

## Research Article

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# Phenology and resource allocation strategies of diploid flowering rush (*Butomus umbellatus*) in Ohio and New York

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## Abstract

Flowering rush (*Butomus umbellatus* L.) is an emergent perennial monocot that has invaded aquatic systems along the U.S.–Canadian border. Currently, there are two known cytotypes of flowering rush, diploid and triploid, within the invaded range. Although most studies have focused on the triploid cytotype, little information is known about diploid plants. Therefore, phenology and resource allocation were studied on the diploid cytotype of flowering rush in three study sites (Mentor Marsh, OH; Tonawanda Wildlife Management Area, NY; and Unity Island, NY) to understand seasonal resource allocation and environmental influences on growth, and to optimize management strategies. Samples were harvested once a month from May to November at each site from 2021 to 2023. Plant metrics were regressed to air temperature, water temperature, and water depth. Aboveground biomass peaked from July to September and comprised 50% to 70% of total biomass. Rhizome biomass peaked from September to November and comprised 40% to 50% of total biomass. Rhizome bulbil densities peaked from September to November at 3,000 to 16,000 rhizome bulbils m<sup>-2</sup>. Regression analysis resulted in strong negative relationships between rhizome starch content and air temperature ( $r^2 = 0.52$ ) and water temperature ( $r^2 = 0.46$ ). Other significant, though weak, relationships were found, including a positive relationship between aboveground biomass and air temperature ( $r^2 = 0.17$ ), a negative relationship between rhizome bulbil biomass and air temperature ( $r^2 = 0.18$ ) and a positive relationship between leaf density and air temperature ( $r^2 = 0.17$ ). Rhizomes and rhizome bulbils combined stored up to 60% of total starch, and therefore, present a unique challenge to management, as these structures cannot be reached directly with herbicides. Therefore, management should target the aboveground tissue before peak production (July) to reduce internal starch storage and aim to limit regrowth over several years.

## Introduction

Flowering rush (*Butomus umbellatus* L.) is an invasive perennial monocot primarily found in aquatic and wetland habitats along the U.S.–Canadian border. *Butomus umbellatus* was first documented in the St Lawrence River, which facilitated spread into the Great Lakes region by the mid-1950s (Bellaud 2009; Gunderson et al. 2016). *Butomus umbellatus* is thought to have had multiple introductions into North America due to two separate cytotypes, a triploid and diploid, that have been identified with at least six amplified fragment-length polymorphism genotypes (Anderson et al. 1974; Gaskin et al. 2021). Both cytotypes of *B. umbellatus* are capable of vegetative reproduction through fragmentation of the rhizome and production of rhizome bulbils developed along the rhizome and leaf axils (Hroudová and Zákřavský 1993; Thompson and Eckert 2004). The diploid cytotype can also sexually reproduce through self-compatible flowers on an inflorescence containing 20 to 50 flowers (Thompson and Eckert 2004). Current information about *B. umbellatus* biology and management largely centers on the triploid cytotype and its invasion within lakes, whereas the diploid cytotype and impacted wetlands remain understudied.

Within aquatic systems, plants must overcome unfavorable conditions such as low light availability and limited CO<sub>2</sub> concentrations, which can impact photosynthesis and growth (Ralph et al. 2007). *Butomus umbellatus* can grow both emergent and submersed leaves (often

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### Management Implications

*Butomus umbellatus* (flowering rush) is an invasive emergent monocot commonly found along the U.S.–Canadian border that can grow rapidly and cause numerous water-use issues as a monotypic population. Within the United States, there are two cytotypes of the species, a diploid and triploid, which have slightly different reproductive strategies, with diploid reproducing both sexually and asexually and triploid plants reproducing only asexually. The current study sought to describe the life history, resource allocation, and phenology of diploid *B. umbellatus*. Diploid *B. umbellatus* in this study produced 8,000 rhizome bulbils  $m^{-2}$  on average, with one site having more than 15,000 rhizome bulbils  $m^{-2}$ . Not only does diploid *B. umbellatus* produce large numbers of propagules, but up to 30% of total plant starch is found in rhizome bulbils, which will allow for longevity in the sediment. A successful management program will need to indirectly target the belowground biomass and starch storage of this plant. Because rhizomes and rhizome bulbils are at or below the sediment surface, they are difficult to directly treat with a herbicide or to harvest. Management programs should target aboveground biomass before July to remove new growth. Additional in-season applications should occur as plants regrow to limit rhizome bulbil formation. Repeat applications should reduce year-to-year recruitment, although management of *B. umbellatus* will be long term, as the propagule bank will need to be exhausted to completely control this species.

together) and can be found in water as deep as 6 m but is usually found at depths of <1.3 m (Carter et al. 2018; Madsen et al. 2016). Triploid *B. umbellatus* is typically found growing in lake systems, whereas diploid *B. umbellatus* is more commonly found in wetland systems with variable hydrology (Gunderson et al. 2016). Alongside reproductive differences, the triploid cytotype is more widespread within the United States, with populations found from Minnesota westward in lake or reservoir systems (Gaskin et al. 2021; Liu et al. 2005). Conversely, diploid *B. umbellatus* has been documented from Minnesota eastward and is prolific especially around the Great Lakes region within wetlands (Treibitz and Taylor 2007). Differences in geography and growing environment can impact a plant's life history, resource allocation, and phenology.

Emergent wetland plants are subject to the environment, which can impact life-history characteristics such as growth and resource accumulation (MacNeill et al. 2017; Scofield et al. 2009). Phenology is studied to understand and generate timelines for critical plant processes such as peak biomass and resource allocation patterns that can provide managers with important information to optimize the timing of control activities (Clarke et al. 2023; Wersal et al. 2011, 2013). Triploid *B. umbellatus* phenology has been documented in the Detroit Lakes system in Minnesota, where it was observed that aboveground biomass peaks in late summer and rhizomatic biomass peaks in late fall (Madsen et al. 2016; Marko et al. 2015). Due to differences in genetics between cytotypes and depths at which diploid *B. umbellatus* plants grow, characterizing phenology and resource allocation could yield important information for managing invaded sites. Whereas phenology focuses on timing of biomass changes or reproduction, resource allocation aims to understand locations within the plant where starch and other carbohydrates are typically stored (Clarke et al. 2023; Haram and Wersal 2023; Wersal et al. 2011, 2013). Management targeting critical tissues and times of low

resource allocation can reduce growth, prevent reproduction, impact nutrient acquisition, and limit annual recruitment (Scofield et al. 2009).

