




RESEARCH ARTICLE

Resistance of sugarcane hybrids to internode borer *Chilo sacchariphagus indicus* (Lepidoptera: Crambidae)

P. Mahesh¹ , B. Singaravelu¹, J. Srikanth¹, K. P. Salin¹, K. Chandran² and R. Nirmala¹

¹Section of Entomology, ICAR-Sugarcane Breeding Institute, Coimbatore 641007, Tamil Nadu State, India and

²ICAR-Sugarcane Breeding Institute Research Center, Kannur 670002, Kerala State, India

Corresponding author: P. Mahesh; Email: agrimahesh@gmail.com

(Received 07 August 2023; revised 29 December 2023; accepted 30 January 2024)

ABSTRACT

A four-year field study (2013–2016) was conducted to screen Indian sugarcane hybrids together with two susceptible checks against internode borer *Chilo sacchariphagus indicus* (Kapur) (Lepidoptera: Crambidae) in endemic locations of Tamil Nadu State, India. Each year, borer incidence on cane basis and intensity on internode basis were assessed at harvest to eliminate susceptible entries. Of the total 535 hybrids screened, only Co 293 emerged as resistant at the end of fourth year trial which was confirmed in tests under controlled conditions with artificial infestation. A modified relative resistance ratio computed using incidence and intensity also confirmed its resistance to the borer. In laboratory oviposition choice tests with excised leaves of the resistant Co 293 and susceptible hybrids Co 86032 and Co 1060, percent of leaf bits oviposited, egg masses laid, and egg numbers deposited were significantly lowest in Co 293. Also, an oviposition preference index computed for both egg mass number and egg number was significantly lowest for Co 293 which suggested antixenosis. Larval survival was significantly lowest in Co 293 with 5 to 10-fold higher neonate mortality than in the two susceptible hybrids. Prolonged larval development period and lower fecundity were observed when the borer was reared on Co 293 which indicated antibiosis. A relative suitability ratio developed from larval and pupal durations also indicated lower suitability of Co 293. Among the plant morphological characters examined, leaf length and cane thickness positively influenced borer incidence; loose sheath-clasp was associated with higher borer incidence. Among 12 shoot phenolics quantified, eight were present in higher quantities in Co 293 suggesting their role in antibiosis. Co 293 identified as resistant hybrid in the present study has the potential to be used as a parent in breeding programs for *C. sacchariphagus indicus* resistance.

Keywords: Sugarcane; *Chilo sacchariphagus indicus*; Indian hybrids; relative resistance; oviposition preference; relative suitability; antixenosis; antibiosis; phenolics

Introduction

Worldwide, moth borers are among the most damaging pests of sugarcane (*Saccharum* spp. hybrids) with about 50 species, mostly belonging to the family Crambidae, reported damaging sugarcane stalks (Long and Hensley, 1972). *Eldana saccharina* (Walker) (Lepidoptera: Pyralidae) and *Diatraea saccharalis* F. (Lepidoptera: Crambidae) are major pests in Africa and the American continent, respectively (Showler and Reagan, 2012). Tunneling of stalks by larvae generally leads to deadheart formation and side shooting, disruption of nutrient flow, stalk breakage, loss of stalk weight, and introduction of pathogens, all of which contribute to reduced yield and juice quality (Long and Hensley, 1972; Milligan *et al.*, 2003; Reay-Jones *et al.*, 2005; White *et al.*, 2008; Showler and Reagan, 2017; Reagan and Mulcahy, 2019). The genus *Chilo* spp. is distributed throughout Asia and Africa with *Chilo sacchariphagus* (Lepidoptera: Crambidae) being the major pest in



Figure 1. *Chilo sacchariphagus indicus* damage in sugarcane: (a) dead heart, (b) bore hole, (c) bunchy top, and (d) grown-up larva in leaf sheath.

China, India, Indonesia, Madagascar, Mauritius, Taiwan, Reunion and the Comoros, Borneo, Java, Bali, Sumatra, Celebes, Japan, Singapore, Sri Lanka, Malaysia, Thailand, and the Philippines (Sallam and Allsopp, 2003) with known yield losses of up to 40 t/ha (Goebel *et al.*, 1999).

In India, more than 14 species of moth borers are reported to attack sugarcane in both tropical and sub-tropical sugarcane-producing provinces (David and Nandagopal, 1986). Among these, the internode borer *Chilo sacchariphagus indicus* (Kapur) (Lepidoptera: Crambidae), a subspecies of *C. sacchariphagus* (David, 1986; Sallam and Allsopp, 2003), is the most destructive borer pest in peninsular India (Srikanth and Kurup, 2011; Mahesh *et al.*, 2018). It generally attacks the crop from internode formation to harvest causing estimated yield losses of 10–35% (Yalawar *et al.*, 2010). Injury extending to three or more internodes or more than 10% length of cane can produce significant deterioration in juice quality (David, 1986). Destruction of apical meristem in young and weak shoots results in the production of deadhearts, axillary bud sprouting, and bunchy top formation (Figure 1) leading to greater crop losses (Mukunthan and Rakkiyappan, 1989; Srikanth and Kurup, 2011). Insecticides are not the best option for the control of *C. sacchariphagus indicus* due to the concealed habit of larvae and unamenable crop canopy (Ananthanarayana and David, 1986). Biological control and sex pheromone traps are known to minimize borer incidence levels but suffer from limitations (Mahesh *et al.*, 2018). In this context, cultivar resistance, a major component of long-term integrated pest management strategy for sugarcane moths in many sugarcane growing regions of the world (Reay-Jones *et al.*, 2005; Mahesh *et al.*, 2018; Wilson *et al.*, 2021), can serve as a potential tool to reduce losses caused by borers.

Screening of sugarcane cultivars against *D. saccharalis* in Louisiana, USA (Bessin *et al.*, 1990; Posey *et al.*, 2006; Reagan *et al.*, 2008), and Brazil (Tomaz *et al.*, 2018), *Eoreuma loftini* (Dyar) (Lepidoptera: Crambidae) in Texas (Reay-Jones *et al.*, 2003; Wilson *et al.*, 2015; Salgado *et al.*, 2022a), *E. saccharina* in South Africa (Keeping, 2006), and other moth borers in Papua New Guinea (Korowi and Samson, 2013) have demonstrated significant differences in the resistance of cultivars to various borers. Commercial sugarcane cultivars display a range of resistance with susceptible ones often suffering 5–10 fold greater levels of injury than resistant cultivars (Salgado *et al.*, 2022b). The Research Center of ICAR-Sugarcane Breeding Institute (ICAR-SBIRC) located at Kannur, Kerala State, India, currently maintains the world's largest collection of sugarcane germplasm. This vast germplasm of diverse genetic makeup includes assemblages of Indian hybrids, foreign hybrids, *Saccharum* spp., and allied genera, constituting a crop island (Mahesh *et al.*, 2019). In earlier work, attempts were made to identify sources of resistance among limited accessions of five *Saccharum* spp., namely *S. barberi*, *S. robustum*, *S. sinense*, *S. spontaneum*, and *S. officinarum* available at the Research Center, against *C. sacchariphagus indicus* under field conditions (Jayanthi, 1988; Mahesh *et al.*, 2018). However, resistance among hybrids to this borer has not been examined seriously hitherto except for the screening of a few commercial hybrids (Jhansi and Rao, 1996; Rajendran *et al.*, 1996; Albert *et al.*, 2007) and prerelease clones (Jayakumar, 2018; Pasupathy *et al.*, 2021). Hence, the present study was undertaken to investigate the resistance of a large collection of Indian hybrids to *C. sacchariphagus indicus* in a series of field screening trials followed by the assessment of selected resistant hybrids in pot culture. Besides, oviposition preference, relative survival and comparative biology of the borer were examined in the laboratory to determine the underlying resistance mechanisms and enable identification of genetic stock for introgression of borer resistance in sugarcane breeding programs.

Materials and methods

Field screening of Indian hybrids

Study site and field layout

A series of four field experiments was conducted to screen Indian hybrids against *C. sacchariphagus indicus* under natural pest pressure at ICAR-SBIRC, Kannur, Kerala State, India, and research farm of M/s Rajshree Sugars and Chemicals Ltd (RSCL), Mundiampakkam, Villupuram district, Tamil Nadu State, India, from 2013 to 2016 (Supplementary Material Table S1). The hybrids were screened adopting a process of elimination of those found susceptible in any one replication in successive years of screening (Mukunthan, 2001). In the first year trial (2013), hybrids showing above 30% incidence level were excluded whereas in the subsequent trials (2014–2016), hybrids showing more than 15% incidence were eliminated as a more rigorous method of screening.

In the first field trial (2013) at ICAR-SBIRC, 535 hybrids (Table S2) were planted in 2 m rows each with a row-to-row spacing of 90 cm. In the subsequent trials (2014–2016) at M/s Rajshree Sugars and Chemicals Ltd., an area endemic for *C. sacchariphagus indicus* (Puthira Prathap *et al.*, 2014), hybrids found resistant in the previous trial were carried forward and screened against the borer in replicated randomized block design. In the 2014 trial, 187 test hybrids, including the two susceptible hybrids, namely Co 86032 (Nalawade *et al.*, 2022) and Co 1060 (Table S1), were planted in four blocks serving as replications, each containing two rows of all entries randomized within the block. In the 2015 trial, 50 hybrids and the two susceptible checks 86032 and Co 1060 were planted in five blocks with two rows for each entry. In the 2016 trial, four hybrids and the two susceptible checks Co 86032 and Co 1060 were planted in six blocks with six rows for each plot. All test entries were planted in 6 m rows with 1.8 m spacing between rows and 1.2 m alley between plots. In all the trials, planting was done in January following recommended agronomical practices (Sundara, 1998) but without pesticide application.

