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## **Invited Article**

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**Corresponding author:** Emily G. Mitchell; Email: ek338@cam.ac.uk From organisms to biodiversity: the ecology of the Ediacaran/Cambrian transition

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### Non-technical Summary

Animals first evolved more than 570 million years ago, during the Ediacaran time period, but it was not until well into the Cambrian time period, around 520 million years ago, that animal evolution really took off and most modern animal groups diversified. It is over this Ediacaran to Cambrian transition that we not only see animals first appear, but also the evolution of movement, the ability to burrow and to swim, and the very first reefs and macroscopic predators. There are likely many different factors that shaped this radiation of animal life, so in this review paper we discuss the ecology underlying this Ediacaran to Cambrian transition and place the individual specimens and taxa in the context of the environment in which they lived. After all, it is the interactions that organisms experienced in their daily lives with one another and their environment that led to the diversification of animal body plans, and the evolutionary patterns we observe over these crucial 75 million years. As early animals evolved, we see diversification in feeding and biological and environmental interactions. These ecological interactions started off relatively weak, with few interactions between organisms, but then increased throughout the Ediacaran and into the Cambrian. By 500 million years ago, the ecosystem structure was similar to that of marine systems today. However, there are time delays between the origins of structuring processes and the time when they have an observed impact on other organisms and their ecosystem. As such, while the key building blocks of ecosystem structure were in place by the end of the Cambrian, it takes evolutionary timescales for the impact of these innovations o be realized.

#### Abstract

The Ediacaran/Cambrian transition (ECT; ~575-500 Ma) captures the early diversification of animals, including the oldest crown-group taxa of most major animal phyla alive today. Key to understanding the drivers underneath the ECT macroevolutionary patterns are the interactions of animals with one another and their environment, and how these interactions scale up to global diversity patterns. Understanding the ecology of ECT organisms is enabled by the abundance of Lagerstätten over this time period, with a relatively large proportion of soft-bodied organisms preserved, often within the communities in which they lived. Here, we review our understanding of organismal, community, and macroecology of the ECT, and how these different scales of ecological analyses relate to the macroevolutionary diversification patterns we see over this 75 Myr time period. Across all ecological scales, we find clear trends, starting with stochastic ecosystem dynamics dominated by generalist taxa in the first Ediacaran communities, to more structured, niche-driven specialist dynamics by Cambrian Epoch 2. These trends are reflected in organism functional morphology, the complexity and strength of organisms' interactions within their communities, and large-scale metacommunity, biogeographic, and biodiversity patterns. Yet there is often a time delay between the origination of a new type of ecological interaction and when it is observed to impact the ecosystem as a whole. As such, while many modern ecological innovations were in place by the end of the Cambrian, the knock-on effects and complexity of these interactions continued to build up throughout the Phanerozoic, leading to the complex biosphere we have today.

### Introduction

One of the most important evolutionary innovations in the history of life on Earth was the evolution of animals and the subsequent transformation of the biosphere. The appearance and diversity of body and trace fossils in the Cambrian is sufficiently quick that it was termed the Cambrian explosion in the 1970s (cf. Brasier 1979), with a rapid expansion of taxonomic and morphological diversity occurring in the Cambrian Epoch 2, where many animal phyla crown groups are first found (e.g., Erwin et al. 2011; Zhang et al. 2021). These likely represent accurate timescales for animal origination and early diversification (Budd and Jensen 2017; Budd and Mann 2023; Moody et al. 2024). Since the 1970s, our understanding of the

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Cambrian Explosion and the origins and early evolution of metazoans have changed dramatically, with an appreciation that animals predate the Cambrian (Ford 1958). Indeed it was only in the 1950s that complex, macroscopic fossils were established to have been found in rocks known to be pre-Cambrian (Ford 1958), with six other localities known that are now dated as Ediacaran age (Billings and Billings 1872; Gürich 1933; Sprigg 1947; Misra 1969; Sokolov 1972; Fig. 1A-D). Yet it is only in the last few years that taxonomic affinities of some Ediacaran macrofossils have been resolved, with support from phylogenetic analyses, as total- and/or crown-group animals (Budd and Jensen 2017; Bobrovskiy et al. 2018; Chen et al. 2019; Dunn et al. 2021), supporting the diversification of early animals in the Ediacaran. The presence of Ediacaran animals (Bobrovskiy et al. 2018; Chen et al. 2019; Dunn et al. 2021), coupled to differences in when different groups radiated (Servais et al. 2023), means that the term "Ediacaran/Cambrian transition" (ECT) (cf. Narbonne et al. 1987) is a more appropriate term than Cambrian explosion for the radiation and diversification of early animals across the Ediacaran and Cambrian (e.g., Wood et al. 2019).

Our understanding of the evolutionary relationships of Cambrian organisms has similarly shifted since the 1970s, greatly influenced by research effort on exceptionally preserved fossils. Detailed redescriptions of Burgess Shale animals first described by Walcott in the 1910s-1920s (e.g., Walcott 1911a,b,c, 1912, 1918, 1920), by Whittington and colleagues, alongside descriptions of new taxa, highlighted the unusual morphology of many Cambrian soft-bodied animals. While these fossil taxa were recognizable as metazoans, they were not always straightforward to place within extant classes or even phyla (Whittington 1971, 1975; Hughes 1975; Briggs 1976; Conway Morris 1976, 1977; Conway Morris and Robison 1982; Bruton and Whittington 1983; Whittington and Briggs 1985). Over the last 30 yr, the integration of these previously problematic Cambrian animals into cladograms (Briggs and Fortey 1989; Budd 1993, 1998) and phylogenetic analyses including extant groups allowed these and other soft-bodied fossil taxa to be recognized as stem-group representatives of modern classes and phyla (Daley et al. 2009; Legg et al. 2013; Smith and Caron 2015; Park et al. 2024; Rahman and Zamora 2024). These exceptional fossils have provided critical data for understanding the evolution of disparate body plans (Fig. 1E-H), although not always without controversy (Daley et al. 2018; Nanglu and Caron 2018; Zhao et al. 2019; Nanglu et al. 2020a, 2023; Kihm et al. 2023; Park et al. 2024).

Other substantial insights into the ECT since 1975 have been gained through the large increase in terms of number of localities and sites (Fig. 2), geographic distribution of these sites, and new data-collection strategies. These data have been leveraged to further our understanding in terms not only of ECT evolutionary relationships, but also their evolutionary drivers through interrogating the ecology and ecological relationships. Whereas specimens were historically often collected in isolation, so taken out of their population and community context, now this information is commonly kept intact, enabling us to reconstruct the interactions and ecology of these organisms on ecological, not only geological, timescales. These data are further leveraged by the vast improvements in computational and imaging power since 1975, enabling more detailed studies of organisms' anatomy through photographic and scanning methods (Y. Liu et al. 2015) and complex ecological statistical analyses (e.g., Clapham et al. 2003; Mitchell et al. 2015), as well as new computationally intensive analyses, such as computational fluid dynamics (CFD; e.g.,

Rahman et al. 2015b) and finite element analysis (FEA; Bicknell et al. 2018, 2021) to investigate ECT organisms. This change in ECT paleontology is also mirrored in extant ecology, with a shift from largely descriptive and observational work to that which is more hypothesis testing-based and data driven (Anderson et al. 2021).

#### Diversity Patterns over the ECT

The term "Cambrian explosion" conjures up images of low (or no) animal life before the Cambrian, followed by a rapid/exponential radiation in terms of taxonomic diversity. This pattern was visualized first in Sepkoski (1978), who showed that the number of orders from the Ediacaran time period (then known as the Vendian) through the Cambrian followed an exponential curve that plateaued in the Cambrian Epoch 2. This historical pattern is seen across multiple groups, including trace fossils and commonly preserved fossil groups such as trilobites and brachiopods (e.g., Z. Zhang et al. 2008; Mángano and Buatois 2017; Paterson et al. 2019). Almost 50 yr later, diversity patterns appear much more complex, with an increase and subsequent decrease in the Ediacaran, followed by a large increase in the Cambrian at the base of Stage 3, with the rapid diversity increase that we associate with the Cambrian Explosion, which then plateaus (Na and Kiessling 2015). Yet these patterns are not robust across all taxa, environments, or metrics. Ichnodiversity follows the classic ECT increase (Mángano and Buatois 2014), while some groups show a pulsed radiation through the ECT (Zhuravlev and Wood 2018), and others begin to radiate at different times, some even starting their radiation in the Ordovician (Servais et al. 2023).

With the variation in timings, and distribution of radiations across different groups, it is likely that a range of different processes drove the ECT. Indeed, the exponential increase in the diversity of a group follows a null model, with background rates of origination and extinction leading to crown-group radiation (Budd and Mann 2020), so arguably requires no drivers. However, to describe the processes behind these patterns, we need to resolve which key mechanisms were operating, and even if the ECT diversification was driven purely by abiotic factors, we still need to understand how animals interacted with these abiotic factors, enabling diversification.

### Ecology

Ecology is a very broad field that includes a range of different subtopics that consider organisms' interactions on a variety of different scales. On an individual specimen level, organismal ecology describes aspects of the individual that reflect aspects of an organism's behavior, functional morphology, and/or life style. In a paleontological context, functional morphology, ichnology, and ecospace studies fall within organismal ecology.