By combining environmental data with trends in plant growth and reproduction, a detailed understanding of phenological timing can be developed for a plant species. Previous studies on *B. umbellatus* phenology focused on the triploid cytotype within the Detroit Lakes, MN (Madsen et al. 2016; Marko et al. 2015), but there is a paucity of data describing life history, resource allocation, and phenological timings for diploid *B. umbellatus*. Documenting diploid *B. umbellatus* phenology would permit management strategies to be optimized if growth of both cytotypes are similar. Therefore, the objectives of this study were to (1) evaluate the phenology of diploid *B. umbellatus* by determining times of peak biomass and starch allocation patterns from three field populations over two growing seasons in Ohio (one population) and New York (two populations) and (2) relate *B. umbellatus* growth to air temperature, water temperature, and water depth. It is hypothesized that diploid *B. umbellatus* will have similar phenological timings with respect to peak aboveground biomass in late summer (August or September) and peak rhizome biomass in late fall (November to December). It is also hypothesized that the starch allocation within the plant will be higher in the belowground structures, similar to the pattern documented in triploid *B. umbellatus*.

### Materials and Methods

Sites chosen for this study were selected based on knowledge of historical *B. umbellatus* infestations, access for sampling, and in close proximity to the Great Lakes. The *B. umbellatus* populations in these three sites were previously determined to be diploid *B. umbellatus* genotype G4 by Gaskin et al. (2021). Subsequent genetic testing by the resource managers associated with each site reconfirmed the cytotype.

### Study Site Description

Mentor Marsh, OH (41.73649°N, 81.29879°W) is a large emergent marsh complex managed by the Cleveland Museum of Natural History (CMNH) that receives hydrologic input directly from Lake Erie. Mentor Marsh historically was used as a dumping site for salt mine tailings, which caused widespread ecosystem degradation and native plant loss. The open niche space was quickly filled by [*Phragmites australis* (Cav.) Trin. ex Steud] haplotype M, which is an aggressive invader (Guo et al. 2014; M Yeager, personal communication, March 6, 2023). In 2013, CMNH began large-scale habitat restoration efforts to manage *P. australis* in the marsh, and the disturbance from the restoration work likely allowed *B. umbellatus* to expand its range within the marsh. Currently, CMNH's goal has been to reduce *B. umbellatus* populations by hand pulling the plants directly. More recently, *B. umbellatus* invaded and has begun to spread in the wetland system.

Tonawanda Wildlife Management Area, NY (Tonawanda WMA; 43.10590°N, 78.48118°W) is a large emergent wetland complex managed by New York State Department of Environmental Conservation that undergoes seasonal water-level management through on-site water control structures. The wetland is naturally flooded in the spring (April) and can be manually or naturally drained before summer and, depending on water availability, reflooded in the fall. Historically, this site was part of glacial Lake Tonawanda until the lake naturally drained through Niagara Falls (Calkin and Brett 1978). Today, there are numerous dikes to control

water flow within the system to create a waterfowl management area through seasonal flooding and draining. *Butomus umbellatus* was first discovered in Tonawanda WMA in 2009 and has since continued to spread throughout the marsh (Kennedy et al., 2018; Roster et al., 2011; U.S. Geological Survey 2023). Tonawanda WMA staff currently aim to control *P. australis* and water chestnut (*Trapa* spp.) through herbicide applications and hand pulling, respectively.

Unity Island, NY (42.92999°N, 78.90453°W) is a wetland adjacent to the upper Niagara River near Buffalo, NY. The wetland was created in 2018 through beneficial use of dredged material from the Buffalo River federal navigation channel. *Butomus umbellatus* was first recorded within the Unity Island site in 2019. *Butomus umbellatus* is common in the mainstem upper Niagara River, where it can grow in at least 3 m of water (Gunderson et al. 2016). It is thought that propagules recruited to the Unity Island site dispersed from the adjacent upper Niagara River, and establishment was likely aided by general site disturbance during restoration efforts. *Butomus umbellatus* continues to spread and be an aggressive invader at this site.

### Biomass

Sampling was conducted once per month at all three sites between May and November over 3 yr (2021 through 2023;  $n = 420$  samples per site). During each sampling event, 20 samples were harvested from each study site using a polyvinyl chloride (PVC) coring device designed to remove the plant and soil within the area of the device (0.018 m<sup>2</sup>) (Madsen et al. 2007; Wersal and Madsen 2018). Before sample collection, geographic coordinates, plant height, water depth, and presence of emergent leaves and inflorescences were recorded. Data loggers (HOBO pendants, Onset Computer Corporation, Bourne, MA) were deployed at two locations at each site to collect air and water temperature every 30 min throughout the sampling season each year. Plant samples were rinsed, placed into labeled plastic bags, and shipped overnight in a cooler on ice to Minnesota State University, Mankato, MN, for further processing.

In the lab, samples were washed and separated into tissue types described by Hroudová and Zákavský (1993), with aboveground tissue (leaves), rhizome and roots, inflorescences, rhizome bulbils, and vegetative bulbils formed at the base of the inflorescence (Hroudová and Zákavský 1993; Hroudová 1989). In the 3-yr span of sampling, bulbils formed at the base of the inflorescence were not recorded in any sample regardless of site. Thus, only rhizome bulbils were collected as a form of vegetative reproduction. The number of rhizome bulbils and leaves from each collected sample were also counted and recorded. Sorted tissues were put into separate paper bags and placed in a drying oven at 48 C for at least 72 h, until dry. Once dried, samples were weighed to the nearest 0.001 g then dry weight (DW) was divided by the area of the PVC coring device (0.018 m<sup>2</sup>) to determine grams of dry weight per square meter (g DW m<sup>-2</sup>) for each tissue. Densities of rhizome bulbils and ramets were calculated in a similar manner.

