholecystokinin, glucagon-like peptide 1, and peptide YY and their relation to appetite and pyloric motility. *Am J Clin Nutr* **69**, 999–1006.
24. Stunkard AJ & Messick S (1985) The three-factor eating questionnaire to measure dietary restraint, disinhibition and hunger. *J Psychosom Res* **29**, 71–83.
25. Tefera S, Gilja OH, Olafsdottir E, Hausken T, Hatlebakk JG & Berstad A (2002) Intragastric maldistribution of a liquid meal in patients with reflux oesophagitis assessed by three dimensional ultrasonography. *Gut* **50**, 153–158.
26. Horowitz M, Maddox AF, Wishart JM, Harding PE, Chatterton BE & Shearman DJ (1991) Relationships between oesophageal transit and solid and liquid gastric emptying in diabetes mellitus. *Eur J Nucl Med* **18**, 229–234.
27. Parker B, Sturm K, MacIntosh CG, Feinle C, Horowitz M & Chapman IM (2004) Relation between food intake and visual analogue scale ratings of appetite and other sensations in healthy older and young subjects. *Eur J Clin Nutr* **58**, 212–218.
28. Shrout PE & Fleiss JL (1979) Intraclass correlations: uses in assessing rater reliability. *Psychol Bull* **2**, 420–428.
29. Choi JS, Wexner SD, Nam YS, Mavrantonis C, Salum MR, Yamaguchi T, Weiss EG, Nogueras JJ & Yu CF (2000) Intraobserver and interobserver measurements of the anorectal angle and perineal descent in defecography. *Dis Colon Rectum* **43**, 1121–1126.
30. Bland JM & Altman DG (1995) Calculating correlation coefficients with repeated observations: part 1 – correlation within subjects. *BMJ* **310**, 446.
31. Sturm K, Parker B, Feinle-Bisset C, Jones KL, Chapman I & Horowitz M (2004) Energy intake and appetite are related to antral area in healthy young and older subjects. *Am J Clin Nutr* **80**, 656–667.
32. Sidery MB, Macdonald IA & Blackshaw PE (1994) Superior mesenteric artery blood flow and gastric emptying in humans and the differential effects of high fat and high carbohydrate meals. *Gut* **35**, 186–190.
33. Hveem K, Jones KL, Chatterton BE & Horowitz M (1996) Scintigraphic measurement of gastric emptying and ultrasonographic assessment of antral area: relation to appetite. *Gut* **38**, 816–821.
34. Jones KL, Tonkin A, Horowitz M, Wishart JM, Carney BI, Guha S & Green L (1998) Rate of gastric emptying is a determinant of postprandial hypotension in non-insulin-dependent diabetes mellitus. *Clin Sci (Lond)* **94**, 65–70.
35. Naslund E, Bogefors J, Gryback P, Jacobsson H & Hellstrom PM (2000) Gastric emptying: comparison of scintigraphic, polyethylene glycol dilution, and paracetamol tracer assessment techniques. *Scand J Gastroenterol* **35**, 375–379.
36. Gilja OH, Detmer PR, Jong JM, Leotta DF, Li XN, Beach KW, Martin R & Strandness DE Jr (1997) Intragastric distribution and gastric emptying assessed by three-dimensional ultrasonography. *Gastroenterology* **113**, 38–49.

37. Rayner CK, Samsom M, Jones KL & Horowitz M (2001) Relationships of upper gastrointestinal motor and sensory function with glycemic control. *Diabetes Care* **24**, 371–381.
38. Horowitz M, Edelbroek MA, Wishart JM & Straathof JW (1993) Relationship between oral glucose tolerance and gastric emptying in normal healthy subjects. *Diabetologia* **36**, 857–862.
39. Matzinger D, Gutzwiller JP, Drewe J, Orban A, Engel R, D'Amato M, Rovati L & Beglinger C (1999) Inhibition of food intake in response to intestinal lipid is mediated by cholecystokinin in humans. *Am J Physiol* **277**, R1718–R1724.
40. Katschinski M, Schirra J, Beglinger C, Langbein S, Wank U, D'Amato M & Arnold R (1996) Intestinal phase of human antro-pyloro-duodenal motility: cholinergic and CCK-mediated regulation. *Eur J Clin Invest* **26**, 574–583.
41. Liddle RA, Goldfine ID, Rosen MS, Taplitz RA & Williams JA (1985) Cholecystokinin bioactivity in human plasma. Molecular forms, responses to feeding, and relationship to gallbladder contraction. *J Clin Invest* **75**, 1144–1152.