Damage assessment

The attack of *C. sacchariphagus indicus* begins at fifth month age of the crop and extends almost until harvest. Neonate larvae, after feeding on the inside of leaf sheath tissue for 7–8 days, typically bore into the top internodes egesting out wet frass from the bore holes which indicates fresh attack; the presence of large bore holes, often blackened by the growth of saprophytes, in the lower internodes (Figure 1) characterizes old attack (Mahesh *et al.*, 2018). At harvest, millable canes in all rows of each replication were counted, cut at the ground level, examined for the presence of injury symptoms, and the number of canes with bore holes was recorded. In the injured canes, total number of internodes and number of internodes with bore holes were recorded and the following three injury parameters were computed.

$$\text{Percent incidence} = \frac{\text{Number of bored canes}}{\text{Total number of canes}} \times 100 \quad (1)$$

$$\text{Percent intensity} = \frac{\text{Number of bored internodes}}{\text{Total number of internodes}} \times 100 \quad (2)$$

$$\text{Infestation index} = \frac{\text{Percent incidence} \times \text{Percent intensity}}{100} \quad (3)$$

While percent incidence indicates between-plant dispersal of the borer, percent intensity signifies within-plant spread and repeated attack by successive generations of the borer. Infestation index accommodates variation in both types of attack and may serve as a composite indicator, and in many cases, the index reflects percent incidence (Mahesh *et al.*, 2018).

Screening under artificial infestation

Internode borer culture

Chilo sacchariphagus indicus required for artificial infestation studies was obtained from a laboratory culture maintained at the Section of Entomology, ICAR-Sugarcane Breeding Institute, Coimbatore, Tamil Nadu, India. The culture originated from borer larvae collected in sugarcane fields which were reared on shoot bits in the laboratory until pupation. After determining the sex of pupae (Duttamajumder, 2020), equal numbers of male and female pupae (10 : 10) were placed in oviposition cages (45 cm L x 45 cm W x 45 cm ht) provisioned with 20 cm leaf bits of the susceptible Co 86032 as substrate for oviposition. Egg masses laid on leaves were surface sterilized with 1% formaldehyde at black head stage and emerging neonate larvae were reared in artificial diet in 10 cm ht x 2.5 cm dia. glass tubes at 25 °C and 65% RH (Mehta and David, 1978; Anonymous, 2018).

Artificial inoculation

After eliminating a large number of susceptible entries under natural conditions of infestation in the 4-year field-screening process, four apparently resistant hybrids, namely Co 293, Co 389, Co 62019, and Co 62213, together with the two susceptible hybrids Co 86032 and Co 1060, were subjected to artificial infestation in pot culture under greenhouse conditions. Single-budded sugarcane setts from 7-month-old crop of the six hybrids were planted in 45 cm diameter pots filled with a 1:1:1 mixture of sand, silt, and cattle farmyard manure (0.5% N - 0.2% P₂O₅ - 0.5% K₂O). Each hybrid was planted in six pots serving as six replications, and a uniform plant stand of six canes was maintained in each pot.

When plants attained 6 months age, neonate larvae of *C. sacchariphagus indicus* obtained from laboratory culture were released on the inner side of leaf sheath of +4 or +5 leaf (Clements and Ghotb, 1969) at 10 per plant using a fine camel hair brush. A month after inoculation, the entries were evaluated based on the appearance of bore holes or deadhearts.

Categorization of hybrids

Percent incidence, percent intensity and infestation index were considered to assess borer damage as described above. However, in view of its predominance in the computation of infestation index, only percent incidence was used as a rapid and low sampling-cost parameter to group hybrids into three categories of resistance (Table S3) (Mukunthan, 2001; Mahesh *et al.*, 2018) under both natural and controlled conditions.

Relative resistant ratio

A modified relative resistance ratio (RRR) was computed for four test hybrids and two susceptible hybrids following Wilson *et al.*, (2015). In the field trial and the test conducted under controlled conditions, hybrids within each replication were ranked for percent of incidence and percent of intensity and RRR was computed as

$$\text{Relative Resistance Ratio} = \frac{RR_{(\text{Inc.})} + RR_{(\text{Int.})}}{2n} \quad (4)$$

where:

n : number of hybrids evaluated

RR_(Inc.): relative resistance ranking based on percent of incidence (the lowest percent has an RR_(Inc.)=1 and the highest has an RR_(Inc.)= n)

RR_(Int.): relative resistance ranking based on percent of intensity (the lowest percent has an RR_(Int.)=1 and the highest has an RR_(Int.)= n)

Oviposition choice tests

Oviposition preference of *C. sacchariphagus indicus* was examined with Co 293, the hybrid found to be resistant in the field and moderately resistant under controlled conditions, and the susceptible hybrids Co 1060 and Co 86032. The hybrids were maintained in micro-plots following normal agronomic practices (Sundara, 1998) and +4 or +5 leaves (Clements and Ghotb, 1969) excised from 5-month-old plants were used in oviposition choice tests. The tests were conducted in plastic containers (20 cm ht x 18 cm dia.) with wire mesh lid serving as oviposition cages. Since the borer was found to prefer middle part of +4 or +5 leaves for oviposition in the wild (Anonymous, 2018), five 17 cm long leaf bits cut from the center of excised leaves of each hybrid were placed vertically by inserting them in moist sand placed in a small plastic cup. The three cups containing leaf bits of each hybrid were placed equidistant from one another in the oviposition cage and freshly eclosed (<24 h) adults (1 male: 1 female) of *C. sacchariphagus indicus* selected from the laboratory culture were released in each cage. The test was replicated 22 times with oviposition cages maintained under ambient laboratory conditions (25 °C and 65% RH). Leaf bits were replaced with fresh ones every 24 h after examining for oviposition. The number of leaf bits oviposited upon, and the number of egg masses and eggs on each leaf bit of the three hybrids were recorded every day until the death of female in about 7 d. From the cumulative data compiled over the observation period for the three hybrids, percent of leaf bits oviposited, and percent of egg masses and egg numbers deposited were calculated for the 22 replications. Besides, egg number in individual egg masses laid in all three hybrids was also assessed.

Oviposition preference index (OPI)

An oviposition preference index (OPI) on per leaf bit basis, per cm² area of leaf bit, and per gram weight of leaf bit for egg mass number and egg number was derived using the equation of Reay-Jones *et al.*, (2007). For each hybrid, area of leaf bit was calculated as the product of length (17 cm) and broadest width measured from four samples and mean area computed; mean weight of leaf bit was computed from seven samples. OPI was computed as

$$\alpha_{ij} = \frac{n_{ij}}{\max n_j} \quad (5)$$

where

α_{ij} = estimated preference shown for the *i*th hybrid for the *j*th dataset

n_{ij} = mean number of egg masses or eggs laid per unit area or unit weight of leaf bit on the *i*th treatment and *j*th dataset

$\max n_j$ = mean maximum number of egg masses or eggs laid per unit area or unit weight of leaf bit across treatments for the *j*th dataset

Larval development and survival

After screening under artificial inoculation in pot culture, the resistant Co 293 and the susceptible Co 1060 and Co 86032 hybrids were assessed for borer development and survival in the laboratory under ambient conditions (25 °C, 65% RH and 12L:12D). Neonate larvae obtained from stock culture were placed in rearing boxes (8.5 cm ht x 6.0 cm dia.) lined with moist filter paper at the bottom. Five larvae were released per box and provisioned with three 5.0 cm long tender shoot bits obtained from 5-month old plants of the three hybrids maintained in a micro-plot. Fifteen boxes were maintained for each hybrid as replicates in a completely randomized design. Shoot bits were changed and larval mortality per box was recorded on alternate days. The experiment was continued until death of larvae or pupation (\approx 6 weeks), and cumulative weekly mortality rates were computed.

Comparative biology on resistant and susceptible hybrids

Life cycle in shoots

Life cycle of the borer was examined in shoots of the field-resistant (Co 293) and field-susceptible hybrids (Co1060 and Co 86032). Five neonate larvae obtained from stock culture were released in each of 10 rearing boxes and reared until pupation as described above. Freshly formed pupae were sexed, weighed, and kept individually in plastic boxes lined with moist filter paper at the bottom for adult emergence. Moths were maintained on 50% honey solution dispensed on cotton wad until death. Total larval duration, pupal period, pupal weight, sex ratio, and adult longevity were recorded for larvae that completed development which were used as replications ($n=17-22$) for analysis. Emerging males and females were held in pairs in oviposition cages until female death and fecundity was recorded as described under oviposition choice tests above.

Relative suitability ratio

A relative suitability ratio (RSR) was developed on the lines of RRR (Wilson *et al.*, 2015) using larval and pupal durations obtained from replicated laboratory rearing on shoots of the three hybrids ($n=17$). Since developmental durations are generally prolonged on unsuitable hosts, longest developmental period was assigned the lowest rank and shortest developmental period was given the highest rank; in case of equal developmental periods, mean ranking was assigned. Consequently, lower RSR indicates unsuitability of the host and vice versa.

$$\text{Relative Suitability Ratio} = \frac{RS_{(\text{Larva})} + RS_{(\text{Pupa})}}{2n} \quad (6)$$

where:

n : number of hybrids evaluated

$RS_{(\text{Larva})}$: relative suitability ranking based on larval period (the highest duration has an

$RS_{(\text{Larva})} = 1$ and the lowest has an $RS_{(\text{Larva})} = n$)

$RS_{(Pupa)}$: relative suitability ranking based on pupal period (the highest period has an $RS_{(Pupa)} = 1$ and the lowest has an $RS_{(Pupa)} = n$)

Morphological characters vs. borer incidence

To establish the relationship between cane morphological characters and borer injury parameters, selected quantitative and qualitative morphological traits were recorded at 9 months age in healthy canes of hybrids maintained at ICAR-SBIRC, Kannur. Leaf characters were measured on third leaf from visible dewlap from each of three randomly selected plants and the means calculated. Leaf length (cm) was measured from ligule to leaf tip, and leaf width (cm) was measured in the middle part of leaf. Cane parameters were measured on three randomly selected canes to compute means. While cane length (cm) was measured from ground level to the top-breaking point, cane thickness (cm) was calculated as the mean of three measurements recorded in top, middle, and bottom parts of the cane using a digital vernier caliper. Internode hardness was measured in top, middle and bottom portions of the cane with a 'rind hardness tester' developed at ICAR-Sugarcane Breeding Institute, Coimbatore, India (Babu *et al.*, 2009; Govindaraj *et al.*, 2014); fiber content was estimated gravimetrically. Yield traits, namely single cane weight (kg), brix and sucrose percentage, were recorded at cane maturity, i.e., 12 months age. Brix was recorded in the middle portion of three independent canes using a hand refractometer (ATAGO, Japan).