Organismal ecology is the product of how different organisms interact with their local environment and other taxa, and these interactions and associations build up to form the network of interactions that form the ecosystem, the community ecology. Community ecology includes interactions such as competition and facilitation, as well as trophic interactions between organisms, that is, food webs. Community ecology can also include the interactions of organisms with their local habitat, and how that then impacts the organisms themselves. Population ecology also acts at a community level (communities are after all just collections of interacting populations), so includes analyses of population

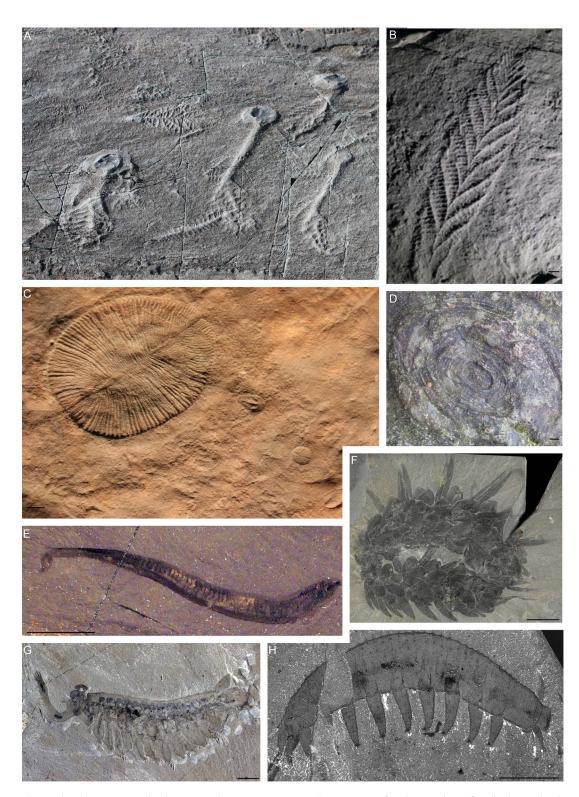


Figure 1. Iconic Ediacaran/Cambrian transition (ECT) organisms known in 1975. **A**, Mistaken Point E surface (~564 Ma), Newfoundland, Canada, showing *Charniodiscus spinous, Charniodiscus arboreus, Beothukis mistakensis*, and *Fractofusus misrai*. **B**, *Charnia masoni* holotype (LEIUG 2328), Bed B (~560 Ma) Charnwood Forest, U.K. Image credit: British Geological Survey. **C**, *Dickinsonia costa* (large specimen, left; and small specimen, right) and *Parvancorina minchami* (right middle) (~550 Ma), South Australia Museum. **D**, *Cyclomedusa* disk (~560 Ma), Charnwood Forest, U.K. **A**, **C**, **D**, Image credit: Emily G. Mitchell. **E**, Stem-group chordate *Pikaia gracilens* Walcott, 1911, Burgess Shale, Canada (Cambrian: Wuliuan) (Walcott 1911c). USNM PAL 83940B. Image courtesy of the Smithsonian Institution (CCO license) (EZID:http://n2t.net/ark:/65665/m37ec4e117-c554-4a97-b352-5deb01b3081f). **F**, Stem-group mollusk *Wiwaxia corrugata* Walcott, 1911, Burgess Shale, Canada (Cambrian: Wuliuan) (Walcott 1911c). USNM PAL 198745. Image credit: Mark Florence. Image courtesy of the Smithsonian Institution (EZID: http://n2t.net/ark:/65665/m31ec24628-518cfdc8c941). **G**, Stem-group euarthropod *Opabinia regalis* Walcott, 1912, Burgess Shale, Canada (Cambrian: Wuliuan). USNM PAL 57683. Image credit: Han Zeng. Image courtesy of the Smithsonian Institution (EZID: http://n2t.net/ark:/65665/m31ec24168-b24e-e565c31a4441). **H**, Total-group edysozoan *Aysheaia pedunculata* Walcott, 1911, Burgess Shale, Canada (Cambrian: Wuliuan) (Walcott 1911c). USNM PAL365608. Image credit: Javier Ortega-Hernández. Scale bars for **A–H**, 10 mm.

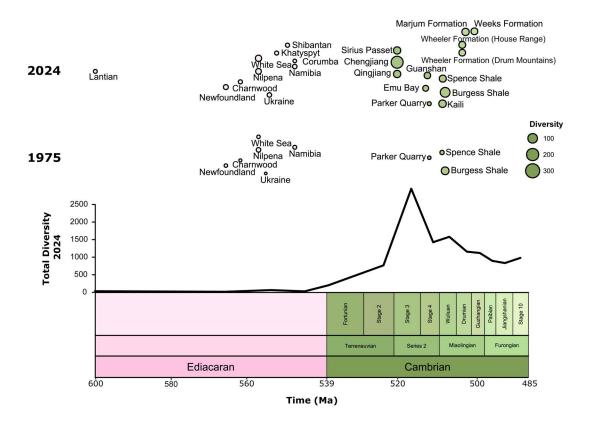


Figure 2. Key Ediacaran (pink) and Cambrian (green) Lagerstätten with their generic diversity in 1975 and 2024, and the total diversity from the Paleobiology Database (Na and Kiessling 2015) (black line).

size distributions, for example, and can also include behavioral ecology, such as the interactions between predators and prey (e.g., the landscape of fear; Laundré et al. 2001). In a paleontological context, community ecology can be inferred from organisms' functional morphology (Haug et al. 2012), with hypotheses now often interrogated with computational methods (Rahman et al. 2015a; Bicknell et al. 2021); gut contents and morphology (trophic ecology and food webs) (Butterfield 2002; Vannier and Chen 2005; Vannier 2012; Vannier et al. 2014; Strang et al. 2016; Peel 2017; Park et al. 2024); trace fossils, including predation traces (Seilacher et al. 2005; Jensen et al. 2006; Chen et al. 2013; Bicknell and Paterson 2018; Mángano and Buatois 2020; Bicknell et al. 2022); and with exceptional preservation, from quantitative analyses of census populations or communities (Wood et al. 2003; Darroch et al. 2013; Zhao et al. 2014; O'Brien and Caron 2016; Mitchell et al. 2019; Nanglu et al. 2020b).

The ecological mechanisms that shape and drive evolutionary patterns act at and within the community scale, over time leading to changes in organismal ecology and to broad patterns across space and time (between-community ecology, or macroecology). Comparisons between different communities encompasses metacommunity (and metapopulation) ecology, landscape ecology, biogeography, global ecology, movement ecology, and macroecology. As such, diversity trends over geological and evolutionary timescales are the result of organism, community, and global ecological interactions of organisms with one another and their environment.

To understand the long-term diversity and evolutionary patterns over the ECT, this review outlines our current knowledge in ECT organismal ecology, community ecology, and macroecology, and how these different scales of interactions feed into and influence the broad-scale diversity patterns we observe over the ECT. Of course, preservational biases can have huge impacts on paleoecological studies (Nanglu and Cullen 2023). However, the exceptional preservation for many fossil localities spanning the ECT means that analytical techniques normally only applicable for extant systems can be used. The preservation of Ediacaran macrofossil communities is commonly in-situ, creating nearcensus preservation, with burial under volcanic ash flow (Benus et al. 1988; Wood et al. 2003; Noble et al. 2015) and pervasive microbial mat coverage of the seafloor leading to widespread in situ preservation of benthic communities between 572 and 550 Ma (Gehling 1999; Grazhdankin 2004; Fedonkin et al. 2007; Laflamme et al. 2011; Darroch et al. 2012; Tarhan et al. 2015, 2017). The terminal Ediacaran (550-539 Ma) also has exceptional soft-bodied preservation, but many of these sites contained specimens that were transported in mass flow events (Hall et al. 2013; Meyer et al. 2014), as well as biomineralized reefs preserved within carbonate platforms (Hofmann and Mountjoy 2001; Grotzinger et al. 2005; Warren et al. 2011).