### Starch Allocation

Tissue samples from each site and month were consolidated into sets of five; thus, tissue samples 1 to 5, 6 to 10, 11 to 15, and 16 to 20 became starch samples 1, 2, 3, and 4 respectively. Samples were then placed in a food processor until the tissue was roughly cut. Rough-cut biomass was then ground using a Cyclone Sample Mill 3010-014 (UDY Corporation, Fort Collins, CO) and sieved through a #40 mesh screen (1 mm). After the biomass samples

were ground, 50 to 55 mg of the sample was transferred to a plastic centrifuge tube for the starch analysis. Starch percent dry weight (% DW) of each sample set was determined using the amylase/amyloglucosidase method via the commercially available STA-20 starch assay kit (Sigma Aldrich, St Louis, MO). Recovery, accuracy, and precision were assessed using several repeated measures with every starch extraction assay. Wheat (*Triticum aestivum* L.; 89% purity) and corn (*Zea mays* L.; 93% purity) starch standards were used to assess recovery, which was 90.0% ± 1.8 SE and 89.6% ± 1.4 SE for wheat and corn, respectively. The starch extraction assay accuracy was demonstrated by a five-point (dilution) standard curve that was constructed for every assay, and all absorbance points were consolidated and produced an  $r^2 = 0.98$ . Finally, the precision of the assays was determined by duplicating three *B. umbellatus* samples for every extraction and computing the percent difference between repeated samples, which on average was 10.0% ± 1.9 SE (Wersal et al. 2011).

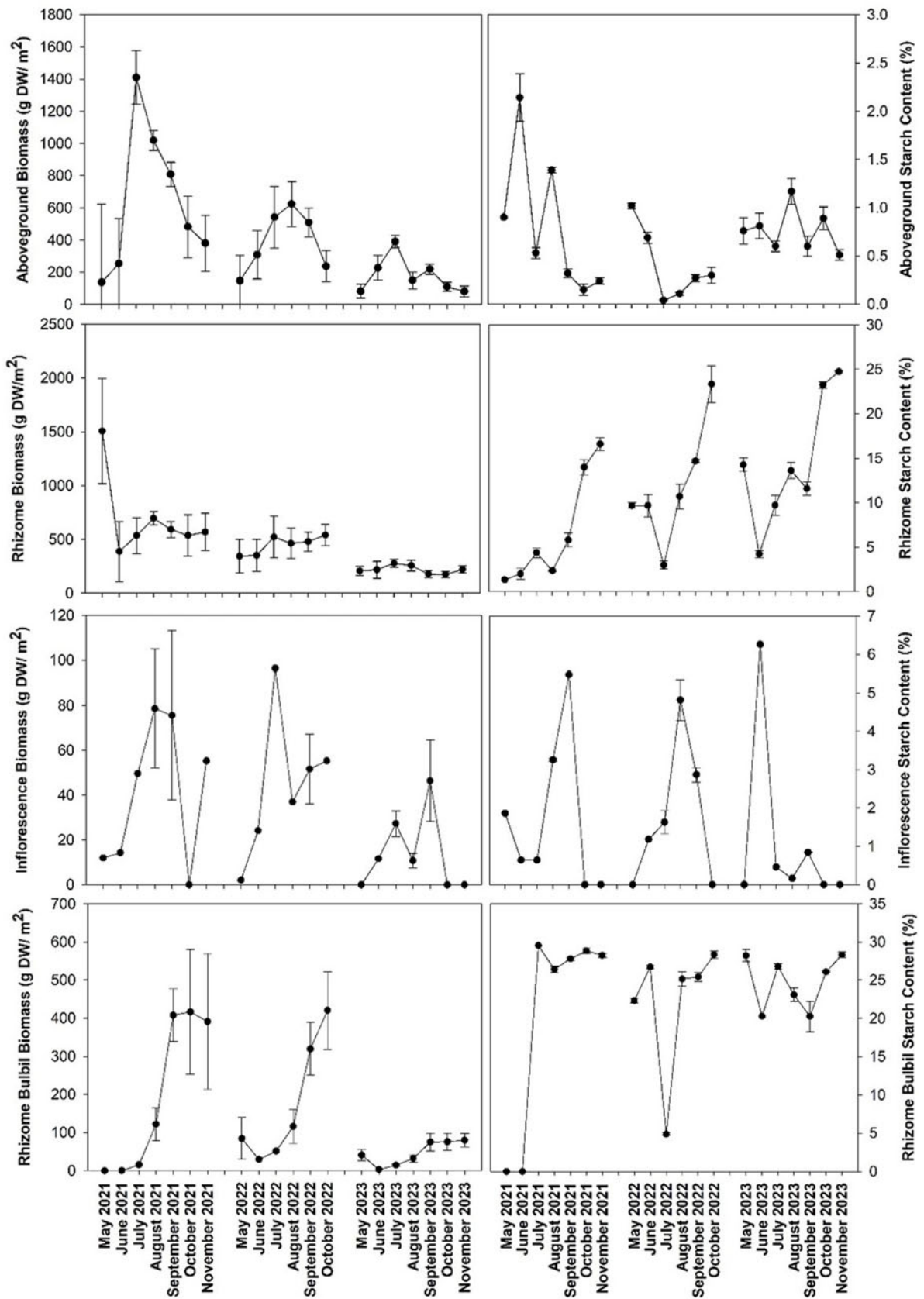
### Statistical Analysis

Monthly averages for tissue biomass, starch content, and environmental variables were calculated and pooled across study sites ( $N = 13,727$  data points). Study site was included as a random variable in the construction of the regression models to account for its influence on model variance. Due to the high variability in the data set, an average and standard deviation were calculated per biomass and starch variable each month, then all data points that were ±1 SD were removed (4,500 of 13,727 data points) (Aguinis et al. 2013; Osborne and Overbay 2019). Linear regression models were fit to determine the relationship between air temperature, water temperature, and water depth and diploid *B. umbellatus* biomass. Relationship strength was defined as no relationship (0 to 0.1), weak relationship (0.1 to 0.4), moderate relationship (0.4 to 0.6), strong relationship (0.6 to 0.9), or a perfect relationship (0.9 to 1; Dancy and Reidy 2004), using absolute values. Kruskal-Wallis analyses were conducted between sites to determine site-specific differences in rhizome bulbil density and leaf density. If a difference was observed, a Dunn's all-pairwise comparison was used to separate site-specific data. All analyses were conducted at  $\alpha = 0.05$  significance level using R statistical software v. 4.4.0 (R Core Team 2017). Packages used for analyses are as follows: DPLYR (Wickham et al. 2024), TIDYR (Wickham et al. 2023), GGPUBR (Kassambra 2023a), RSTATIX (Kassambra 2023b), LME4 (Bates et al. 2015), LMERTEST (Kuznetsova et al. 2017), and MUMIN (Bartoń 2024).