Besides, qualitative plant characters such as rind color (14 classes), pith (three classes), sheath-clasp (four classes), and leaf hairiness (two classes) were recorded at 9 months age as per the descriptor classes and descriptor states adopted for sugarcane germplasm (Artschwager and Brandes, 1958).

Estimation of phenolics

Sample collection and preparation

Phenolics in shoots of the three hybrids Co 293, Co 1060, and Co 86032 were estimated (Tonnesen and Karlsen, 1983) at 6 months age. Top shoots were collected from three uninfested canes for each hybrid maintained in the field and processed immediately. Shoots were chopped finely, macerated in liquid nitrogen, extracted with methanol, and filtered through a filter paper (47 mm, Whatmann, UK) to remove residues and pigments. The extract was stored at -20°C until further analysis in HPLC.

HPLC analysis

Shoot phenols were estimated in Shimadzu LC-8A HPLC system with C18 column (Phenomenex Gemini 250 x 4.6 mm; 5 μm particle size) and photodiode array detector. Methanol extracts were filtered through 0.2 μm membrane, and 25 μl of sample was injected using Rheodyne injector. A gradient mixture of methanol and 0.1% phosphoric acid (in water) as the mobile phase at a flow rate of 1 ml/min was maintained; the detection was performed at 285 nm. The ratio of methanol : 0.1% phosphoric acid in water was increased from 0 : 100 to 5 : 95 in the first 5 min, 10 : 90 in the next 5 min, 15 : 85 in the next 15 min, 40 : 60 in the subsequent 20 min, and 95 : 5 in the next 10 min. The ratio was then brought down to the initial level of 5 : 95 in the last 10 min. Serial dilutions of standards (Sigma-Aldrich, USA) of the phenolic compounds were prepared in methanol and used to obtain standard curves. Concentrations (ppm) of individual phenolics in the samples were estimated by comparing the retention time and peak area of sample chromatograms with those of standards.

Statistical analysis

Repeated measures analysis of variance (ANOVA) was applied to field data (2014–2016) with six hybrids as the levels of between-group factor and three study years as the levels of within-subject repeated measure factor. One-way ANOVA was used to analyze data from the pot culture experiment with six hybrids under controlled conditions, laboratory oviposition preference test with three hybrids under free-choice situation, fecundity data of females reared on shoots, RRR and RSR; data were subjected to square root or arcsin transformation wherever needed. Repeated measures ANOVA was used for arcsin-transformed mortality rates of the borer reared on three hybrids with hybrids as the levels of between-group factor and cumulative mortality in different weeks after setup as the levels of within-subject repeated measure factor. Two-way factorial ANOVA was applied to laboratory life cycle parameters with hybrid (three) and sex (male and female) serving as factors. Tukey HSD test for equal or unequal replications was employed for multiple comparisons of treatments in all ANOVA. Simple correlation coefficients were computed between infestation parameters and quantitative plant morphological characters of different hybrids. To assess the effect of qualitative plant characters on infestation parameters, Mann–Whitney U test was applied for characters with two categories, and Kruskal–Wallis ANOVA by ranks and multiple comparison tests were used for characters with more than two categories. Bar diagrams were used for graphical depiction and mean comparison of field attack rates, RRR, OPI on leaf basis, fecundity, and RSR. The analyses were performed using StatSoft, Inc. (2004).

Results

Chilo sacchariphagus indicus infestation levels among hybrids

Screening of 535 hybrids through the process of elimination of susceptible hybrids in four successive years (2013–2016) led to the selection of four hybrids, namely Co 293, Co 389, Co 62019 and Co 62213 by the end of 2016. In the first trial (2013) at ICAR-SBIRC, Kannur, attack rates on cane basis (percent incidence) were lower in these four hybrids (4.55–15.15%) than in the susceptible checks, i.e., Co 86032 (29.17%) and Co 1060 (30.43%) (Table S4). However, attack rates on internode basis (percent intensity) (3.85–11.76%) did not show clear-cut trend among the hybrids.

In the three trials (2014–2016) conducted at RSCL, Mundiampakkam, injury levels were generally higher (Table 1) than those observed in 2013 at Kannur (Table S4). Besides, injury levels varied widely among the three years with percent incidence and infestation index showing far higher values in 2016. Repeated measures ANOVA of percent incidence was significant for hybrids ($F_{5, 12}=60.89$; $p < 0.01$) and years ($F_{2, 24}=51.75$; $p < 0.01$) but not their interaction ($F_{10, 24}=2.25$; $p > 0.05$). Mean percent incidence was significantly lowest in Co 293 (10.04 ± 2.56) and highest in the standards Co 1060 (65.59 ± 2.56) and Co 86032 (46.30 ± 2.56); the differences were not significant among the remaining three hybrids. Among the years, the mean percent incidence did not differ between 2014 and 2015 but was significantly highest in 2016. Repeated measures ANOVA of percent intensity was not significant for hybrids ($F_{5, 12}=2.22$; $p > 0.05$) but significant for years ($F_{2, 24}=17.41$; $p < 0.01$) and their interaction ($F_{10, 24}=3.52$; $p < 0.01$); it was significantly lower in 2015. With repeated measures ANOVA of infestation index being significant for hybrids ($F_{5, 12}=46.07$; $p < 0.01$), years ($F_{2, 24}=91.18$; $p < 0.01$), and their interaction ($F_{10, 24}=4.79$; $p < 0.01$), the parameter followed the same trend as percent incidence showing a slight decrease in 2015.

Screening of hybrids for resistance

Under natural conditions

Of the 535 Indian hybrids screened for their relative degree of resistance in the field, four hybrids remained resistant for three consecutive years. In the final confirmatory screening studies (2016), when the four resistant hybrids were evaluated along with two susceptible hybrids, only one

Table 1. Comparative levels of injury by *Chilo sacchariphagus indicus* among six sugarcane hybrids screened in the field at Mundiampakkam, Tamil Nadu State, India, during 2014–2016

| Hybrid | Year of evaluation | | | Mean |
|------------------------|-----------------------|---------------|--------------|-----------------------|
| | 2014 | 2015 | 2016 | |
| Percent of infestation | | | | |
| Co 62019 | 14.48 ± 0.68 | 14.49 ± 1.57 | 52.24 ± 4.73 | 27.07 a ^{@1} |
| Co 62213 | 14.60 ± 0.63 | 12.03 ± 2.23 | 46.97 ± 9.17 | 24.53 a |
| Co 389 | 14.87 ± 0.30 | 12.17 ± 2.74 | 55.93 ± 8.67 | 27.66 a |
| Co 293 | 8.41 ± 2.30 | 7.15 ± 0.30 | 14.55 ± 1.39 | 10.04 b |
| Co 86032 | 34.12 ± 4.64 | 31.44 ± 6.47 | 73.35 ± 5.07 | 46.30 c |
| Co 1060 | 69.02 ± 9.95 | 53.75 ± 10.33 | 74.01 ± 6.53 | 65.59 d |
| Mean | 25.92 A ^{@1} | 21.84 A | 52.84 B | |
| Percent of intensity | | | | |
| Co 62019 | 9.66 ± 0.97 | 8.69 ± 0.70 | 15.00 ± 1.67 | 11.12 a ^{@2} |
| Co 62213 | 11.09 ± 2.84 | 10.35 ± 0.38 | 13.01 ± 0.67 | 11.48 a |
| Co 389 | 12.84 ± 1.65 | 9.88 ± 2.39 | 12.63 ± 2.15 | 11.78 a |
| Co 293 | 11.20 ± 1.61 | 6.99 ± 0.19 | 6.34 ± 0.95 | 8.18 a |
| Co 86032 | 14.13 ± 0.94 | 7.88 ± 0.39 | 10.94 ± 0.48 | 10.98 a |
| Co 1060 | 11.61 ± 0.63 | 7.89 ± 0.18 | 13.25 ± 0.72 | 10.92 a |
| Mean | 11.76 A ^{@2} | 8.61 B | 11.86 A | |
| Infestation index | | | | |
| Co 62019 | 1.39 ± 0.07 | 1.25 ± 0.14 | 7.81 ± 1.09 | 3.48 a ^{@2} |
| Co 62213 | 1.64 ± 0.47 | 1.23 ± 0.19 | 5.99 ± 0.93 | 2.95 a |
| Co 389 | 1.92 ± 0.27 | 1.08 ± 0.06 | 7.03 ± 1.37 | 3.34 a |
| Co 293 | 1.01 ± 0.39 | 0.50 ± 0.02 | 0.95 ± 0.24 | 0.82 b |
| Co 86032 | 4.77 ± 0.51 | 2.47 ± 0.53 | 7.98 ± 0.25 | 5.07 c |
| Co 1060 | 8.03 ± 1.24 | 4.21 ± 0.75 | 9.89 ± 1.33 | 7.38 d |
| Mean | 3.13 A ^{@2} | 1.79 B | 6.61 C | |

[@]Means ± SE followed by the same lower case letter in the column and upper case letter in the row are not significantly different ($p > 0.05$) by repeated-measures ANOVA and Tukey HSD test on ¹arcsin and ²square root transformed values.

hybrid (Co 293) emerged as resistant (Table S3) on the basis of highest incidence among the four years (14.55%), whereas the remaining five were found to be susceptible (Figure 2a) with an incidence range of 46.97–74.01%.