Dozens of deposits with exceptional soft-bodied preservation are known from the Cambrian (Gaines 2014; Muscente et al. 2017). These provide census-level data for deposits that have not undergone significant transport, as well as a rich record of the anatomy of animals completely lacking hard parts, as well as soft tissues such as appendages, nervous tissues, and guts not usually preserved in the shelly fossil record (Gaines 2014), just as for Ediacaran deposits with similar preservation. However the oldest of these Cambrian deposits are known from Stage 3 (Fig. 2), leaving a large gap in the soft-bodied fossil record spanning the entire Terreneuvian. Insights into the organismal ecology of Terreneuvian animals can be gleaned from trace fossils, for which a continuous record exists (Seilacher et al. 2005; Mángano and Buatois 2017; Buatois et al. 2018, 2020), as well as the small shelly and small carbonaceous fossil records (Butterfield and Harvey 2012; Kouchinsky et al. 2012); however, the lack of exceptional deposits precludes community ecology studies. Compared with the Ediacaran, exceptional soft-bodied deposits from the Cambrian Series 2 and 3 provide a more heavily sampled interval, including multiple deposits of similar age from geographically constrained areas (e.g., Burgess Shale and nearby localities [Collins et al. 1983; Caron et al. 2010; Nanglu et al. 2020b]; the Great Basin of the United States [Lieberman 2003; Lerosey-Aubril et al. 2018, 2020; Kimmig et al. 2019]; and South China [Steiner et al. 2005; X. Zhang et al. 2008; Hou et al. 2017; Fu et al. 2019]). Just as for softbodied preservation in the Ediacaran and post-Cambrian, these deposits provide a rich record important for our understanding of organismal ecology. Compared with Ediacaran macrofossils, Cambrian macrofossils are generally more readily identifiable to the phylum or class level, facilitating a more straightforward comparison with modern analogues to infer aspects of their ecology (see "Organismal Ecology"). Importantly, alongside the temporal gap in the Terreneuvian, there are geographic limitations to our understanding of softbodied Cambrian animals. Most pressingly, there is a paucity of deposits preserving soft tissues from high latitudes, and those that are known do not have the same fidelity or abundance of fossils as the best low-latitude Lagerstätten (they are Tier 3, rather than Tier 1; sensu Gaines 2014). Thus comparable preservation modes and types facilitate comparison of organism, community, and macroecology across the ECT, albeit with some temporal and geographic gaps and differences in sampling intensity.

## **Organismal Ecology**

Organismal ecology, or autecology, focuses on the adaptations that an individual organism has in response to its biotic and abiotic environment. These changes include morphological, physiological, and behavioral adaptations. Insights into autecology can be gained through functional ecology analyses such as CFD and FEA to infer trophic relationships (e.g., Gibson et al. 2020; Bicknell et al. 2021) and trace fossil analyses to understand habitat and feeding modes (Seilacher et al. 2005; Jensen et al. 2006; Mángano and Buatois 2020). These analyses can also be visualized in ecospace, whereby different axes are used to quantify the different niches in which organisms can live and how this niche or ecospace occupation changes through time (Bambach 1983). For marine organisms, the axes used are normally: extent of mobility, position relative to the substrate, and feeding modes (Bambach et al. 2007). Body and trace fossil data are used to determine whether an area of ecospace (or "Bambach cube") is occupied for a particular time slice. During the ECT, ecospace occupation changed dramatically between the Ediacaran and Cambrian, with the advent of animals, mobility, and many new feeding modes. In the Ediacaran, 12 Bambach cubes were occupied, with 30 in the Cambrian, and 92 in the modern (Bambach et al. 2007; Bush et al. 2011).

## Marine Animal Forests: Lantian and Avalon Assemblages (Pre-558 Ma)

The first putative animals were sessile, benthic organisms that lived attached to the seafloor. The oldest are putative cnidarians from the Lantian biota, China (Wan et al. 2016) (up to 600 Ma), and widespread, abundant, large, complex fossils found predominantly in Newfoundland, Canada, and Charnwood Forest, U.K. (572-560 Ma), known as the Avalon assemblage (Waggoner 2003; Noble et al. 2015; Matthews et al. 2021). Avalon communities are dominated by rangeomorphs, a group of fractally branching organisms unique to the Ediacaran (Narbonne 2004; Brasier et al. 2012; Hoyal Cuthill and Conway Morris 2014), with large numbers of arboreomorphs (Laflamme and Narbonne 2008) also present (Clapham et al. 2003; Wilby et al. 2011). Phylogenetic analyses have resolved rangeomorphs as eumetazoans (Dunn et al. 2021) with other taxonomically resolved taxa, including crown-group cnidarians (Liu et al. 2014; Dunn et al. 2022) and putative sponges (Sperling et al. 2011; Suarez and Leys 2022). In these early Ediacaran assemblages, namely the Lantian and Avalon, there is no evidence for mobility within animal communities. Putative cnidarian trails have been found in nearby strata (Liu et al. 2010), but these may instead be microbial aggregates (Mariotti et al. 2016; Warren et al. 2020).

Feeding modes for these sessile, benthic organisms are hard to establish, because comparisons with extant sessile benthic organisms such as sponges and sea pens are limited (Antcliffe and Brasier 2007; Laflamme et al. 2009). The widespread presence of rangeomorphs in deep-water strata below the photic zone demonstrates these sessile organisms were not algae or photosynthetic (Benus et al. 1988; Wood et al. 2003). Instead, the fractal-style branching of rangeomorphs (Hoyal Cuthill and Conway Morris 2014) suggests utilization of resources from the water column (Narbonne 2004, 2005). This feeding could be through suspension or filter feeding, as is seen with extant sessile benthic animals, or osmotrophy like giant bacteria and other microorganisms. As rangeomorphs exhibit surface area to volume ratios similar to those of extant osmotrophs, and no feeding apparatus for suspension or filter feeding is preserved at a micron-level scale, this unusual mode of feeding was thought likely (Laflamme et al. 2009). However, suspension feeding is now considered the most likely mode for rangeomorphs, as osmotrophy could not occur at the high Raleigh numbers and advective fluid dynamics that operate at macroscopic scales (Butterfield 2022). Furthermore, if the filter/suspension-feeding apparatuses were sufficiently small (<8 microns), they might not have been preserved (Riisgård and Manríquez 1997), especially due to the preferential loss of labile tissue (Gibson et al. 2018). Finally, rangeomorphs display growth patterns that put them within Eumetazoans (Dunn et al. 2021), where osmotrophy is unknown. CFD simulations have lent further support for suspension feeding in Ediacaran organisms. Tribrachidium and Ernietta are now interpreted as a suspension feeders (Rahman et al. 2015a; Gibson et al. 2021), whereas the rangeomorph Pectinifrons shows feeding modes inconsistent with suspension feeding, osmotrophy, and carnivory (e.g., the carnivorous sponges Lyra; Darroch et al. 2023a).

The passive uptake of food from the water (by osmotrophy or passive suspension feeding) can be supplemented by making the most of canopy flow (Ghisalberti et al. 2014), but these are not active feeding modes, as they do not expend energy in order to gain further energy. This contrasts with filter feeding, the active pumping of water through feeding apparatuses. Before 559 Ma, the putative sponge *Thectardis* (572 Ma) is the only organism for which filter feeding has been proposed. Support for this feeding mode is based on length/width ratios and comparisons of the morphology of Cambrian and extant sponge oscula and surface area of flattened specimens with those of *Thectardis* (Sperling et al. 2011). As such, before 559 Ma, known life modes are limited to nonmobile, passive feeders attached to the substrate (Bush et al. 2011).

## Origins of Mobility and Morphological Diversification: White Sea Assemblage (558–550 Ma)

Taxonomic, morphological, and functional diversity increases dramatically ~555 Ma during the Avalon explosion or Ediacaran second wave (Shen et al. 2008; Droser and Gehling 2015; Evans et al. 2018), with key localities for this time period in South Australia and the White Sea Region, Russia (Boag et al. 2016). In contrast to the deep-water Avalon assemblage, the White Sea assemblage records a shallow-water environment (Grazhdankin 2004; Tarhan et al. 2015). Phototrophs in the form of macroalgae are present within some communities (Wang et al. 2014; Droser and Gehling 2015; Xiao et al. 2020), as are the dominant Avalon taxa, rangeomorphs and arboreomorphs, but in reduced diversity and abundance (Evans et al. 2018). Communities of the tubular organism Funisia reach extremely high densities-more than 6000 specimens/m<sup>2</sup> (Suprenaut et al. 2020), likely due to reproductive spawning akin to that of corals (Droser and Gehling 2008). Besides these frondose organisms, the White Sea assemblage includes matsticking organisms with a range of symmetries not present today, such as triradial and tetraradial (Xiao and Laflamme 2009), as well as facultative mobile organisms in their colonization life stage (Darroch et al. 2017; Coutts et al. 2018). Tissues diversified, with hard tissues functioning as structural supports in the form of long (up to 37 cm) spicules in association with a cone (Serezhnikova and Ivantsov 2007; Clites et al. 2012). During this Ediacaran second wave, there is also a diversification of bilaterians (Evans et al. 2018), including mobile taxa (Glaessner 1959; Fedonkin and Waggoner 1997) and a corresponding increase in ichnodiversity (Mángano and Buatois 2020). Some bilaterians are interpreted to have burrowed in and out of the microbial mats (Xiao et al. 2019), underneath them (Ivantsov et al. 2019), and through them (Evans et al. 2019). It is also around 555 Ma that within-substrate habitats become well occupied with erniettomorphs (Glaessner 1959; Ivantsov et al. 2016). Mat grazing by Kimberella (Fedonkin and Waggoner 1997) is complemented by trails of Yorgia and Dickinsonia that suggest they fed directly on the microbial mat (Ivantsov and Malakhovskaya 2002; Gehling et al. 2005; Ivantsov et al. 2019). Trails that wind underneath decaying body fossils suggest the first occurrences of scavenging (Gehling and Droser 2018). The irregular morphology of Attenborites, in contrast to most other Ediacaran taxa, suggests that it settled out of the water column and was pelagic (Droser et al. 2020). Thus, the White Sea assemblage records a radiation of life habitats, feeding, and mobility.