### Results and Discussion

Overall, aboveground biomass peaked from July to September and comprised 50% to 70% of peak total biomass. Peak total *B. umbellatus* biomass was 1,970.40, 2,394.07, and 2,399.28 g DW m<sup>-2</sup> for plants sampled at Mentor Marsh, Tonawanda, and Unity Island, respectively. Aboveground biomass in Mentor Marsh peaked at 1,410.18 ± 1,167.22, 624.10 ± 192.98, and 390.72 ± 49.11 g DW m<sup>-2</sup> July 2021, August 2022, and July 2023, respectively (Figure 1). Aboveground biomass at Tonawanda peaked at 1,303.04 ± 98.77, 543.64 ± 29.26, and 574.47 ± 26.94 g DW m<sup>-2</sup> in July 2021, June 2022, and July 2023, respectively (Figure 2). Aboveground biomass at Unity Island peaked at 1,006.31 ± 155.65, 1,306.55 ± 191.03, and 1,141.65 ± 61.33 g DW m<sup>-2</sup> in September 2021, September 2022, and August 2023, respectively (Figure 3). Aboveground tissues on average stored less than 2.5%





**Figure 1.** Mean ( $\pm 1$  SE) biomass and starch content in diploid *Butomus umbellatus* harvested from Mentor Marsh, OH. Scaling for y axes varies based on the collected values for each measurement associated with the axis title.