Under artificial conditions

Under artificial infestation, all hybrids sustained higher levels of damage (Table 2) than in natural conditions. Percent of incidence differed significantly among the six hybrids ($F_{5, 12}=8.70$; $p < 0.01$) with the lowest value in Co 293 (26.10%). However, the higher levels of incidence in the remaining five hybrids did not differ among them. On the other hand, percent of intensity ($F_{5, 12}=1.08$; $p > 0.05$) and infestation index did not differ ($F_{5, 12}=1.98$; $p > 0.05$) among different hybrids.

Relative Resistance Ratio (RRR)

RRR computed from the field data of the hybrids was significantly ($F_{5, 12}=9.37$; $p < 0.01$) lowest in Co 293 (0.167), whereas it was uniformly higher in the remaining five hybrids (Figure 2b). On the other hand, RRR for the hybrids tested under artificial conditions showed nonsignificant differences ($F_{5, 12}=1.06$; $p > 0.05$) though the value was the lowest for Co 293 (0.208) and uniformly higher for the remaining five hybrids (Figure 2c).

Oviposition behavior

The three hybrids showed significant differences in all three oviposition parameters examined for *C. sacchariphagus indicus* under free-choice situation in the laboratory. At the end of the 7-day

Table 2. Resistance status of six sugarcane hybrids against *Chilo sacchariphagus indicus* under artificial infestation in pot culture

| Hybrid | % of infestation ¹ | % of intensity ² | Infestation index ² | Resistance category [@] |
|----------|-------------------------------|-----------------------------|--------------------------------|----------------------------------|
| Co 62019 | 73.38 ± 7.96 a ¹ | 6.37 ± 2.32 a | 5.13 ± 2.60 a | Susceptible |
| Co 62213 | 65.45 ± 3.78 a | 8.76 ± 0.94 a | 6.01 ± 1.04 a | Susceptible |
| Co 389 | 63.37 ± 8.32 a | 8.29 ± 1.86 a | 5.81 ± 2.30 a | Susceptible |
| Co 293 | 26.10 ± 1.57 b | 5.93 ± 0.17 a | 1.62 ± 0.06 a | Moderately Resistant |
| Co 86032 | 76.45 ± 6.62 a | 9.25 ± 0.61 a | 6.98 ± 1.48 a | Susceptible |
| Co 1060 | 69.24 ± 2.26 a | 8.39 ± 0.34 a | 5.99 ± 0.35 a | Susceptible |

¹Means ± SE followed by the same letter are not significantly different ($p > 0.05$) by one-way ANOVA and Tukey's HSD test on ¹arcsin and ² $(x+0.5)^{0.5}$ transformed values.

[@]On the basis of categorization followed for field screening.

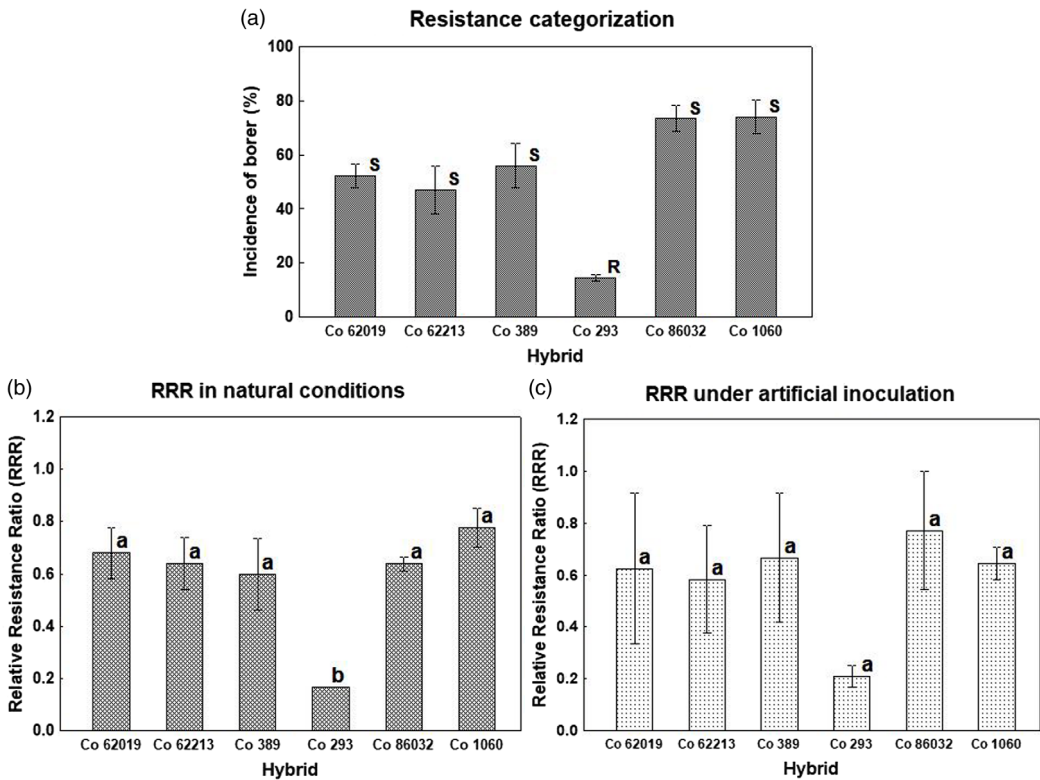


Figure 2. Relative resistance status of six sugarcane hybrids against *Chilo sacchariphagus indicus*: (a) categorization on the basis of highest infestation rate among four years (2013–2016); R – resistant; S – susceptible, (b) relative resistance ratio (RRR) in natural conditions, and (c) RRR under artificial inoculation. Vertical bars denote mean ± SE; bars with same lower case letter are not significantly different ($p > 0.05$) for RRR.

observation period, percent of leaf bits oviposited differed significantly ($F_{2, 63} = 12.48$; $p < 0.01$) among the three hybrids (Table 3). It was significantly lowest in Co 293, whereas it did not differ between the other two hybrids. The percent of egg masses deposited on the resistant hybrid (Co 293) was significantly ($F_{2, 63} = 17.03$; $p < 0.01$) lowest among the three hybrids; the susceptible Co 86032 and Co 1060 showed similar percent of egg masses. Percent of egg numbers deposited also varied significantly ($F_{2, 63} = 29.35$; $p < 0.01$) among the hybrids with Co 293 receiving significantly lowest percent of eggs while the other two recording about four times higher, yet

Table 3. Oviposition behavior of *Chilo sacchariphagus indicus* on three sugarcane hybrids under free-choice situation

| Hybrid | % of leaf bits oviposited ¹ | % of egg masses deposited ² | % of egg numbers deposited ¹ | Egg no./mass ² |
|----------|--|--|---|---------------------------|
| Co 293 | 2.83 ± 0.47 a ¹ | 15.75 ± 3.33 a | 11.74 ± 2.30 a | 11.71 ± 1.17 a |
| Co 86032 | 7.58 ± 1.09 b | 45.35 ± 4.30 b | 47.20 ± 4.14 b | 14.52 ± 0.78 a |
| Co 1060 | 7.88 ± 0.92 b | 38.90 ± 3.96 b | 41.06 ± 3.83 b | 14.00 ± 0.95 a |

¹Mean of 22 replications; ² n = 17 – 73 egg masses.

³Means ± SE followed by the same letter in a column are not significantly different ($p > 0.05$) by one-way ANOVA and Tukey's HSD test for equal or unequal replications on $(x+0.5)^{0.5}$ transformed values.

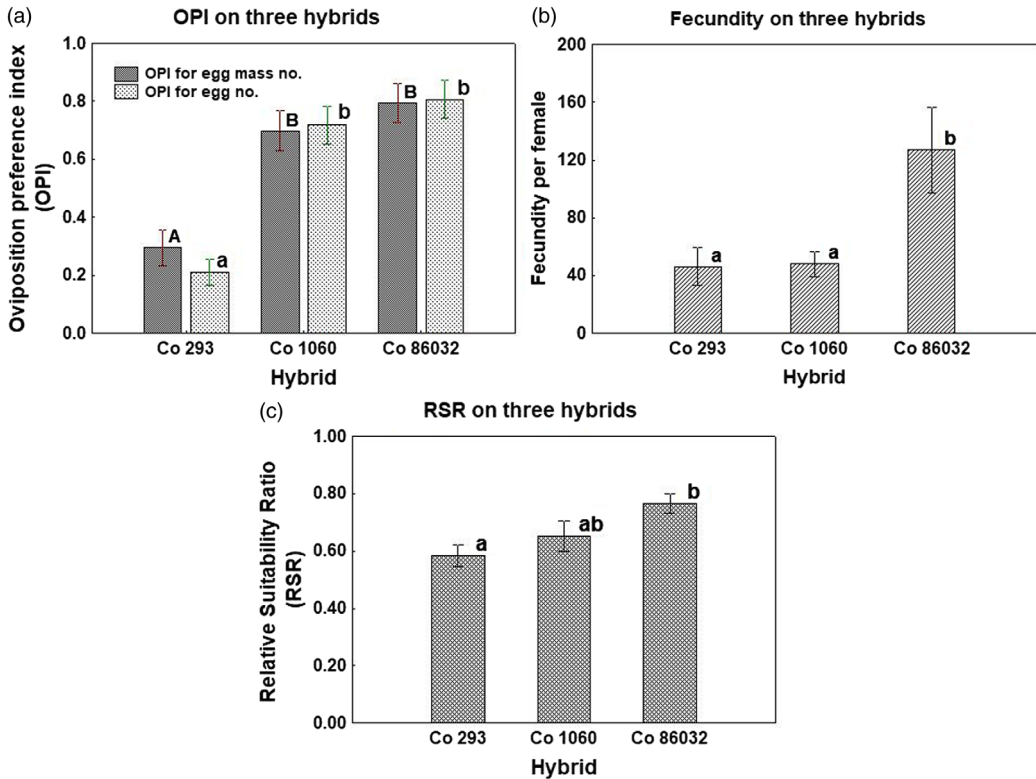


Figure 3. Behavioral and biological suitability of three sugarcane hybrids to *Chilo sacchariphagus indicus*: (a) per leaf basis oviposition preference index for egg mass number and egg number, (b) mean fecundity on shoots, and (c) relative suitability ratio on shoots. Vertical bars denote mean ± SE; bars with same upper or lower case letter are not significantly different ($p > 0.05$).

statistically similar, percent of egg numbers. Egg number/mass did not differ significantly ($F_{2, 136} = 1.14$; $p > 0.05$) among the three hybrids.