Bridging the White Sea assemblage (558–550 Ma) and the terminal Ediacaran assemblage, the Nama (550–539 Ma), is the Shibantan assemblage (551–543 Ma) (Condon et al. 2005; Huang et al. 2020). In common with the South Australian and Russian sites, Shibantan communities include rangeomorphs, arboreomorphs, bilaterians, and macroalgae, with an increased abundance of ichnofossils as well as erniettomorphs (Xiao et al. 2021). What is notably different between the White Sea and South Australia localities and the Shibantan is the abundance of trace fossils, for which the Shibantan has a consistently high proportion of trace fossils that dominate many communities (Xiao et al. 2021), some in association with complex body fossils of bilobed, segmented bilaterians (Chen et al. 2019).

## Advent of Biomineralization: Nama Assemblage and the Terminal Ediacaran (550–539 Ma)

Crucially, the Nama assemblage is where abundant biomineralizers are first found forming metazoan reefs (Hofmann and Mountjoy 2001; Wood et al. 2003; Cortijo et al. 2010; Warren et al. 2011; Becker-Kerber et al. 2017). These reefs capture the first evidence of macropredation in the form of drill holes in the reef-building Cloudina (Hua et al. 2003). As well as reef complexity, increasing behavioral complexity is observed within ichnodiversity in the form of more complex and 3D burrows of treptichnids (Jensen et al. 2000), as well as evidence of sediment bulldozing Parapsammichnites (Buatois et al. 2018). Soft-bodied organisms such as rangeomorphs and erniettomorphs lived attached to and within the substrate (Pflug 1966; Meyer et al. 2014), but mat stickers are not found, and soft-bodied mobile taxa are not preserved (Boag et al. 2016). Thus, at the end of the Ediacaran, we have occupation of pelagic, erect, surficial, semi-infaunal, and shallow infaunal habitat occupation, the full complement of feeding strategies, from suspension feeding, deposit feeding, mining, grazing, and predatory modes with a range of mobility from sessile attached to fully mobile.

# Digging Deeper, Building Bigger: The Terreneuvian (539–521 Ma)

Microbial mats persisted into the Terreneuvian, with evidence for mat grazers, scratchers, and miners continuing into the Fortunian (Buatois et al. 2014). Over the 20 Myr of the Terreneuvian, the maximum depth of burrows increased dramatically (from ~6 to 100 cm), and a range of hitherto unrecorded behaviors and structures can be observed in the ichnofossil record, including penetrative feeding traces, farming burrows, and domiciles for suspension feeders (Buatois et al. 2020). Archaeocyath reefs of active suspension feeders emerged during the early Terreneuvian, ca. 535 Ma, initially as small patch reefs in shallow-water environments of the Siberian Platform (Zhuravlev et al. 2015; Antcliffe et al. 2019; Gibson et al. 2023). The Terreneuvian also records the first appearance of a new group of metazoan predators-chaetognaths--into the pelagic realm (Szaniawski 1983, 2002; Vannier 2007) and the earliest evidence for durophagous predation (Bengtson 1968; Conway Morris and Bengtson 1994), while the number of predatory drill holes-and diversity of groups with them-increased (Bicknell and Paterson 2018; Vinn 2018).

## Radiation of Feeding Modes and Mobility: Cambrian Epoch 2 and Beyond (Post-521 Ma)

Ichnodiversity, disparity, and the modes of life represented in the trace fossil record increased again in Cambrian Epoch 2, across siliciclastic and carbonate environments (Buatois et al. 2020), complemented by a huge array of morphological innovations across the sensory, trophic, and movement spectra, as well as an overall increase in the maximum size of animals. Archaeocyath

diversity and geographic spread increased, and meter-scale reefs appear in the fossil record, with different biohermal communities and structures reflecting the latitudes and depths of their locations (Gandin and Debrenne 2010; Zhuravlev and Wood 2018).

The widespread appearance of sensory organs (antennae, eyes, palps) in animals from Cambrian deposits of Epoch 2 and younger, and corresponding increases in brain complexity evidenced by exceptional fossils of nervous tissues (Ma et al. 2012b; Tanaka et al. 2013; Cong et al. 2014; Ortega-Hernández et al. 2019, 2022; Parry and Caron 2019; Parry et al. 2021; Wolfe and Ortega-Hernández 2021) has been termed the "Cambrian information revolution" (Plotnick et al. 2010), due to the sharp increase in the ability of organisms to sense and process their environment. Eyes are best known, most abundant, and most disparate in euarthropods (Zhao et al. 2013), but originated across a range of phyla in animals occupying very different niches. The oldest eyes in the fossil record-focal apposition eyes in the trilobite Schmidtiellus-are from the base of Stage 3 (Schoenemann et al. 2017). Some total-group euarthropod compound eyes contained thousands to tens of thousands of lenses (Zhao et al. 2013; Paterson et al. 2020), with variation in eye morphology indicating specializations for different depths and feeding ecologies, including mesopelagic suspension feeders (Schoenemann and Clarkson 2011; Paterson et al. 2020). Camera-type eyes have been reported in the mollusk Nectocaris (Smith 2013), with simpler eyes and eye spots recorded in annelids (J. Zhao et al. 2023), chordates (Shu et al. 2003), lobopodians (Ma et al. 2012a), and even priapulids (Schoenemann 2006). Beyond eyes, metazoan sensory structures first recorded at this time include antennulae and antennae in total-group euarthropods (Ma et al. 2012b; Ortega-Hernández et al. 2018; Zhai et al. 2019a, 2019b), and annelid palps (Eibye-Jacobsen 2004; J. Liu et al. 2015; Nanglu and Caron 2018; Parry and Caron 2019).

The first large active swimmers are also found in Cambrian Stage 3 deposits, adding multiple trophic levels to the pelagic realm. The largest swimming animals in Cambrian oceans were stem-group euarthropods such as radiodonts, predators that could reach up to 1 m in length at this time (Potin and Daley 2023; Wu et al. 2024), and stem-group chaetognaths, reaching up to about 30 cm (Park et al. 2024). Adaptations for active swimming can be seen across a range of groups, including specialized swimming flaps, fins, and streamlined fusiform bodies (Whittington and Briggs 1985; Chen et al. 1995; Shu et al. 1999; Vannier and Chen 2000; Usami 2006; Daley et al. 2009; Vannier et al. 2009; Conway Morris and Caron 2012; Smith 2013; Daley and Edgecombe 2014; J. Liu et al. 2015; Lerosey-Aubril and Pates 2018; Izquierdo-López and Caron 2019; Pates et al. 2021a; Lerosey-Aubril and Ortega-Hernández 2022; Potin and Daley 2023). Closer to the seafloor, specialized walking appendages facilitated more rapid movement for vagrant epifaunal animals (Minter et al. 2011).

The Cambrian Epoch 2 also saw increased specialization in feeding appendages across groups, in particular within euarthropods. Multisegmented appendages for raptorial predation (Briggs 1979; Daley and Budd 2010; Daley and Edgecombe 2014; Jiao et al. 2021; Potin and Daley 2023), sediment sifting and sweep feeding (Daley et al. 2013; Yang et al. 2013; Moysiuk and Caron 2019; Potin and Daley 2023), and durophagy (Pratt 1998; Haug et al. 2012) are found across a number of total-group euarthropods, complemented by robust gnathobases, gnathobase-like structures, mandibles, and mandible-like structures (Harvey and Butterfield 2008, 2022; Zacaï et al. 2016; Aria and Caron 2017a,

b; Cong et al. 2017, 2018; Vannier et al. 2018; Yang et al. 2018; Bicknell et al. 2021; Moysiuk and Caron 2021).

Computational fluid dynamics analyses support both active pharangeal filter feeding and passive suspension feeding in Cambrian echinoderms (Rahman et al. 2015b, 2020), while numerous groups display feeding apparatuses with rows of finely spaced spines, setae, cilia, or comb-like elements, suitable for suspension or filter feeding. These include total-group euarthropods (Vinther et al. 2014), luolishaniids (Ma et al. 2009; Yang et al. 2015; Caron and Aria 2017; Howard et al. 2020), hemichordates (Nanglu et al. 2016, 2020a), and the problematica *Siphausauctum* (O'Brien and Caron 2012) and dinomischids (Ou et al. 2017; Y. Zhao et al. 2023). This time period also sees the first documentations of brood care and extended parental care in the fossil record (Caron and Vannier 2016; Fu et al. 2018; Ou et al. 2020; Ma et al. 2023), demonstrating a diversification of behavioral ecology, as well as trophic ecology.

#### Summary of Organismal Ecology over the ECT

A fundamental shift in organismal ecology occurred over the ECT. Before the Ediacaran, macro-organisms were exclusively primary producers, such as algae, and all heterotrophic organisms were microbial. The earliest animals still fed, like their microbial counterparts, on microscopic life such as plankton and dissolved organic carbon (DOC), particulate organic matter (POM) and particulate organic carbon (POC) in the water column. Key innovations such as large body size, coupled with differentiated tissues likely enabled further diversification of feeding modes, and the expansion to grazing, scavenging, and indeed predation. Coupled to this trophic diversification was the advent of mobility, facilitating access to nutrients that were becoming increasingly concentrated on the seafloor. Yet, despite the origination of mobility and predation in the Ediacaran, it is not until Cambrian Epoch 2 that specializations really start to intensify. The specializations and variation in form of feeding appendages, sensory equipment, movement types, and burrowing employed by a diverse range of animal groups set the Cambrian Epoch 2 apart from the Terreneuvian and Ediacaran and laid the groundwork for the rest of the Phanerozoic.