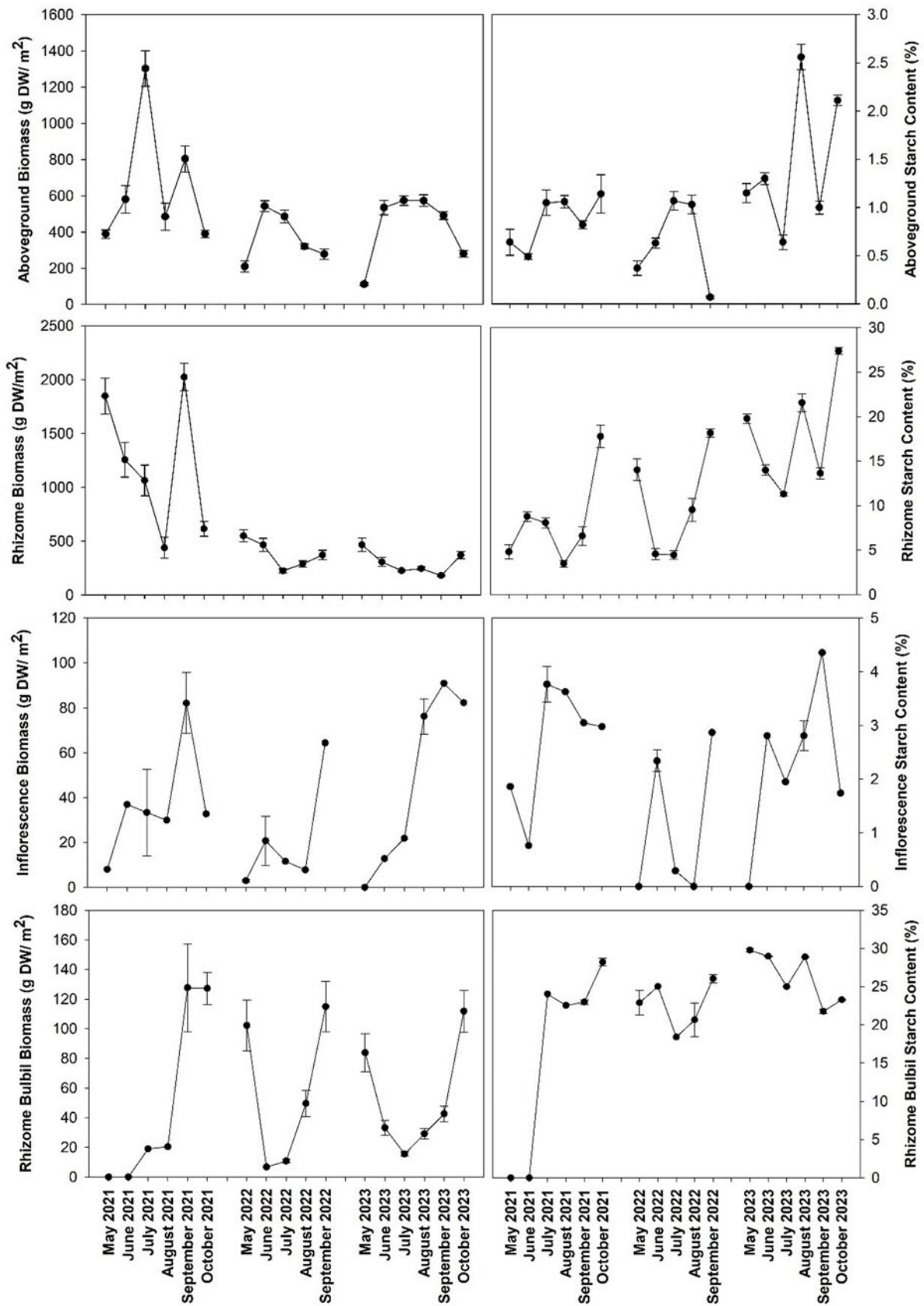
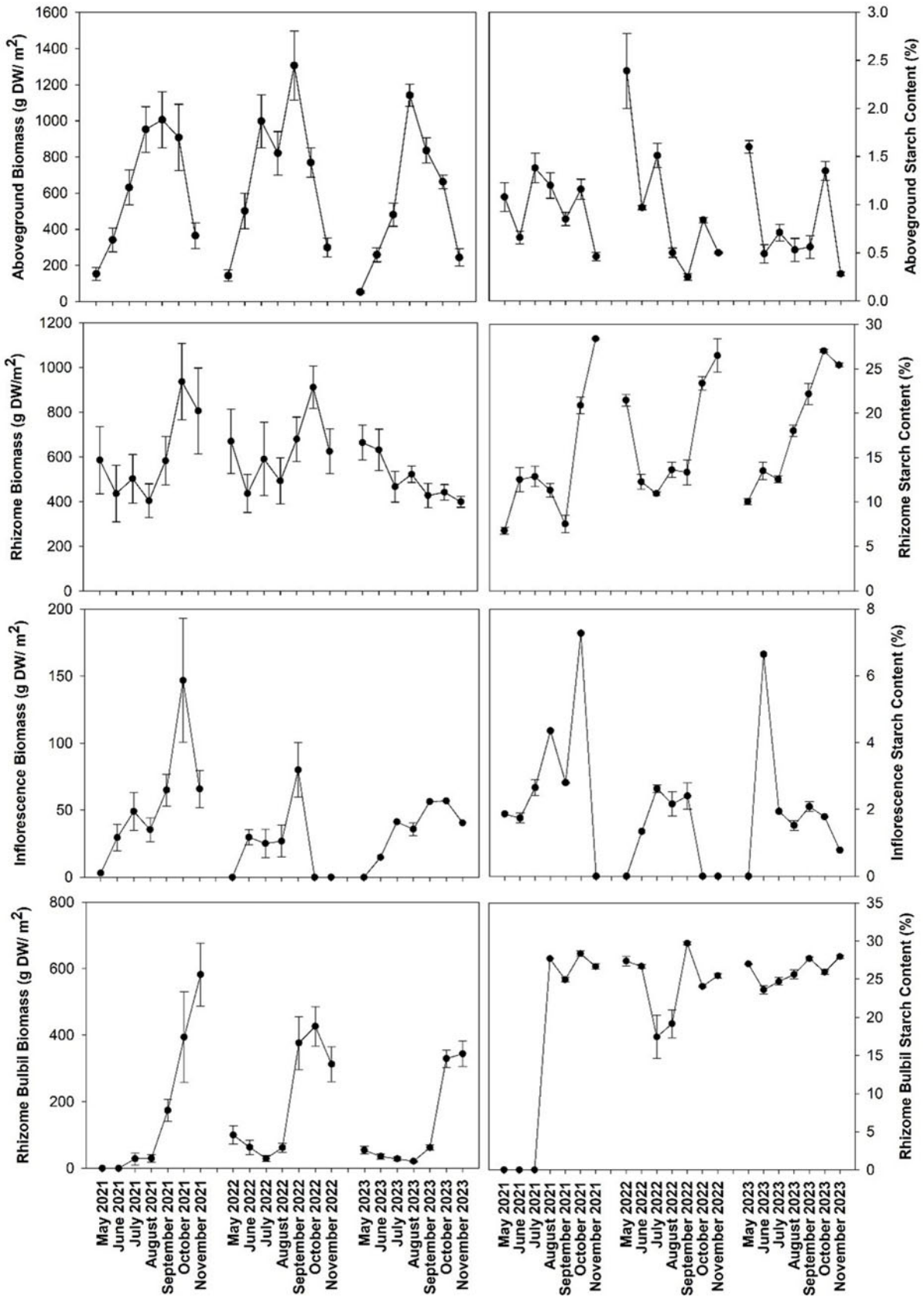
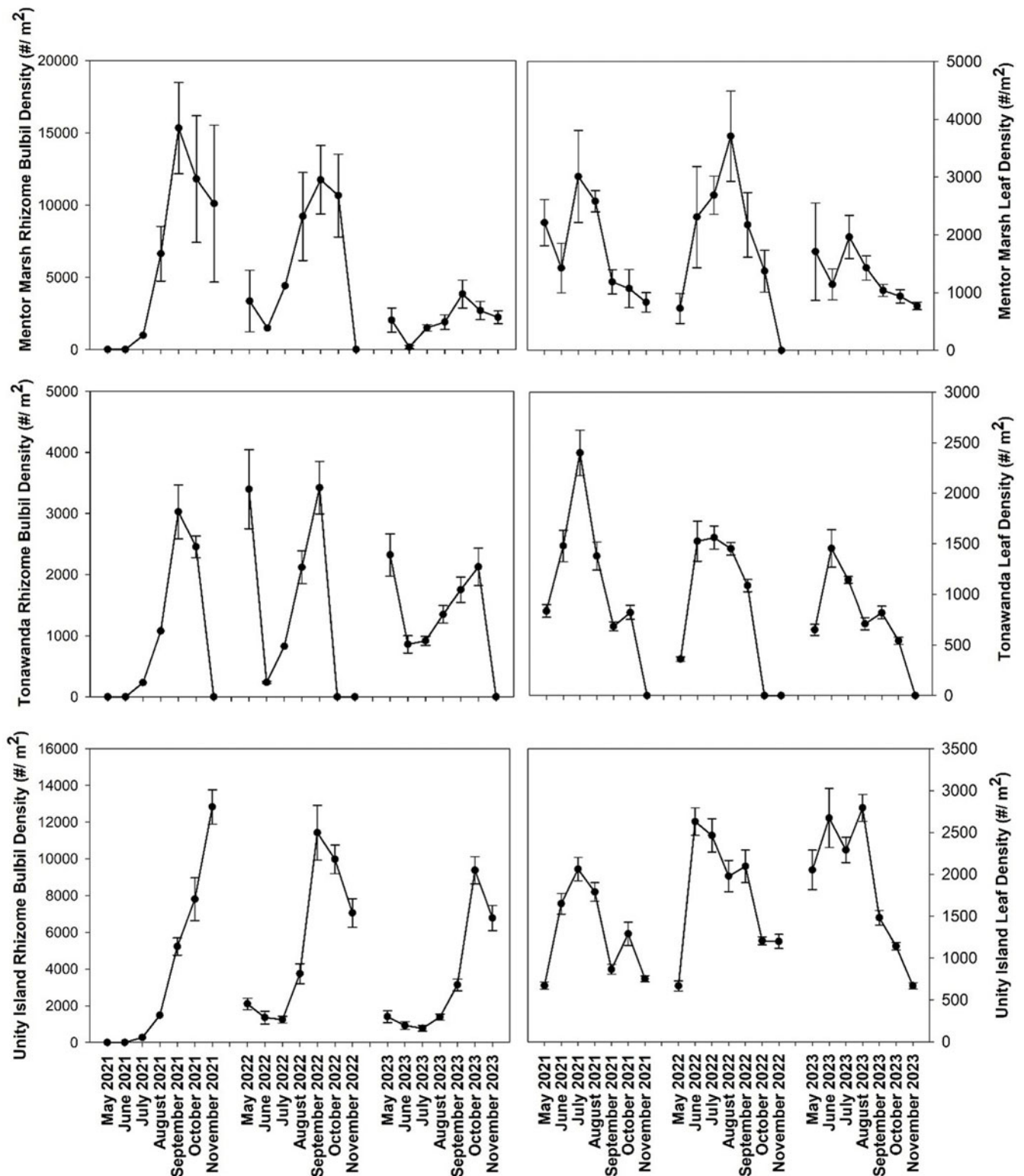


Figure 2. Mean ( $\pm 1$  SE) biomass and starch content in diploid *Butomus umbellatus* harvested from Tonawanda, NY. Scaling for y axes varies based on the collected values for each measurement associated with the axis title.