Oviposition preference index

OPI on per leaf bit basis for the three hybrids differed significantly for egg mass number ($F_{2, 63} = 15.81$; $p < 0.01$) and egg number ($F_{2, 63} = 29.66$; $p < 0.01$) (Figure 3a). Both parameters were significantly lowest on Co 293, whereas they did not differ between Co 1060 and Co 86032. When computed on the basis of weight of leaf bits provided as food, OPI did not differ among the three hybrids for both egg mass number and egg number (Table 4). Similarly, OPI calculated on the

Table 4. Oviposition preference index (OPI) for *Chilo sacchariphagus indicus* attack on three sugarcane hybrids

| Hybrid | Mean weight of leaf bit (g) | OPI (weight basis) | | Mean area of leaf bit (cm ²) | OPI (area basis) | |
|----------|--------------------------------|--------------------|-----------------|--|------------------|-----------------|
| | | Egg mass no. | Egg no. | | Egg mass no. | Egg no. |
| Co 293 | 0.4893 ± 0.0423 a ¹ | 0.626 ± 0.093 a | 0.579 ± 0.093 a | 20.26 ± 0.14 a | 0.543 ± 0.089 a | 0.438 ± 0.077 a |
| Co 86032 | 1.9405 ± 0.1102 b | 0.596 ± 0.068 a | 0.686 ± 0.068 a | 53.12 ± 0.67 b | 0.678 ± 0.072 a | 0.719 ± 0.071 b |
| Co 1060 | 1.4338 ± 0.0274 c | 0.638 ± 0.067 a | 0.731 ± 0.060 a | 37.40 ± 0.61 c | 0.738 ± 0.065 a | 0.800 ± 0.057 b |
| F value | 110.85 | 0.077 | 1.089 | 961.74 | 1.724 | 7.587* |
| df | 2 & 18 | 2 & 63 | 2 & 63 | 2 & 9 | 2 & 63 | 2 & 63 |
| p | < 0.01 | > 0.05 | > 0.05 | <0.01 | > 0.05 | <0.01 |

¹Means ± SE followed by the same letter in a column are not significantly different ($p > 0.05$) by one-way ANOVA and Tukey's HSD test.

Table 5. Cumulative larval mortality rates (%) of *Chilo sacchariphagus indicus* reared on three sugarcane hybrids in the laboratory

| Hybrid | Week after setup | | | | | | Mean |
|----------|---------------------|--------------|--------------|--------------|--------------|--------------|----------------------|
| | 1 | 2 | 3 | 4 | 5 | 6 | |
| Co 293 | 13.33 ± 5.04 | 30.67 ± 4.31 | 53.33 ± 6.07 | 77.33 ± 5.47 | 85.33 ± 3.63 | 89.33 ± 2.67 | 58.22 a ¹ |
| Co 86032 | 2.67 ± 1.82 | 28.00 ± 3.27 | 38.93 ± 4.41 | 49.33 ± 5.11 | 65.33 ± 4.96 | 70.67 ± 4.31 | 42.49 b |
| Co 1060 | 1.33 ± 1.33 | 24.00 ± 2.89 | 38.67 ± 3.07 | 57.33 ± 4.31 | 74.67 ± 3.63 | 84.00 ± 2.89 | 46.67 b |
| Mean | 5.78 A ¹ | 27.56 B | 43.64 C | 61.33 D | 75.11 E | 81.33 E | |

¹Means ± SE followed by the same lower case letter in the column and upper case letter in the row are not significantly different ($p > 0.05$) by repeated-measures ANOVA and Tukey HSD test on arcsin transformed values.

basis of area of leaf bit did not differ for egg mass number. However, it was significantly lowest in Co 293 and similar for the other two hybrids for egg number.

Larval mortality rates on resistant/susceptible hybrids

Larval survival examined for *C. sacchariphagus indicus* reared on shoots showed significantly ($F_{2, 42} = 9.13$; $p < 0.01$) highest mean mortality rate in the hybrid Co 293 (58.22%) (Table 5). In contrast, larval mortality rates were lower and similar when reared on the susceptible hybrids Co 86032 and Co 1060. Mean mortality rates of larvae increased progressively over the period of observation with significant differences ($F_{5, 210} = 223.87$; $p < 0.01$) among different weeks. Neonate larval mortality in Co 293 was 10-fold greater than that in Co 1060 and five-fold greater than that in Co 86032 in the first week of observation. Thereafter, mortality gradually increased in all the hybrids. The hybrid × week interaction was also significant ($F_{10, 210} = 2.41$; $p < 0.01$).

Biology of *C. sacchariphagus indicus* on three hybrids

Life cycle in shoots

Larval developmental parameters did not vary considerably when reared on shoot bits of different hybrids in the laboratory. Larval and pupal periods were the highest on Co 293 but did not differ significantly either among the hybrids or between sexes (Table 6). However, pupal weight differed significantly among hybrids and sexes: mean pupal weight was the highest in the susceptible Co 86032, while it was similar in the other two hybrids; it was significantly higher in female than in male. Just as larval and pupal periods, adult longevity did not differ either among hybrids or sexes. Sex ratio of emerging adults ranged from 1.2: 1.0 (♂ :♀) in Co 293 and Co 86032 to 1.4: 1.0 (♂ :♀) in Co 1060 resulting in a slightly higher percent of males (58.8%) in Co 1060 than in Co 293 and Co 86032 (54.5%). Fecundity of females emerging from rearings on the three hybrids differed

Table 6. Life cycle of *Chilo sacchariphagus indicus* reared on shoots of three sugarcane hybrids in the laboratory

| Hybrid | Male | Female | Mean |
|------------------------|-----------------------------|-------------------|-----------------------------|
| Larval period (days) | | | |
| Co 293 | 65.75 ± 2.38 | 67.00 ± 2.04 | 66.32 ± 1.57 a ¹ |
| Co 86032 | 59.67 ± 4.16 | 60.70 ± 3.77 | 60.14 ± 2.78 a |
| Co 1060 | 61.90 ± 2.99 | 60.29 ± 5.08 | 61.24 ± 2.64 a |
| Mean | 62.47 ± 1.90 A ¹ | 62.93 ± 2.07 A | |
| Pupal period (days) | | | |
| Co 293 | 7.92 ± 0.83 | 9.10 ± 0.60 | 8.45 ± 0.53 a |
| Co 86032 | 8.58 ± 0.47 | 7.30 ± 0.52 | 8.00 ± 0.37 a |
| Co 1060 | 8.40 ± 0.90 | 6.71 ± 1.51 | 7.71 ± 0.81 a |
| Mean | 8.29 ± 0.42 A | 7.81 ± 0.51 A | |
| Pupal weight (g) | | | |
| Co 293 | 0.0483 ± 0.0032 | 0.0678 ± 0.0020 | 0.0572 ± 0.0029 a |
| Co 86032 | 0.0561 ± 0.0039 | 0.0836 ± 0.0050 | 0.0686 ± 0.0043 b |
| Co 1060 | 0.0495 ± 0.0019 | 0.0651 ± 0.0039 | 0.0559 ± 0.0027 a |
| Mean | 0.0514 ± 0.0019 A | 0.0729 ± 0.0027 B | |
| Adult longevity (days) | | | |
| Co 293 | 6.33 ± 0.64 | 7.60 ± 0.62 | 6.91 ± 0.46 a |
| Co 86032 | 7.75 ± 0.62 | 5.90 ± 0.72 | 6.91 ± 0.50 a |
| Co 1060 | 6.40 ± 0.37 | 6.00 ± 0.38 | 6.24 ± 0.26 a |
| Mean | 6.85 ± 0.34 A | 6.56 ± 0.39 A | |

¹Means ± SE followed by the same lower case letter in the column and upper case letter in rows for each parameter are not significantly different ($p > 0.05$) by factorial ANOVA and Tukey unequal N HSD test.

significantly ($F_{2, 38}=6.21$; $p < 0.01$); it was significantly lower in Co 293 and Co 1060 than in Co 86032 (Figure 3b).

Relative suitability ratio (RSR)

RSR computed from ranks based on larval and pupal durations in shoots differed significantly among the three hybrids ($F_{2, 48}= 4.585$; $p < 0.05$). Co 293 emerged as the most unsuitable host with significantly lowest RSR of 0.583 (Figure 3c), whereas Co 86032 was the most suitable for larval development with the highest RSR of 0.765; Co 1060 showed intermediate response (RSR=0.652).

Plant characters vs. borer injury

Role of plant morphology

Among the selected quantitative plant morphological characters, leaf length and cane thickness were positively and significantly correlated with borer incidence for the six hybrids that entered the fourth field trial; however, only leaf length showed significant positive correlation with intensity (Table 7). On the other hand, morphological characters were not significantly correlated with infestation parameters for the 52 hybrids selected at the end of three field trials. Nonparametric analysis of qualitative characters vis-à-vis borer attack rates showed nonsignificant relationship for rind color, pith, sheath-clasp, and leaf hairiness (Table 8). For sheath-clasp, despite nonsignificant ANOVA, percent infestation was significantly ($z=2.672$) lowest in medium class (mean rank=21.23) and highest in loose class (mean rank=33.28).