#### **Community Ecology**

#### Dominant Ecological Modes within Communities

The simplest way to consider the community ecology of an ecosystem is in terms of the relative abundance of ecological modes-that is, what is the primary niche of the community, and how are niches distributed within it? The oldest Ediacaran communities, the Lantian assemblage (~600 Ma), are dominated by algae with a few putative cnidarians (Yuan et al. 2013), so were mainly autotrophic with some possible suspension feeders (Wan et al. 2016). The majority of communities before 560 Ma, from Newfoundland, Canada, and Charnwood Forest, U.K. (Clapham et al. 2003; Wilby et al. 2011), only have suspension-feeding organisms, rendering them ecologically monotypic. Ecological diversification of relative abundance of ecological modes starts to occur around 550 Ma, with communities from Nilpena, South Australia (Droser et al. 2019), and the White Sea, Russia (Zakrevskaya 2014), hosting communities with only suspension feeders (cf. Funisia beds; Droser and Gehling 2008), as well as more ecologically diverse communities with algae, suspension

feeders, scavengers (Gehling and Droser 2018), and microbial mat feeders (both under and on top of the mat) (Ivantsov and Malakhovskaya 2002; Droser et al. 2019; Evans et al. 2019; Fig. 3). Less is known about the period between 550 Ma and 539 Ma, as fewer communities are preserved in situ, but the Shibantan (551-543 Ma) records a diversity of complex mobile animals such as Yilingia (Chen et al. 2019) and ichnofossils as well as a diversity of suspension feeders and macroalgae (Xiao et al. 2021). The relative abundances of Shibantan communities are not (yet) known, but from the diversity of organisms, appear ecologically similar to White Sea assemblage communities. Note, that because the feeding mode of some complex, endemic species such as Yilingia are unresolved, there could be as yet unreported additional feeding complexity. As in older (~555 Ma) communities, the terminal Ediacaran (550-539 Ma) has a plethora of suspension-feeding tubular organisms preserved alongside trace fossils, but biomineralized forms dominate (Becker-Kerber et al. 2017, 2020; Warren et al. 2023), and the relative proportions of predators are not known (Hua et al. 2003). These suspension feeding-dominated communities form two distinct community types. In Namibia, Brazil, and Paraguay, Cloudina and Namaclathus reefs dominate (Hofmann and Mountjoy 2001; Grotzinger et al. 2005; Warren et al. 2011; Penny et al. 2014; Becker-Kerber et al. 2017). Soft-bodied rangeomorphs and erniettomorphs are also present, occupying different settings than the reefs (Pflug 1966; Narbonne et al. 1997; Grazhdankin and Seilacher 2002) and are found mainly in transported deposits (Bowyer et al. 2017).

Less is known about the Terreneuvian, due to the lack of deposits preserving exceptional soft tissues (see "Digging Deeper, Building Bigger"), so ecological dynamics cannot be resolved until the Cambrian Epoch 2. Deposits of this age and younger, for which the Chengjiang and Burgess Shale have been studied in most detail, show a variety of different dominant ecological modes. Chengjiang communities (518 Ma), are dominated by carnivorous taxa (mainly polychaetes) (Zhao et al. 2010, 2014), similar to near-contemporaneous Sirius Passet communities (Harper et al. 2019). In contrast, Burgess Shale localities (506 Ma; Fig. 3) have a mixture of deposit feeding- (Walcott Quarry) or suspension feeding-dominated communities (Tulip Beds) (O'Brien and Caron 2016), although it is important to note that recent flume experiments and reanalysis of sedimentary structures suggest that organisms from the classic Walcott locality in the Burgess Shale may have been transported large distances (Bath Enright et al. 2021). When the localities are sampled at a finer scale (10 cm sample intervals), a more complex pattern emerges, with four community types: suspension feeding, hunting/scavenging, deposit feeding, and nektobenthic-vagrant-deposit feeding (Nanglu et al. 2020b), with a more general Cambrian Stage 3 pattern of the four Burgess Shale-type community types (Hou et al. 2017; Fu et al. 2019) alongside those dominated by archaeocyath reefs (Sun et al. 2020).

While the diversification of feeding modes throughout the ECT is high, the innovation of predation-dominated systems does not lead to a unidirectional change in ecosystem functioning, but more a broadening of different sorts of relative abundances of ecological niches across a spectrum. This point is illustrated by the notable overlap that occurs in the ranking of Nilpena Ediacaran communities (Droser and Gehling 2015; Droser et al. 2019) with Burgess Shale Cambrian communities (Nanglu et al. 2020b) when ordered by proportion of suspension feeding (Fig. 3).

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## Abiotic and Non-trophic Biotic Interactions within Communities

The relative abundances of different ecological groups within each community are a result of the interactions of taxa within the community, and how they interact with the local resources/environment. The environmental impacts on organisms, and the feedback relationships between organisms and their abiotic environment, are dependent on the relative needs of the organisms and the extent of abiotic nutrients. Organisms adapt to their environments to optimize their efficiency, for example, rangeomorphs maximized their surface area to volume ratios in order to optimize nutrient/resource exchange with the water column (Narbonne 2004; Laflamme et al. 2009). These abiotic impacts can, and most commonly do, include organisms only existing in suitable habitats-these can be very clear, such as only organisms with good cold tolerance existing at high latitudes, or more subtle, such as trees having higher densities in regions of higher soil nutrients. These habitat associations and influences can be detected through a variety of different methods. In the Ediacaran, spatial point process analyses have been used to detect increased densities where taxa (or pairs of taxa) exhibit higher densities in areas of good-quality patchy habitat (Baddeley and Turner 2005; Illian et al. 2008; Wiegand and Moloney 2013; Mitchell et al. 2020). Furthermore, these increased densities or aggregations can be statistically distinguished from other aggregation processes such as dispersal limitation and reproductive processes and facilitation. These methods have shown that environmental, or niche, influences are limited from 572 to 560 Ma (Mitchell and Kenchington 2018; Mitchell et al. 2019), and increase from 560 to 550 Ma (Mitchell et al. 2020). Preferential settlement (Boan et al. 2023) with community composition corresponding to different microbial mat maturity/complexity (Droser et al. 2022) demonstrates the importance of local environment to communities. In the terminal Ediacaran (550-539 Ma), the reef builders Cloudina, Corumbellina, and Namacalathus show clear associations with biolaminites such stromatolites and thrombolites (Hofmann and Mountjoy 2001; Grotzinger et al. 2005; Warren et al. 2011; Cai et al. 2014), reflecting either biotic interactions and/or mutual habitat associations.

The importance of local habitat patchiness increases into the Cambrian, as can be seen through the trace fossil record, through an increase in sediment bulldozing into the Fortunian as sediment feeders diversified (Mángano and Buatois 2014; Buatois et al. 2018) and where the complexity of interactions of ichnotaxa with patchiness increases (Mitchell et al. 2022). As the depth and complexity of burrowing increased into Cambrian Stage 2, with the onset of mix-ground ecology and suspension feeding from within the sediment (Mángano and Buatois 2014), these subtle differences of substrate local patchiness likely decreased in importance, because large-scale and more significant habitat variations took over, and the benthic realm became more akin to the modern, with other factors becoming a first-order process of habitat variation, rather than subtle differences in microbial mat or sediment variations.

Biotic interactions, that is, the interactions between two taxa, consist of interactions such as competition, facilitation and mutualisms, and trophic interactions. For the oldest Ediacaran communities, there is limited evidence of competition for resources, with only rare intra- and interspecific competition (Mitchell and Butterfield 2018; Mitchell and Kenchington 2018). As the extent of intra- and interspecific competition increases throughout the

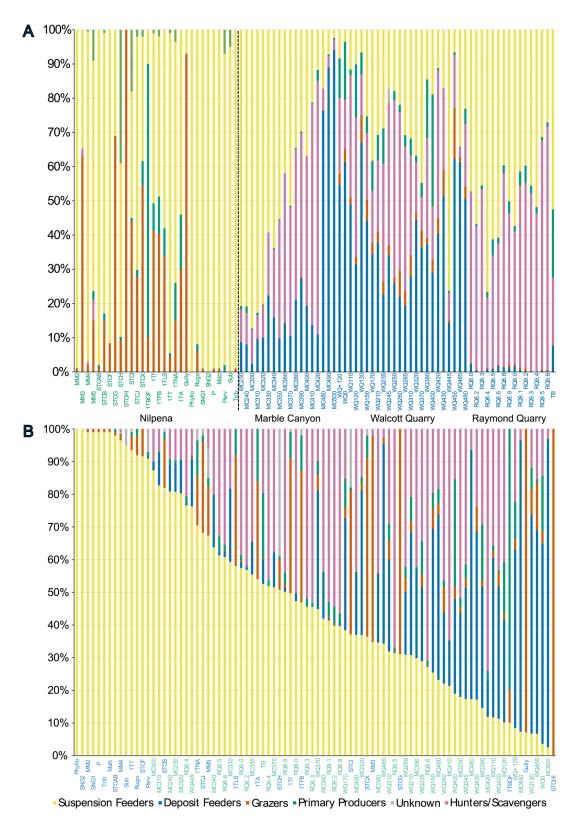


Figure 3. The relative proportions of feeding type in Ediacaran (green) from Nilpena, South Australia (Droser and Gehling 2015; Droser et al. 2019) and Cambrian (blue) from the Burgess Shale, Canada (Nanglu et al. 2020b). Feeding types are suspension feeding (yellow), deposit feeding (blue), grazing (orange), primary production (green), hunters and scavengers (red), and unknown in gray. The top plot is ordered temporally, and the bottom plot is ordered by relative proportion of suspension feeders from left to right. Note that while there is a trend of Ediacaran on the left to Cambrian on the right, there is a significant overlap between them.