**Figure 3.** Mean ( $\pm 1$  SE) biomass and starch content in diploid *Butomus umbellatus* harvested from Unity Island, NY. Scaling for y axes varies based on the collected values for each measurement associated with the axis title.



**Figure 4.** Mean ( $\pm 1$  SE) leaf density and rhizome bulbil density in diploid *Butomus umbellatus* harvested from each study site. Scaling for y axes varies based on the collected values for each measurement associated with the axis title.

starch across all sampling sites and years, with the highest concentrations of starch occurring in May to June (Figures 1–3). Inflorescence tissue at each site was less than 10% of the total biomass and peaked between August and October. Inflorescence starch was more variable throughout the year, with low points around 0.6% and

the highest concentration at 7% (Figures 1–3). Leaf density in Mentor Marsh peaked at  $3,011 \pm 796$ ,  $3,708 \pm 780$ , and  $1,963 \pm 371$  ramets m<sup>-2</sup> in July to August for all sampling years (Figure 4). Leaf density at Tonawanda peaked at  $2,400 \pm 223$ ,  $1,560 \pm 114$ , and  $1,453 \pm 185$  ramets m<sup>-2</sup> in June to July for each sampling year (Figure 4). Leaf



**Table 1.** Results of the linear regression analyses between the plant and environmental metrics of diploid *Butomus umbellatus* pooled across all study sites.

Plant metric	Environmental metric								
	Air temperature			Water temperature			Water depth		
	P-value	Regression equation	r <sup>2</sup>	P-value	Regression equation	r <sup>2</sup>	P-value	Regression equation	r <sup>2</sup>
Aboveground biomass	<0.01	$y = 25.26x - 84.2$	0.17	<0.01	$y = 32.87x - 196$	0.19	<0.01	$y = 2.727x + 359.7$	0.03
Rhizome biomass	0.03	$y = -5.50x + 529.6$	<0.01	0.64	$y = -1.76x + 473.6$	<0.01	<0.01	$y = -5.545x + 532.2$	0.07
Inflorescence biomass	<0.01	$y = -2.101x + 80.98$	0.05	0.04	$y = -1.515x + 67.25$	0.02	0.41	$y = -0.0745x + 35.8$	<0.01
Rhizome bulbil biomass	<0.01	$y = -11.83x + 331.3$	0.18	<0.01	$y = -12.36x + 357.4$	0.11	<0.01	$y = -1.93x + 143.6$	0.05
Total biomass	0.03	$y = 9.864x + 814.8$	<0.01	<0.01	$y = 21.09x + 671.3$	0.02	0.14	$y = -2.335x + 1,062$	<0.01
Rhizome bulbil density	<0.01	$y = -137.9x + 5,939$	0.03	<0.01	$y = -161.5x + 6,862$	0.03	<0.01	$y = -38.81x + 4,029$	0.03
Leaf density	<0.01	$y = 93.45x - 457.2$	0.17	<0.01	$y = 104.3x - 656.9$	0.15	0.14	$y = 3.62x + 1,339$	<0.01
Plant height	<0.01	$y = 0.622x + 62.2$	0.01	<0.01	$y = 0.873x + 59.76$	0.02	<0.01	$y = 0.369x + 68.06$	0.06
Aboveground starch	0.08	$y = 0.007x + 0.69$	<0.01	<0.01	$y = 0.015x + 0.5582$	0.01	<0.01	$y = 0.0034x + 0.889$	0.01
Rhizome starch	<0.01	$y = -1.028x + 34.02$	0.52	<0.01	$y = -1.169x + 34.84$	0.46	<0.01	$y = -0.1064x + 15.59$	0.06
Inflorescence starch	0.02	$y = -0.0757x + 4.361$	0.02	<0.01	$y = -0.1463x + 5.96$	0.04	0.80	$y = -0.00135x + 2.677$	<0.01
Rhizome bulbil starch	<0.01	$y = -0.2709x + 30.32$	0.09	<0.01	$y = -0.241x + 29.9$	0.05	0.52	$y = -0.007x + 25.33$	<0.01

density at Unity Island peaked at  $2,061 \pm 254$ ,  $2,629 \pm 318$ , and  $2,793 \pm 271$  ramets  $m^{-2}$  in June to August for each sampling year (Figure 4). Overall, leaf density was not different ( $P = 0.93$ ) between Mentor Marsh and Unity Island, although *B. umbellatus* had fewer ( $P < 0.01$ ) leaves per square meter at Tonawanda when compared with the other sampling sites.

Peak rhizome biomass occurred September to November and comprised 40% to 50% of total biomass. Peak rhizome biomass was  $1,506.57 \pm 488.3$ ,  $2,025.24 \pm 128.82$ , and  $936.41 \pm 170.54$  g DW  $m^{-2}$  for plants sampled at Mentor Marsh, Tonawanda, and Unity Island, respectively. Rhizome biomass in Mentor Marsh peaked at  $593.27 \pm 75.44$ ,  $540.94 \pm 96.91$ , and  $222.63 \pm 33.08$  g DW  $m^{-2}$  in September 2021, October 2022, and November 2023, respectively (Figure 1). Rhizome biomass at Tonawanda peaked at  $2,025.24 \pm 128.82$ ,  $372.28 \pm 44.03$ , and  $371.65 \pm 32.78$  g DW  $m^{-2}$  in September 2021, September 2022, and October 2023, respectively (Figure 2). Rhizome biomass at Unity Island peaked at  $936.41 \pm 170.54$ ,  $911.60 \pm 94.47$ , and  $442.44 \pm 34.25$  g DW  $m^{-2}$  in October of all years (Figure 3). Rhizomes stored a large proportion of total starch (16% to 28%) across sites and years, with peak storage between October and November. Low points in rhizome starch storage occurred from May to July.