Phenolics vs. borer incidence

Among the phenolics determined in shoots of the three hybrids, rutin, vanillic acid, ellagic acid, ferulic acid, coumarin, flavone, catechin, and orcinol showed a clear trend of highest quantity in Co 293 followed by Co 1060 and Co 86032 in lower amounts (Table S5). The two phenolics gallic acid and phloroglucinol were totally absent in Co 293. While gallic acid was present in very high

Table 7. Correlations between plant characters and *Chilo sacchariphagus indicus* attack rates on sugarcane hybrids screened under natural conditions

| Parameter | % infestation | | % intensity | |
|--|---------------|----------|-------------|----------|
| | <i>r</i> | <i>p</i> | <i>r</i> | <i>p</i> |
| Correlations for 52 hybrids after three field trials | | | | |
| Leaf length (cm) | −0.060 | 0.673 | 0.068 | 0.630 |
| Leaf width (cm) | −0.043 | 0.763 | 0.145 | 0.304 |
| Cane thickness (cm) | 0.192 | 0.174 | 0.073 | 0.608 |
| Cane length (cm) | 0.130 | 0.357 | 0.157 | 0.266 |
| Single cane weight (kg) | 0.035 | 0.808 | 0.111 | 0.435 |
| Brix % | −0.103 | 0.467 | −0.014 | 0.923 |
| Sucrose % | −0.043 | 0.761 | −0.065 | 0.649 |
| Correlations for six hybrids after four field trials | | | | |
| Leaf length (cm) | 0.927** | 0.008 | 0.883* | 0.020 |
| Leaf width (cm) | 0.730 | 0.099 | 0.516 | 0.295 |
| Cane length (cm) | 0.640 | 0.172 | 0.755 | 0.082 |
| Cane thickness (cm) | 0.817* | 0.047 | 0.677 | 0.140 |
| Mean internode hardness | 0.290 | 0.577 | 0.386 | 0.450 |
| Fiber | −0.674 | 0.142 | −0.516 | 0.294 |
| Single cane weight (kg) | 0.524 | 0.286 | 0.676 | 0.140 |
| Brix % | −0.029 | 0.957 | −0.577 | 0.231 |
| Sucrose % | 0.344 | 0.505 | −0.361 | 0.483 |

* $p < 0.05$; ** $p < 0.01$; rest not significant.

Table 8. Relationship of stem and leaf characteristics with attack rates of *Chilo sacchariphagus indicus* in 52 sugarcane hybrids

| Parameter | Rind color ^a | Pith ^a | Sheath-clasp ^a | Leaf hairiness ^b |
|---------------|---|---------------------------------------|---------------------------------------|---------------------------------|
| Infestation % | $\chi^2 = 7.244^{ns}$ (df=13, N=52) | $\chi^2 = 0.597^{ns}$ (df=2, N=52) | $\chi^2 = 7.158^{ns}$ (df=3, N=52) | U=276.5 ^{ns} 35, 17 |
| Intensity % | $\chi^2 = 12.953^{ns}$ (df=13, N=52) | $\chi^2 = 1.092^{ns}$ (df=2, N=52) | $\chi^2 = 3.339^{ns}$ (df=3, N=52) | U=243.5 ^{ns} 35, 17 |

^aKruskal–Wallis ANOVA

^bMann–Whitney *U* test.

^{ns} $p > 0.05$.

quantities in the remaining two hybrids, phloroglucinol was observed in far smaller quantities. Caffeic acid was the lowest in Co 293 and highest in Co 1060; syringic acid did not show clear-cut trend.

Discussion

The first trial with 535 hybrids was conducted at ICAR-SBIRC to avoid the movement of germplasm out of the center. The process of elimination of susceptible hybrids adopted in the study ensured that a manageable number of hybrids was carried forward for subsequent replicated trials in the pest endemic area. In three trials (2014–2016) conducted in the borer endemic area, incidence was the lowest in 2014 but continued to increase reaching its peak in 2016, the highest being in the susceptible hybrids in all three years. Such increased levels of incidence, which could partly be due to variation in weather factors during the study period, ensured repeated screening of possible escapes. Consequently, the number of resistant hybrids decreased progressively over the study period ultimately leaving only Co 293 as resistant at the end of the final field trial.

Under intense borer attack facilitated by artificial inoculation, however, Co 293 turned out to be moderately resistant, whereas the remaining three hybrids maintained their susceptible status. The categorization protocol of Mukunthan (2001) and Mahesh *et al.*, (2018) for *C. sacchariphagus*

indicus followed in the present study demarcated lower ranges of infestation in each category than those proposed by Radadia and Shinde (2013) in their system of classification. Despite following such more stringent classification, the hybrid Co 293 maintained its resistance status.

To evaluate sugarcane cultivars against *D. saccharalis*, Bessin *et al.*, (1990) developed the parameter of relative survival as the number of adult emergence holes relative to the number of bored internodes which was later adopted by Reay-Jones *et al.*, (2005). Subsequently, Wilson *et al.*, (2015) developed a RRR for *E. loftini* based on rankings given to proportions of bored internodes and relative survival to accommodate the discrepancies observed in the levels of resistance based on these two considered individually. In contrast, canes affected by *C. sacchariphagus indicus* do not exhibit adult emergence holes as the larvae pupate in leaf axils after exiting the bored internode (David 1986). Hence, we used rankings based on percent of incidence (cane-based damage) and percent of intensity (internode-based damage) to modify RRR. The lowest RRR recorded for Co 293, though significantly different from that of the remaining hybrids in the field trial alone, clearly indicated its resistance status.

Eoreuma loftini oviposits exclusively on dry leaves, dry tips of leaves, dry leaf sheaths, between leaf sheath and stalk, and in concealed sites on dried sugarcane leaves located on the lower part of the plant (reviewed in Reay-Jones *et al.*, 2007). Hence, Wilson *et al.*, (2015) measured host preference in terms of early instar larval injury recorded as percentage of bored internodes which reflects not only establishment of early instar larvae but also oviposition preference. However, *C. sacchariphagus indicus* oviposits only on fresh leaves, showing preferences between adaxial and abaxial surfaces; upper, middle, and lower portions; and lamina, midrib, and leaf margin (Anonymous 2018). First instar larvae feed on inner side of top leaf sheath for a week and only second instar tunnels into the top internode (David 1986). In view of the gap between oviposition and larval tunneling, proportion of internodes tunneled cannot taken as an index of oviposition preference. Hence, oviposition preference was examined in the laboratory on excised leaves, giving due consideration to the preferences exhibited by borer females for egg laying (Anonymous 2018).

In studies on *E. loftini*, a sugarcane cultivar with greater volume of dry leaves was more attractive than another cultivar for oviposition and this decreased egg laying in the latter was suggested as antixenosis conferring resistance (Reay-Jones *et al.*, 2007). Further, complete lack of oviposition by *D. saccharalis* on two cultivars was suggested to be due to nonpreference by ovipositing females (Salgado *et al.*, 2022b). In the present study, that the moths of *C. sacchariphagus indicus* experienced antixenosis on leaf bits of Co 293 was evident from the significantly lowest proportions of leaf bits oviposited, egg masses deposited and egg numbers deposited in this hybrid in free-choice tests. However, the slightly yet nonsignificantly smaller egg number per mass in this hybrid indicated that Co 293 did not induce behavioral change suggesting lack of complete immunity.

The significantly lowest OPI on per leaf bit basis in Co 293 for both egg mass number and egg number was suggestive of antixenosis. Since the mean weight and area of the ≈ 17 cm L leaf bits of the three hybrids provisioned in the studies showed considerable variation, which indicated a gradation in biomass, OPI was computed on the basis of weight or area of leaf bit. Such standardization also distinguished the hybrids, though significant for egg number on area basis alone. Since volatile production is related to leaf size (Agelopoulos *et al.*, 2000) and leaf size is smallest in Co 293 among the three hybrids, probably due to the presence of *S. spontaneum* in its lineage (Balasundaram *et al.*, 2005), the antixenosis in this resistant hybrid could be due to lower quantities of attractive volatiles and higher quantities of repellents emitted, besides the possible influence of variable free amino acids (Reay-Jones *et al.*, 2007) and headspace volatiles produced by sugarcane (Salin *et al.*, 2021). In addition, the presence of *S. spontaneum* in the lineage may have conferred borer resistance in the form of nonpreference for oviposition known for the species (David 1986).

An earlier study with *D. saccharalis* on sugarcane cultivars suggested that reduced frequency of establishment of young larvae in the stalk would be a major component of resistance to the borer

(White 1993). Significantly lowest survival of the borer in laboratory rearing tests established Co 293 as the most unsuitable among the three hybrids. Further, 5-10 fold higher levels of initial mortality of neonate larvae in Co 293 than in the other two hybrids suggested possibility of this hybrid showing early resistance against the borer probably mediated by the high quantities of the eight phenolics detected in HPLC studies. When reared on shoots, larval development of *C. sacchariphagus indicus* was not affected unlike in earlier studies wherein prolongation of larval duration was observed in resistant sugarcane cultivars for *Chilo infuscatellus* Snellen (Lepidoptera: Crambidae) (Bhavani *et al.*, 2012) and *D. saccharalis* (Salgado *et al.*, 2022b). However, pupal weight was significantly lower for *C. sacchariphagus indicus* in Co 293 than in the susceptible Co 86032. Apparently, minor nutritional or biochemical differences, including phenolics, in the hybrids did not affect larval duration upon sustained feeding but reduced metabolism and conversion of food into biomass resulting in lower pupal weight with females showing greater tolerance. Lower fecundity in the resistant Co 293, an apparent effect of reduced pupal weight, could probably be due to nutritional or biochemical factors. Examination of amount of food consumed and digested in relation to biomass accumulated would throw more light on the role of nutritional biochemistry of the hybrids on borer biology and possible antibiosis effect. RSR computed using the ranks of larval and pupal durations in shoots accommodated the variations in these parameters and exhibited a definite trend, once again establishing the unsuitability of Co 293.