Ediacaran (Droser et al. 2017), coupled to an increase in relative importance of niche/environment interactions (Mitchell et al. 2020; Droser et al. 2022), so does the complexity of interactions, with facilitation first detected in the terminal Ediacaran in terms of preferred settling near favorable fluid dynamics flow (Gibson et al. 2021).

Complexity of types of ecological interactions increases in the Cambrian, with symbiosis (Vinn 2017), parasitism (Peel 2015; Li et al. 2020; De Baets et al. 2021; Chen et al. 2022), mutualisms and commensalism (Topper et al. 2014; Peel 2015; Nanglu and Caron 2021), and epibionts (Zhang et al. 2009; Topper et al. 2015) all present. Yet, in terms of observed competition within Cambrian ecosystems, there is only limited evidence of its impact. Competitive exclusion is extremely rare in Phanerozoic benthic marine ecosystems (Klompmaker and Finnegan 2018), likely due to the higher levels of bioturbation, predation, and disturbance preventing ecosystems from reaching carrying capacity (Stanley 2008; Klompmaker and Finnegan 2018). An exception is possible in the case of archaeocyath reefs. Distorted specimens likely resulted from competition for space (Zhuravlev and Wood 1995), in an ecosystem lacking known predators and-due to its biomineralized nature-less prone to environmental disturbance. Even then, both niche and neutral processes appear to have played a role in these early sponge reef ecosystems (Zhuravlev et al. 2015).

#### **Trophic Interactions**

Trophic interactions, namely the interactions that occur between an organism and its food source, have been suggested to be a key driver of the Cambrian Explosion, with predators and their prey driving the morphological changes and diversity seen during the Cambrian (Bengtson 2002). Trophic complexity started relatively simply (Fig. 4), with early Ediacaran organisms feeding primarily/exclusively on plankton and POM/POC from the water column (Laflamme et al. 2009; Butterfield 2022). New trophic links formed between organisms and the microbial mat, with Dickinsonia and Yorgia feeding on it (Ivantsov and Malakhovskaya 2002; Sperling and Vinther 2010; Ivantsov 2011; Evans et al. 2019; Ivantsov et al. 2019; Ivantsov and Zakrevskaya 2021; Bobrovskiy et al. 2022), and with the origination of the first scavengers (Gehling and Droser 2018; Fig. 4). There is no evidence of pelagic macroscopic predation until the terminal Ediacaran (Hua et al. 2003). Further insights into the extent of resource niche overlap found that the majority of Ediacaran communities from 572 to 560 Ma had shared niches, so likely shared food sources; niche complexity increased with complex/partitioned niches within communities during 560-550 Ma, then decreased with simpler/shared niches in 550-540 Ma (Darroch et al. 2018a). These data likely reflect an increase in trophic complexity around 555 Ma, potentially followed by a decrease in the terminal Ediacaran.

These matground-dominated dynamics persisted into the early Cambrian (Fortunian), with a shift in trophic dynamics starting in Cambrian Age 2, with a similar style of trophic dynamics to the modern present in Age 3 (Cambrian Epoch 2) (X. Zhang et al. 2021; Fig. 4). There is a distinct shift in the importance of biotic interactions within Cambrian food webs from Age 3 and younger, that is, with trophic interactions, where we see a sharp increase in the functional morphology of feeding apparatuses, predatory apparatuses, and defensive tools (see "Radiation of Feeding Modes and Mobility"). Initially it was thought these trophic chains were relatively short, with only

primary predators (Vannier and Chen 2005), but more recent work has suggested that there can be relatively long trophic chains four to five species long (Hu et al. 2007; Dunne et al. 2008). Such complex chains are not necessarily present in all Cambrian ecosystems, such as those dominated by suspension feeders or detritovores (cf. O'Brien and Caron 2016; Fig. 3).

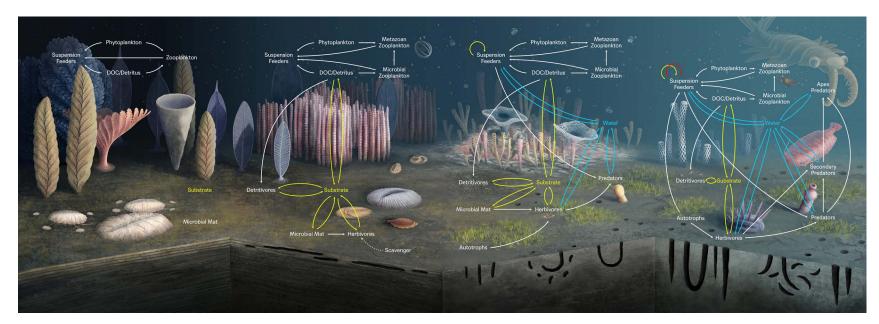
All these trophic interactions translate into relatively complex trophic networks (or food webs), with food webs of the Burgess Shale and Chengjiang displaying topologies similar to those of modern ones in terms of distributions of feeding links (Dunne et al. 2008). These Cambrian food webs show a dependence of their structure on the number of taxa and feeding links, which is found in modern webs modeled by simple "niche models." However, the Chengjiang web has long paths (feeding chains) and more feeding loops than modern webs. One cause of this is that the prey species with the most predator links have many more predators than expected from modern webs, perhaps due to a lack of specialized defense mechanisms. Both Burgess Shale and Chengjiang food webs show a higher variation in the number of trophic links per taxa than is seen in the modern. These results are thought to suggest that as ecosystems evolved after the Cambrian, the trophic structure became more constrained (Dunne et al. 2008).

#### Ecosystem Resilience and Complexity

For ecosystems and the organisms within them to persist over geological timescales, these ecosystems must be stable, that is, have resilience. Ecosystem stability is strongly correlated to taxonomic and functional diversity, because more diverse systems have a better capacity to cope with biotic and abiotic changes and fluctuations through feedbacks within the network of interactions (abiotic, biotic, and trophic) within the community (May 1974; Tilman 1994; McCann 2000). These feedbacks and redundancy mean that the more complex an ecosystem is, the more stable it is (Ruiter et al. 1995; Neutel et al. 2007), in contrast to theoretical (random) networks, where complex networks are less stable (May 1974, 2019).

Ecological complexity is most simply measured in terms of taxonomic diversity, with more diversity corresponding to more stable ecosystems (Ives and Carpenter 2007). However, ecological stability is not as straightforward as just taxonomic diversity stabilizing the ecosystems. If there is a single species that is crucial for ecosystem function/stability, then the system is vulnerable (Paine 1969; Jain and Krishna 2002). As such, communities in which many species occupy similar niches tend to be more stable, so that if a specialist predator or pathogen or habitat changes, other species can fill that ecological role (Petchey and Gaston 2006). The capacity for ecosystems to have this redundancy is most simply measured using evenness (Shannon 1948), that is, the distribution of relative abundances of taxa within an community. If the community is very skewed, and so has a few very highabundance taxa and many rarer taxa, the system is likely much more sensitive to perturbations compared with communities that are more even (Hillebrand et al. 2008).

Over the ECT, some Ediacaran communities exhibit relative high evenness, corresponding to many relatively abundant taxa and complex niches, alongside simpler, more monospecific communities (Darroch et al. 2018a). As such, it is likely that Ediacaran ecosystems were not all equally resilient. The stabilizing mechanisms for this resilience can vary, with systems stabilized primarily through primary production availability (bottom-up), and



**Figure 4.** Ecological networks across the Ediacaran/Cambrian transition (ECT), with four time periods illustrated (from left to right: 565 Ma, 539–521 Ma, 521 Ma). Suspension-feeding communities and simple trophic interactions (white arrows) are present 565 Ma. An increase in habitat modification (yellow arrows) in the form of grazing and scavenging appear 550 Ma. In the Terreneuvian (539–521 Ma), the active pumping of archaeocyaths and swimming of chaetognaths increase aquatic bioturbation (blue arrows), archaeocyath reefs modify the substrate, and vertical as well as horizontal bioturbation complexity and depth increase dramatically. In the Cambrian Epoch 2 (521 Ma), there is a further increase in trophic interactions, aquatic bioturbation, and vertical bioturbation, as well as symbiotic interactions and epibionts (red arrows). DOC, dissolved organic carbon. Reconstruction by Franz Anthony.

other systems stabilized by predators (top-down). Due to the lack of higher predators and short trophic links (Fig. 4), Ediacaran ecosystems were stabilized from the bottom-up, so dependent on abiotic resources and environmental factors. The lack of resource competition suggests that such resources are not limited (Mitchell and Kenchington 2018; Mitchell et al. 2019), likely making these Ediacaran systems relatively stable. This bottom-up stability likely persists until Cambrian Ages 2 and 3, when some (but not all; see "Dominant Ecological Modes within Communities") systems exhibit many trophic levels, which could potentially lead to top-down stabilizing mechanism. However, longer feeding chains seen in the Chengjiang are sufficiently long that species may be functionally separated from one another, potentially destabilizing the systems (Dunne et al. 2008). The potential (relative) lack of stability in the Cambrian was also possibly present within Burgess Shale communities, where the Tulip Bed and Walcott Quarry communities have high abundances of a few dominant taxa, with many rarer taxa (O'Brien and Caron 2016).