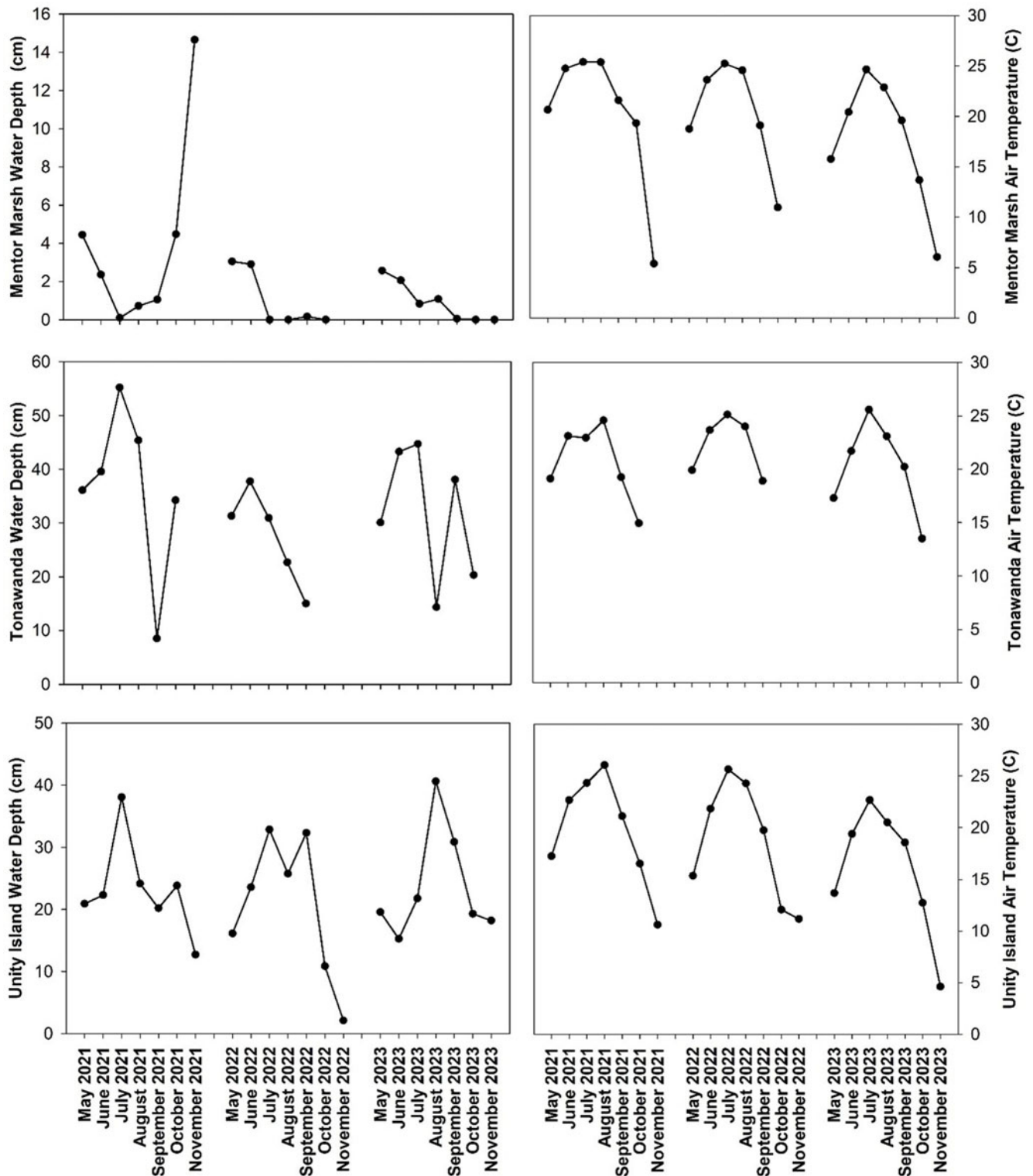
Rhizome bulbil biomass was always less than 600 g DW  $m^{-2}$  for all sites over the course of the study (Figure 4). Rhizome bulbil densities peaked in the fall from September to November at 3,000 to 16,000 rhizome bulbils  $m^{-2}$  for each sampling site. Rhizome bulbil density in Mentor Marsh peaked at  $15,333 \pm 3,153$ ,  $11,757 \pm 2,368$ , and  $3,845 \pm 967$  bulbils  $m^{-2}$ , respectively, for 2021, 2022, and 2023 (Figure 4). Rhizome bulbil density at Tonawanda peaked at  $3,026 \pm 442$ ,  $3,422 \pm 429$ , and  $2,124 \pm 306$  bulbils  $m^{-2}$ , respectively for 2021, 2022, and 2023 (Figure 4). Rhizome bulbil density at Unity Island peaked at  $12,802 \pm 2,023$ ,  $11,422 \pm 2,582$ , and  $9,379 \pm 1,490$  bulbils  $m^{-2}$ , respectively, for 2021, 2022, and 2023 (Figure 4). Rhizome bulbil densities at Mentor Marsh and Unity Island were not different ( $P = 0.34$ ); however, fewer ( $P < 0.01$ ) bulbils were produced on an annual basis at Tonawanda when compared with the other sampling sites. Even though rhizome bulbils accounted for a small proportion of total biomass, there were high amounts of stored starch that ranged from 15% to 30% throughout most of the year at each sampling site (Figures 1–3).