The weak and nonsignificant correlations between morphological characters and infestation parameters of *C. sacchariphagus indicus* for the 52 hybrids at the end of third field trial could be due to the presence of large number of susceptible entries. On the other hand, among the six hybrids evaluated in the final year, correlations were generally high, though only a few were significant, probably due to the smaller number and minimum variability. Nevertheless, these trends indicated a possible role for plant morphological characters: positive influence of leaf length and width in terms of site selection for oviposition; positive relationship with cane length and thickness suggested the selection of long and thick canes that offer greater quantity of food; negative correlation with fiber content in the cane was suggestive of reduced larval feeding. Cane length, and length and girth of vulnerable portions are known to positively influence borer infestation in genotypes (Asha *et al.*, 2019). In our earlier study with *Saccharum* spp. clones, cane thickness positively influenced borer incidence in *S. officinarum* alone (Mahesh *et al.*, 2018). Internode hardness, fiber, and pith did not show strong relationship with borer incidence in the present study though in an earlier study with progeny from a cross, internode rind hardness and fiber were more closely associated with resistance than pith (White *et al.*, 2006). Higher infestation in loose sheath-clasp varieties suggested their suitability since tight leaf sheath is known to act as an adverse barrier for initial establishment of neonate larvae (Easwaramoorthy and Nandagopal 1986), and it may also render leaf axils inaccessible for pupation. Such a trend was noticed earlier with the borer even in *S. barberi* clones (Mahesh *et al.*, 2018).

Higher quantities of eight phenolics in Co 293 indicated their possible role as resistant factors. On the other hand, two phenolics absent in Co 293 but present in high quantities in the other two hybrids suggested that they could be acting as possible phagostimulant factors conferring susceptibility. Total phenols in the cane were shown to be negatively related to incidence levels of the borer in selected genotypes earlier (Asha *et al.*, 2019). The limited number of hybrids examined for phenolic composition in the present study disallowed computation of such correlations. Establishment of such relationships and examination of borer biology on phenolic-incorporated diet would establish their role as resistant or phagostimulant factors. Besides, significant impact of constitutive phenolics in sugarcane hybrids on the abundance of woolly aphid *Ceratovacuna lanigera* Zehntner (Hemiptera: Aphididae) (Srikanth *et al.*, 2009) indicated their role in imparting resistance against not only borers but also sucking pests.

In the present study, 535 sugarcane hybrids were selected from the Indian hybrid collection based on regularity of flowering to facilitate easy introgression in breeding programs. However, the four-year elaborate screening process in the field and controlled conditions, supported by laboratory tests

on borer biology, culminated in the identification of only one resistant hybrid. Such low levels of resistance in hybrids could be due to the general lack of borer resistance in their predominant progenitor *S. officinarum* (Mahesh *et al.*, 2018). Nevertheless, the resistance observed in Co 293 can be exploited in breeding programs for borer resistance by deploying it as a parent.

Acknowledgements. We thank the personnel of M/s Rajshree Sugars and Chemicals Ltd., Mundiampakkam, Tamil Nadu State, India, for facilitating field experiments and data collection in their research farm. We thank our Director for academic encouragement. We are grateful to Dr. P. Govindaraj, Principal Scientist, Division of Crop Improvement, for arranging rind hardness test and fiber analysis. The help rendered by Ms. M. Dhanalakshmi in maintaining insect cultures and recording observations in the laboratory is gratefully acknowledged.

Supplementary material. To view supplementary material for this article, please visit <https://doi.org/10.1017/S0014479724000024>

References

- Agelopoulos N.G., Chamberlain K. and Pickett J.A. (2000). Factors affecting volatile emissions of intact potato plants, *Solanum tuberosum*: variability of quantities and stability of ratios. *Journal of Chemical Ecology* **26**, 497–511. <https://doi.org/10.1023/A:1005473825335>
- Albert S., Thirumalai M., Krishnamurthi M., Ramya M., Lourdusamy A. and Gopinathan M.C. (2007). Evaluation of tolerance and susceptibility against internode borer of sugarcane. *Sugar Tech* **9**, 308–311.
- Ananthanarayana K. and David H. (1986). Chemical control. In David H., Easwaramoorthy S. and Jayanthi R. (eds), *Sugarcane Entomology in India*. Coimbatore, India: Sugarcane Breeding Institute, pp. 423–435.
- Anonymous (2018). *Annual Report 2017-18*, Coimbatore: ICAR-Sugarcane Breeding Institute, 208p
- Artschwager E. and Brandes E. W. (1958). Sugarcane (*Saccharum officinarum* L.): Origin, classification, characteristics and descriptions of representative clones. United States Department of Agriculture Handbook. No. 122, 307 p.
- Asha N., Patel V.N., Sugeetha G., Pankaja N.S., Mahdev J. and Kamaraddi V. (2019). Antixenosis and antibiosis of sugarcane varieties on the incidence of sugarcane internode borer, *Chilo sacchariphagus indicus*. *International Journal of Chemical Studies* **7**, 1008–1012.
- Babu C., Koodalingam K., Natarajan U. S., Shanthi R. M. and Govindaraj P. (2009). Assessment of rind hardness in sugarcane (*Saccharum* spp. Hybrids) genotypes for development of non-lodging erect canes. *Advances in Biological Research* **3**, 48–52.
- Balasundaram N., Pramachandran M.N., Chandran K. and Natarajan U.S. (2005). Catalogue on Sugarcane Genetic Resources VI. Indian Hybrids at Sugarcane Breeding Institute, Kannur. Sugarcane Breeding Institute, Coimbatore, India. 237 p.
- Bessin R. T., Reagan T. E. and Martin F. A. (1990). A moth production index for evaluating sugarcane cultivars for resistance to the sugarcane borer (Lepidoptera: Pyralidae). *Journal of Economic Entomology* **83**, 221–225. <https://doi.org/10.1093/jee/83.1.221>
- Bhavani B., Reddy K.D., Rao N.V. and Lakshmi M.B. (2012). Biochemical basis for antibiosis mechanism of resistance in sugarcane to early shoot borer, *Chilo infuscatellus* Snellen. *Tropical Agricultural Research* **23**, 126–141. <http://doi:10.4038/tar.v24i2.7998>
- Clements H.F. and Ghotb A. 1969. The numbering of leaves and internodes for sugarcane nutrition studies. *Proceedings of the International Society of Sugarcane Technologists XIII Congress*, Taiwan, pp. 569–584.
- David H. (1986). The internode borer, *Chilo sacchariphagus indicus* (Kapur). In David H., Easwaramoorthy S. and Jayanthi R. (eds), *Sugarcane Entomology in India*. Coimbatore, India: Sugarcane Breeding Institute, pp. 121–134
- David H. and Nandagopal V. (1986). Pests of sugarcane – distribution, symptomology of attack and identification. In David H., Easwaramoorthy S. and Jayanthi R. (eds), *Sugarcane Entomology in India*. Coimbatore, India: Sugarcane Breeding Institute, pp. 1–29
- Duttamajumder S.K. (2020). *Moth borers of sugarcane*. Daya Publishing House, New Delhi, India. 639 p.
- Easwaramoorthy S. and Nandagopal V. (1986). Life tables of internode borer, *Chilo sacchariphagus indicus* (K.) on resistant and susceptible varieties of sugarcane. *Tropical Pest Management* **32**, 221–228. <https://doi.org/10.1080/09670878609371067>
- Goebel R., Fernandez E., Tibere R. and Alauzet C. (1999). Damage and yield losses to sugarcane caused by the stem borer *Chilo sacchariphagus* (Bojer) (Lep.: Pyralidae) on Reunion. *Annales- Societe Entomologique de France* **35**, 476–481.
- Govindaraj P., Parthiban S., Hari K., Naik R. and Kotwaliwale N. (2014). Non-destructive and simple estimation of fibre content – an aid to energy cane development programme. In Viswanathan R., Bhaskaran A., Hemaprabha G., Ramasubramanian T. and Nair N.V. (eds), *Proceedings of the Symposium on Bioenergy for Sustainable Development – The Potential Role of Sugar Crops*. Coimbatore, India: Sugarcane Breeding Institute (ICAR), pp. 123–126.
- Jayakumar J. (2018). Screening of sugarcane clones for resistance to early shoot borer and internode borer. *Annals of Plant Science* **26**, 270–275. <https://doi.org/10.5958/0974-0163.2018.00061.7>