#### Macroecology

Determining the processes that shape community-level ecological metrics such as diversity and community composition is enhanced through comparisons with other communities, enabling the teasing apart of the relative importance of different temporal and environmental drivers, and how they interact with the organisms at a community level.

#### **Beta Diversity**

Beta diversity is a measure of how much variation there is between communities (Anderson et al. 2011), with high diversity indicating little species overlap and thus different species in different communities, and low beta diversity indicating high levels of overlap and thus similar species are found across different communities. Regions that exhibit high beta diversity are more stochastic, that is, the community compositions are not the systematic result of adaptions to their local environments and biotic interactions, but instead reflect colonization and dispersal dynamics, for example, as seen at modern hydrothermal vents (Giguère and Tunnicliffe 2021). If biotic interactions and environmental filtering are strong drivers of community composition, we would expect the same community compositions to emerge within a given environment, and so beta diversity would be low, for example, in shallow-water reefs (Harborne et al. 2006). As such, quantifying beta diversity can help inform the key factors underlying community composition and development.

Ediacaran beta diversity is significantly higher than that of the Cambrian and the rest of the Phanerozoic. This beta-diversity increase has been suggested to be due to the rarity of mobile taxa, which may have increased the patchiness of communities (Finnegan et al. 2019). Strong dispersal limitation, such as is produced via asexual stoloniferous reproduction (Mitchell et al. 2015), provides a complementary cause for community patchiness. Competitive communities can also increase beta diversity (Stanley 2008), but the rarity and weakness of resource competition within Avalonian Ediacaran communities suggest that that niche processes and biotic interactions shape the communities less than in extant systems (Na and Kiessling 2015; Finnegan et al. 2019). As such, high Ediacaran beta diversity may result from processes similar to those in hydrothermal vent systems.

These vent systems often exhibit high beta diversity (Giguère and Tunnicliffe 2021) due to the stochastic nature of the creationsuitable habitat (the vents). Thus, the identity of organisms that reach and colonize the new habitat first are more stochastic than most extent benthic systems. Over the ECT, patterns of beta diversity do not necessarily follow those of alpha diversity. Alpha diversity increases from the Ediacaran through to Cambrian Age 3 (Fig. 2), yet beta diversity decreases into the Fortunian before increasing again through Cambrian Ages 2 and 3. Middle-level beta diversity later in the Cambrian suggests an increase in the influence of these niche and biotic interactions throughout (Zhao et al. 2014). In these first three stages of the Cambrian, alpha/beta-diversity dynamics are consistent with a low-competition dynamics, that is, little pressure on resources (Hautmann 2014). New species appear either in previously unexploited niches, or communities have more species within a single niche, potentially due to predation exerting a top-down control (Na and Kiessling 2015). Beta diversity increases from the Cambrian to the Devonian as resources are reduced, increasing competition and constricting niches (Na and Kiessling 2015; Penny and Kröger 2019). These beta-diversity patterns suggest that stochasticity is key to Ediacaran communities with limited niche and biotic interactions. Niches constrict and biotic interactions increase throughout the Cambrian, feasibly as a result of predation and top-down control.

## Biogeography

Biogeographic patterns require sufficient sample sizes to resolve, which means that inferences over the ECT are limited. Within the Ediacaran, detailed biogeographic analyses are limited to differences between high and low paleolatitudes, due to the number and geographic placement of localities (Boddy et al. 2021). The frondomorph group, which includes rangeomorphs and arboreomorphs, radiated from high to low paleolatitudes, in contrast to algal and protist groups, which showed no paleolatitudual differences (Boddy et al. 2021). Further paleolatitudual differences are found with the bilaterian group, which radiated from low to high latitudes, unlike algal, tubular, soft-bodied, and biomineralizing taxa (Boddy et al. 2021), showing Ediacaran biogeographic differences with taxon distributions through time, even at this coarse spatial scale. The drivers behind these two patterns are not yet resolved, but the frondomorph high to low radiation is likely due to evolutionary processes with a deep-water, highlatitude, origination (Griffiths et al. 2023), while the bilaterian paleolatitude signal could be a reflection of high rates of diversification at the tropics leading to quicker evolution at these lower latitudes (Mittelbach et al. 2007).

In the Cambrian, the presence and strength of biogeographic biodiversity patterns increased, showing increase in provinciality, or bioregions, coupled to a decrease in geographic ranges (Hendricks et al. 2008; Na et al. 2023). This increase in biogeographic patterns has been suggested to relate to several factors, including the geographic differences in origination of groups (Lieberman 2002, 2008; Lieberman and Meert 2004); changes from low-competition to high-competition models with a change from wide to narrower geographic ranges (Na and Kiessling 2015); differences within groups, relating to dispersal abilities (e.g., with soft-bodied, pelagic Cambrian arthropods showing greater geographic ranges than trilobites; Hendricks et al. 2008); and more broadly to biogeographic differentiation at regional and local scales (Na et al. 2023). However, in some cases, for example, the trilobites, their origination within the Cambrian (Paterson et al. 2019) does not correlate with the breakup of Pannotia tens of millions of years earlier, leaving many patterns still to be explained.

#### Metacommunity Ecology

The many different types of ECT communities (see "Dominant Ecological Modes within Communities") could be due to evolutionary/temporal environmental filtering, and/or the result of different biotic interactions. Cluster analyses based on community composition can shed led on these drivers, because if environment dominates, we expect the groups of communities that share similar composition to have lived in similar environments. If the communities represent a progressive evolution, then the community groups of similar geological ages should cluster together (Anderson 1971). Once different groups of communities, or metacommunities, have been established, the metacommunity structure within these groups can shed further light on the driving forces within them, elucidating differences in environmental and biotic drivers and the relative importance of stochasticity between metacommunities (Leibold et al. 2004).

The three classic assemblages of the Ediacaran-Avalon, White Sea, and Nama-as first proposed by Waggoner in 2003 occupied distinct temporal, environmental as well as species-specific assemblages (Waggoner 2003), with a direct reading suggesting an evolutionary radiation from deep-water, highlatitude sites. Since then, the number of sites has increased dramatically from 15 in Waggoner's initial analyses to 127, with more overlap between environments and time, yet these three key assemblages (or biozones) remain robust when examined using a variety of different clustering and network approaches (Boag et al. 2016; Muscente et al. 2019; Boddy et al. 2021), so are likely fundamental to the Ediacaran. Because the environmental and temporal range of the Ediacaran localities are largely (but not totally) non-overlapping, it is not possible to determine the extent to which the differences between the oldest, Avalon assemblage and the much more diverse White Sea assemblage are evolutionary or environmental (Grazhdankin 2004; Boag et al. 2016). As work on other sites, such as the Lantian, Shibantan, and Corumbá localities increases, it possible that additional assemblages may emerge, such as with the Shibantan (Xiao et al. 2021), with the potential of overlapping temporal and species ranges of the new data enabling a teasing apart of environmental versus evolutionary trends.

Ediacaran diversity increases between the Avalon and White Sea, then decreases between the White Sea and Nama, which at a global scale is not due to sampling differences (Muscente et al. 2019; Boddy et al. 2021). This diversity decrease has been suggested to be caused by a catastrophic mass extinction (Darroch et al. 2018b, 2023b; Zhuravlev and Wood 2018; Muscente et al. 2019; Evans et al. 2022); change in taphonomic window (Laflamme et al. 2013); or a biotic replacement model, whereby Phanerozoic-type animals outcompeted Ediacaran-type animals, driving them extinct (Darroch et al. 2015), potentially through "wormworld" fauna (Schiffbauer et al. 2016). This diversity decrease is robust to sampling differences on a global scale (Muscente et al. 2019); however, there are paleolatitudinal differences in sampling biases, with low-latitude diversity not significantly different from the number of low-latitude localities, and high-latitude decoupled with only weak significance (Boddy et al. 2021). This latitudinal difference suggests that some (but not all) of the drop in diversity in the terminal Ediacaran may be an artifact of sampling biases. Further differences in diversity could be due to internal restructuring: analyses of the metacommunity structure between the three large assemblages suggest diversity differences between the White Sea and Nama assemblages are more likely due to internal ecological restructuring, because there is an increase in turnover, a maintenance/increase in significant pairwise taxon associations, and an increase in depth specialization, which are all inconsistent with a catastrophic mass extinction (Eden et al. 2022). As such, biological (within-organism) and ecological complexity likely increased throughout the Ediacaran.