Rhizome starch content had strong relationships with air temperature ( $r^2 = 0.52$ ) and water temperature ( $r^2 = 0.46$ ) (Table 1). Other significant, though weak, relationships were found, including a positive relationship between aboveground biomass and air

temperature ( $r^2 = 0.17$ ), a negative relationship between rhizome bulbil biomass and air temperature ( $r^2 = 0.18$ ), and a positive relationship between leaf density and air temperature ( $r^2 = 0.17$ ). Both water and air temperature are important factors that drive photosynthesis, plant development, and phenological timing of plants (Henne et al. 2007). During this study, when water temperature approached 25 C, there was a general decrease in the production of aboveground, rhizome, and inflorescence biomass, as well as a shift in the peak rhizome starch content. Water depth has an influence on the temperature that plants will experience, in that deeper water will disperse heat energy and thus result in cooler temperatures, while shallow water will result in higher temperatures (Erwin 2009). Temperature is a notable stressor on plants, mainly due to the production of reactive oxygen species (ROS), which inhibits gas movement, resulting in potential changes of phenological patterns in the plant (Hassanuzzaman et al. 2013; Yamamoto et al. 2008). The creation of ROS causes oxidative stress in plants, which can lead to damage in the leaf and shoot tissues, thus reducing photosynthetic potentials (Marchland et al. 2005). Heat stress can occur from modest increases (1 to 2 C) in temperature. In warm, dry years, heat stress may reduce aboveground biomass and change timing of starch storage and growth of rhizomes.

Deeper water (1 m) is not ideal for *B. umbellatus* growth, as a previous study reported that water depth had a negative relationship with triploid *B. umbellatus* growth (Madsen et al. 2016). For diploid *B. umbellatus*, only plant height was related to water depth in the current study, and the relationship was positive (Table 1). Differences in growth response relating to water depth between the cytotypes may be due to the characteristics of the water bodies they were growing in. Triploid *B. umbellatus* is often found in areas with deeper and more stable hydrology (i.e., lake, reservoirs, and slow-moving rivers). In contrast, the sites sampled in this study for diploid *B. umbellatus* were shallow wetland complexes with fluctuating hydrology. Mentor Marsh had little standing water throughout the growing season, but water depth at Tonawanda WMA and Unity Island fluctuated between 10 and 60 cm (Figure 5). Differences in phenology demonstrate that *B. umbellatus* is influenced by the flooding regime that is present within an invaded wetland system (Pigliucci and Kolodynska 2002). Wetlands likely offer the most optimal environment for *B. umbellatus*, as water levels often remain below 1 m, which allows for increased biomass and production of high densities of rhizome bulbils due to receding water levels (Hroudová 1989; Madsen et al. 2016c).





**Figure 5.** Water depth (cm) and mean air temperature (C) for study sites where diploid *Butomus umbellatus* samples were harvested. Scaling for y axes varies based on the collected values for each measurement associated with the axis title.

Diploid *B. umbellatus* can reproduce and spread vegetatively via rhizome fragmentation, rhizome bulbils, and inflorescence bulbils formed at the base of the inflorescence, although there were no inflorescence bulbils observed during this study. Diploid *B. umbellatus* has the potential to produce >15,000 rhizome bulbils m<sup>-2</sup>. In contrast, triploid *B. umbellatus* produces fewer but

larger reproductive structures (600 rhizome buds m<sup>-2</sup>) (Marko et al. 2015). Diploid *B. umbellatus* also reproduces sexually through flowering and seed production. *Butomus umbellatus* can produce 20 to 50 flowers per inflorescence and approximately 200 seeds per flower, with each seed having roughly 31% viability (Eckert et al. 2000). Individual plants can produce multiple inflorescences, thus

increasing the amount of seeds produced per square meter as well. Considering both reproductive strategies, diploid *B. umbellatus* could produce up to 18,000 individuals  $m^{-2}$  (15,000 rhizome bulbils  $m^{-2}$  with 3,000 viable seeds per inflorescence) during the reproductive season (fall) under optimal conditions, which is almost 30-fold higher than triploid *B. umbellatus*.

Overall, many of the life-history traits between triploid and diploid *B. umbellatus* are similar. Rhizome biomass and peak periods are similar (Marko et al. 2015). Aboveground biomass in triploid biomass reached a peak of just over 500 g DW  $m^{-2}$ , with diploid *B. umbellatus* producing between 600 and 1,100 g DW  $m^{-2}$  (Marko et al. 2015). Both cytotypes had aboveground biomass peak between July and August, with the rhizome and rhizome bulbils reaching peak production October to November. Management strategies for both cytotypes should be similar and focus on two goals: (1) long-term biomass reduction and (2) reduction in vegetative propagules. *Butomus umbellatus* management is challenging, as rhizomes and rhizome bulbils combined store up to 60% of total starch and present a unique challenge to management, as these structures cannot be reached directly with herbicides (Parsons et al. 2019; Turnage et al. 2017). However, rhizome biomass and starch content decrease as aboveground biomass increases in June and July. Management programs should target aboveground biomass before July to remove new growth and further deplete starch content in the rhizome tissue. Additional herbicide applications should be utilized, as plants can recover, which ultimately exhausts starch reserves and limits rhizome bulbil formation (Bellaud 2009; Erwin 2009). Repeat applications should be sufficient to reduce year-to-year recruitment, although management of *B. umbellatus* will be long term, as the propagule bank will need to be exhausted to completely manage this species. Sites invaded by diploid *B. umbellatus* may see slightly different trends in production, with those sites further south potentially experiencing earlier biomass peaks. Monitoring populations closely would provide important information for when management should occur.

In addition to site-specific management, large-scale prevention programs should be developed to identify water bodies that have a high probability of invasion. Diploid *B. umbellatus* possesses some differences to its triploid counterpart due to the differences in reproductive output and environmental response (Banerjee et al. 2020; Gebhart and Wersal 2023). There is also evidence of differential diploid *B. umbellatus* growth based on site-specific characteristics. These site-specific characteristics should suggest that populations of *B. umbellatus* may need a particular approach when aiming for management and control. High reproductive output, especially through vegetative bulbils, increases the likelihood of diploid *B. umbellatus* spread, especially through aquatic systems that are associated with rivers. Based on the current distribution in the Great Lakes region, diploid *B. umbellatus* has access to the largest lakes and numerous river systems in North America that can move seeds, rhizome bulbils, and rhizome fragments. With the ability to produce almost 18,000 individuals  $m^{-2}$  during peak production, diploid *B. umbellatus* is an invasive plant cytotype that poses a high potential to expand to most of North America. However, a differential response to environmental conditions may not allow diploid *B. umbellatus* to expand westward due to increased temperatures seen in the western United States (Levin 1983). Further temperature and phenology studies could be conducted to create a coherent profile for diploid *B. umbellatus* reproduction and life-history strategies within the United States.

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