- Jayanthi R. (1988). Evaluation of *Saccharum* clones for resistance to the sugarcane internode borer. *Sugar Cane* (Spring Supplement):17–18.
- Jhansi K. and Rao B.R. (1996). Evaluation of promising varieties of sugarcane for shoot borer and internode borer under different dates of plantings. *Cooperative Sugar* 27, 675–679.
- Keeping M.G. (2006). Screening of South African sugarcane cultivars for resistance to the stalk borer, *Eldana saccharina* Walker (Lepidoptera: Pyralidae). *African Entomology* 14, 277–288.
- Korowi K. T. and Samson P. R. (2013). Screening for borer resistance among sugarcane clones in Papua New Guinea, 2010–2012. *Proceedings of Australian Society of Sugarcane Technologists* 35, 1–9.
- Long W.H. and Hensley S.D. (1972). Insect pests of sugar cane. *Annual Review of Entomology* 17, 149–176. <https://doi.org/10.1146/annurev.en.17.010172.001053>
- Mahesh P., Srikanth J., Chandran K. and Singaravelu B. (2018). Resistance of *Saccharum* spp. against *Chilo sacchariphagus indicus* (Kapur) (Lepidoptera: Crambidae) in India. *Experimental Agriculture* 54, 83–95. <https://doi.org/10.1017/S0014479716000697>.
- Mahesh P., Srikanth J., Salin K.P., Singaravelu B., Chandran K. and Mahendran B. (2019). Phenology of sugarcane leaf hopper *Pyrrilla perpusilla* (Walker) (Homoptera: Lophopidae) and its natural enemies in a crop island scenario. *Crop Protection* 120, 151–162. <https://doi.org/10.1016/j.cropro.2019.02.020>
- Mehta U.K. and David H. (1978). A laboratory technique for rearing the sugarcane internode borer, *Chilo sacchariphagus indicus* K. on artificial medium. *Indian Sugar* 28, 38–41.
- Milligan S.B., Balzarini M. and White W.H. (2003). Broad-sense heritabilities, genetic correlations, and selection indices for sugarcane borer resistance and their relation to yield loss. *Crop Science* 47, 1729–1735. <https://doi.org/10.2135/cropsci2003.1729>
- Mukunthan N. (2001). Reaction of *Erianthus* to sugarcane pests. In Sreenivasan T.V., Amalraj V.A. and William Jebadas A. (eds), *Catalogue on Sugarcane Genetic Resources- IV. Erianthus species*. Coimbatore, India: Sugarcane Breeding Institute, pp. 73.
- Mukunthan N. and Rakkiyappan P. (1989). Bunchy top formation in sugar cane caused by the internode borer and its effect on yield and quality. *Sugar Cane* 2, 17–19.
- Nalawade S.V., Indi D.V., Bhilare R.L., Thorvae D.S., Raskar B.S. (2022). Field screening of promising sugarcane genotypes against whip smut disease and borer pests. *Pharma Innovation* 11, 2017–2019.
- Pasupathy S., Shanmuganathan M., Ravichandran V. and Babu C. (2021). Field screening of sugarcane clones against the internode borer, *Chilo sacchariphagus indicus*. *Uttar Pradesh Journal of Zoology* 42, 54–61.
- Posey F.R., White W.H., Reay-Jones F.P.F., Gravois K., Salassi M.E., Leonard B.R. and Reagan T.E. (2006). Sugarcane borer (Lepidoptera: Crambidae) management threshold assessment on four sugarcane cultivars. *Journal of Economic Entomology* 99, 966–971. <https://doi.org/10.1093/jee/99.3.966>
- Puthira Prathap D., Karpagam C., Bhaskaran A. and Nair N.V. (2014). Compendium of Research Articles and Status Papers. 45th meeting of Sugarcane Research and Development Workers of Tamil Nadu and Pondicherry, August 26–27, 2014, Tiruchirappalli. Sugarcane Breeding Institute, Coimbatore. 328 p. ISSN: 0973-8185.
- Radadia G.G. and Shinde C.U. (2013). *Research methodology for recording observations of sugarcane pests*. AICRP on Sugarcane, Indian Institute of Sugarcane Research, Lucknow, India. 35 p.
- Rajendran B., Gopalan M. and Hanifa A.M. (1996). Screening for field reaction of sugarcane to internode borer *Chilo sacchariphagus indicus* (Kapur). *Indian Sugar* 46(4): 257–261.
- Reagan T.E. and Mulcahy M.M. (2019). Interaction of cultural, biological, and varietal controls for management of stalk borers in Louisiana sugarcane. *Insects* 10, 305. <https://doi.org/10.3390/insects10090305>
- Reagan T.E., Way M.O., Beuzelin J.M. and Akbar W. (2008). Assessment of varietal resistance to the sugarcane borer and the Mexican rice borer. Sugarcane Annual Progress Reports 2008, Louisiana State University Agricultural Center, Baton Rouge. 182p.
- Reay-Jones, F.P.F., Way M.O., Tamou M.Se., Legendre B.L. and Reagan T.E. (2003). Resistance to the Mexican rice borer (Lepidoptera: Crambidae) among Louisiana and Texas sugarcane cultivars. *Journal of Economic Entomology* 96, 1929–1934. <https://doi.org/10.1093/jee/96.6.1929>
- Reay-Jones F.P.F., Showler A.T., Reagan T.E., Legendre B.L., Way M.O. and Moser E.B. (2005). Integrated tactics for managing the Mexican rice borer (Lepidoptera: Crambidae) in sugarcane. *Environmental Entomology* 34, 1558–1565. <https://doi.org/10.1603/0046-225X-34.6.1558>
- Reay-Jones F.P.F., Wilson L.T., Showler A.T., Reagan T.E. and Way M.O. (2007). Role of oviposition preference in an invasive Crambid impacting two graminaceous host crops. *Environmental Entomology* 36: 938–951. <https://doi.org/10.1093/ee/36.4.938>
- Salgado L.D., Wilson B.E., Richard R.T., Penn H.J. and Way M.O. (2022a). Characterization of resistance to the Mexican rice borer (Lepidoptera: Crambidae) among Louisiana sugarcane cultivars. *Insects* 13, 890. <https://doi.org/10.3390/insects13100890>
- Salgado L.D., Wilson B.E., Villegas J.M., Richard R.T. and Penn H.J. (2022b). Resistance to the sugarcane borer (Lepidoptera: Crambidae) in Louisiana sugarcane cultivars. *Environmental Entomology* 51, 196–203. <https://doi.org/10.1093/ee/nvab118>

- Salin K.P., Srikanth J., Singaravelu B. and Nirmala R. (2021). Decoding chemical signals in head-space volatiles involved in tri-trophic interactions and induced resistance: A case study in sugarcane. *Proceedings of the International Conference on Sugarcane Research: Sugarcane for Sugar and Beyond*, 516-517. Coimbatore, India: ICAR-Sugarcane Breeding Institute.
- Sallam M. and Allsopp P. (2003). Preparedness for borer incursion: SRDC final report BSS. 249p.
- Showler A.T. and Reagan T.E. (2012). Ecology and tactics of control for three sugarcane stalk boring species in the Western Hemisphere and Africa. In Gonclaves J. and Correia K. (eds), *Sugarcane: production, cultivation and uses*. Nova Science Publishers, Ha, pp. 1–15
- Showler A.T. and Reagan T.E. (2017). Mexican rice borer, *Eoreuma loftini* (Dyar) (Lepidoptera: Crambidae): range expansion, biology, ecology, control tactics, and new resistance factors in United States sugarcane. *American Entomologist* **63**, 36–51. <https://doi.org/10.1093/ae/tmx013>
- Srikanth J. and Kurup N.K. (2011). Damage pattern of sugarcane internode borer *Chilo sacchariphagus indicus* (Kapur) in Tamil Nadu State, southern India. *International Sugar Journal* **113**, 590–594.
- Srikanth J., Salin K.P., Kurup N.K., Karthikeyan J., Mukunthan N. and Singaravelu B. (2009). Reaction of sugarcane clones to woolly aphid, *Ceratovacuna lanigera*, attack and its relationship with leaf phenolics. *Sugar Cane International* **27**, 204–209.
- StatSoft. (2004). STATISTICA (data analysis software system), version 7. Available from: www.statsoft.com.
- Sundara B. (1998). *Sugarcane Cultivation*. New Delhi, India: Vikas Publishing House. 292p.
- Tomaz A., Coutinho A., Soares B., Peternelli L., Pereira E. and Barbosa M. (2018). Assessing resistance of sugarcane varieties to sugarcane borer *Diatraea saccharalis* Fab. (Lepidoptera: Crambidae). *Bulletin of Entomological Research* **108**, 547–555. <https://doi.org/10.1017/S0007485317001183>
- Tonnesen H.H. and Karlsen J. (1983). High-performance liquid chromatography of curcumin and related compounds. *Journal of Chromatography A* **259**, 367–371. [https://doi.org/10.1016/S0021-9673\(01\)88022-5](https://doi.org/10.1016/S0021-9673(01)88022-5)
- White W.H. (1993). Movement and Establishment of Sugarcane Borer (Lepidoptera: Pyralidae) Larvae on Resistant and Susceptible Sugarcane. *Florida Entomologist* **76**, 465–473. <https://doi.org/10.2307/3495647>
- White, W.H., Tew, T.L. and Richard Jr, E.P. (2006). Association of sugarcane pith, rind hardness, and fiber with resistance to the sugarcane borer. *Journal of American Society of SugarCane Technologists* **26**, 87–100.
- White W.H., Viator R.P., Dufrene E.O., Dalley C.D., Richard Jr. E. P. and Tew T.L. (2008). Re-evaluation of sugarcane borer (Lepidoptera: Crambidae) bioeconomics in Louisiana. *Crop Protection* **27**, 1256–1261. <https://doi.org/10.1016/j.cropro.2008.03.011>
- Wilson B.E., Van Weelden M.T., Beuzelin J.M., Reagan T.E., Way M.O., White W.H., Wilson L.T. and Showler A.T. (2015). A relative resistance ratio for evaluation of Mexican rice borer (Lepidoptera: Crambidae) susceptibility among sugarcane cultivars. *Journal of Economic Entomology* **108**, 1363–1370. <https://doi.org/10.1093/jee/tov076>
- Wilson B.E., White W.H., Richard R.T. and Johnson R.M. (2021). Evaluation of sugarcane borer, *Diatraea saccharalis*, resistance among commercial and experimental cultivars in the Louisiana sugarcane cultivar development program. *International Sugar Journal* **123**, 256–261.
- Yalawar S., Pradeep S., Ajith Kumar M.A., Hosamani V. and Rampure S. (2010). Biology of sugarcane internode borer, *Chilo sacchariphagus indicus* (Kapur). *Karnataka Journal of Agricultural Sciences* **23**, 140–141.

Cite this article: Mahesh P, Singaravelu B, Srikanth J, Salin KP, Chandran K, and Nirmala R. Resistance of sugarcane hybrids to internode borer *Chilo sacchariphagus indicus* (Lepidoptera: Crambidae). *Experimental Agriculture*. <https://doi.org/10.1017/S0014479724000024>