In the Cambrian, the paleocommunity data from contemporaneous Cambrian sites within different paleoenvironments enable the teasing apart of niche partitioning (Na et al. 2023). However, Lagerstätten with soft-bodied preservation that are sufficiently sampled to be able to make ecological inferences are relatively constrained to an ~20 Myr between 518 and 499 Ma, which limits the ability to infer evolutionary or temporal trends from the data. In addition, the vast majority of these Lagerstätten are found in the paleotropics. Nonetheless, temporal trends are observable in some cases, such as in echinoderms, where the differences in their presence between the Chengjiang and Burgess Shale localities could be evolutionary (O'Brien and Caron 2016). Temporal trends can also be detected at a fine-scale resolution through collection of data from narrow sample intervals of Burgess Shale communities (10 cm, ~600 yr Nanglu et al. 2020b). These fine-scale analyses reveal periods of relative ecological stability, followed by a turnover in community composition, suggesting periodic environmental disturbances and then recolonization (Caron and Jackson 2008), with similar community composition between successive sample intervals. Temporal trends also changed Burgess Shale community compositions, with the youngest communities (Tulip Bed) lacking hemichordates and annelids, which are abundant in the older localities. One such suggestion for these changes is competitive exclusion within a niche, for example, of sponges by hyoliths and hemichordates (Nanglu et al. 2016, 2020b; Moysiuk et al. 2017).

The relative effects of environmental filtering (i.e., different taxa living within different environments) varies in the Cambrian. In the House Range (Utah), the higher diversity of pelagic animals in the Marjum Formation compared with the slightly older Wheeler Formation was suggested to be due to the deepening of the environment in the Marjum Pass area of the Great Basin and/or changing regional ocean circulation (Pates et al. 2021b). Older Chengjiang communities (518 Ma) also show evidence of environmental filtering with a gradient from lower diversity nearshore to higher diversity offshore (Zhao et al. 2012), and a high facies control on community compositions (Sun et al. 2020), but such signals are not present in Burgess Shale data (O'Brien and Caron 2016).

Other differences between the Chengjiang and Burgess Shale localities include differences in specialization. In contrast to Chengjiang taxa, dominant Burgess Shale taxa show broad stratigraphic and geographic ranges, with the four different community types not split by locality or environment, but instead varying at local and regional scales (Nanglu et al. 2020b). The reoccurrence of the abundant taxa within Walcott Quarry communities suggests that these abundant taxa are likely environmental and/or geographic generalists, with the majority of abundant Walcott Quarry taxa also present in Tulip Beds, albeit as rare taxa (e.g., *Marella*) (Caron and Jackson 2008). There are signals of localization, because many of the taxa from the Walcott Quarry and Tulip Beds occur only within a single horizon, suggesting that some taxa are adapted to their local environments, but not as well adapted to the wider regional environment (O'Brien and Caron 2016). Note that some of these differences could be taphonomic or due to transportation, as the mobile detritivores that dominate Walcott Quarry communities may have escaped from the small burial events at the Tulip Beds (O'Brien and Caron 2016), and fossils from the classic Burgess Shale quarry may have undergone significant transport (Bath Enright et al. 2021). While these patterns suggest recolonization from a species pool of taxa with good dispersal and/or colonization ability was important, that is, community composition was still relatively stochastic (as suggested by the beta-diversity patterns; Na and Kiessling 2015), the dispersal/recolonization cannot account for all the community differences found, with priapulids and sponges abundant throughout most Cambrian localities, but not at Marble Canyon, suggesting some temporal or environmental filtering (Nanglu et al. 2020b).

There are differences in ecological dominance between Chengjiang and Burgess Shale communities, with Chengjiang dominated by carnivorous taxa (Zhao et al. 2014) in contrast to deposit- and suspension-feeding ecological dominance in Burgess Shale localities (O'Brien and Caron 2016). A comparable signal is seen in the large swimming radiodonts, with raptorial amplectobeluids and anomalocaridids dominant in Chengjiang, but sediment-sifting hurdiids more diverse in the Burgess Shale and Great Basin (Wu et al. 2022). Again, these could be the result of environmental differences, rather than temporal or geographic differences, but to resolve the underlying drivers of these differences requires a series of community-level samples from sites spanning a broad temporal range but limited geographically. Thus, data from Chengjiang and the Burgess Shale could be complemented with similar high-resolution studies from Kaili or the Great Basin respectively.

#### Stochastic to Deterministic Trends

Metacommunity comparisons reveal patterns of change across the ECT. In the Ediacaran, such metacommunity analyses have shown a pattern of increasing specialization not only of organisms, but of communities (Eden et al. 2022). The oldest Ediacaran assemblage, the Avalon, is populated by generalists, with little environmental influence. In the White Sea assemblage, there is increased metacommunity variation, with metacommunity structure dependent on environment and biogeographic differences in the pairwise interactions of taxa. The terminal Ediacaran assemblage, the Nama, shows strong niche segregation (Eden et al. 2022). Patterns of increasing specialization and habitat segregation are also seen in early Cambrian ichnotaxa, which start out as generalists in the Fortunian, but then increase in specialization, leading to habitat segregation in the Cambrian Stage 2 (Meek et al. 2023). However, the overarching change over the ECT from stochasticity populated by generalists to more deterministic, with more specialists is not necessarily a unidirectional process. Cambrian archaeocyath sponge reef communities on the Siberian Platform (525-514 Ma) show changes between individually driven metacommunity structure to within-community interactions and metacommunity complexity as reef habitat expands, and then a subsequent reduction in metacommunity complexity as reef habitat is reduced (Zhuravlev et al. 2022). Coupled to diversity patterns in and between different Cambrian clades (Servais et al.

2023), these patterns demonstrate that while ecological complexity increased, it was not a simple linear process, but instead was likely pulsed and multidirectional (Zhuravlev and Wood 2018) and heterogeneous environmentally and geographically.

#### **Ecosystem Engineering**

Interactions and feedbacks within and between ecosystems can not only stabilize, but also drive ecological dynamics through ecosystem engineering, whereby taxa change their local environment in a way that makes it more habitable for other organisms. The precise definition of ecosystem engineering is much debated: on one side, every living organism changes its environment (Jones et al. 1994), but on the other side, it is not useful to discuss ecosystem engineering in terms of ecosystem changes that do not significantly change and improve the environment for other organisms (Reichman and Seabloom 2002). As such, we will define ecosystem engineering as occurring when organisms have impacted their environments sufficiently to produce observable (positive) impacts on other organisms (cf. Hastings et al. 2007).

There are four main ecosystem engineering mechanisms that have been suggested to contribute to the ECT diversification: (1) Savanna hypothesis, (2) reefs, (3) vertical substrate bioturbation, and (4) aquatic bioturbation. The Savanna hypothesis is the oldest form of metazoan ecosystem engineering, whereby early bilaterian evolution is driven by the search for patchy food sources on the seafloor that come from the decaying remains of sessile organisms (Plotnick et al. 2010; Budd and Jensen 2017). This mechanism persists until Cambrian Stage 3, when multiple different processes, namely pervasive burrowing, detritivores, and scavenging, ensure that such patchy resources do not remain in the sediment long enough to be evolutionary drivers. This hypothesis is consistent with analyses from Ediacaran community ecology. These analyses show an increase in the amount of interactions that Ediacaran organisms had with their environments (Mitchell et al. 2020), increased scavenging (Evans et al. 2019), and increased ichnodisparity (Mángano and Buatois 2014), and the extent and complexity to which trace fossil interact substrate patchiness (Mitchell et al. 2022).

The second group of ECT ecosystem engineers are reefs, which dramatically changed their environment through the creation of hard substrate and complex 3D space. Through this reef creation they created more niches, which in modern systems are highly correlated with diversity, as reef animals use coral structures for food, protection from predation, habitat, and nurseries (Bozec et al. 2013; Brandl et al. 2019). However, when these opportunities within reefs started to be fully utilized is not clear-cut. The Ediacaran reefs of Cloudina, Corumbella, and Namacalathus demonstrate significant ability to build large structures, but there is no direct evidence of facilitative ecological interactions of other animals with these Ediacaran reefs (Warren et al. 2017; Wood et al. 2017), although the extent to which this could be a preservational artifact is not clear, because soft-bodied Ediacaran organisms are not easily preserved within such carbonate settings (Hall et al. 2013). Archaeocyath reefs are abundant from the early Cambrian, yet again the evidence of direct facilitation of diversity and habitat creation is missing. A second potential ecosystem engineering effect of archaeocyath reefs is to increase nutrient flux to the benthos through increased fluid flow over the reefs, which then creates capacity for an increase in diversity of scavengers and detrivores as well as increasing the capacity for higher trophic-level predators (Manzuk et al. 2023). The reduction in

diversification after the extinction of archaeocyaths suggests that these reefs may have contributed to diversification over the ECT (Na and Kiessling 2015; Figs. 1, 2). The first demonstration of colonization of biotic hard substrates is not until Cambrian Stage 4, when brachiopods form the Guanshan Biota are the first to host epibionts (Chen et al. 2022). While the presence of reefs correlates with increasing diversity (Reaka-Kudla et al. 1996; Small et al. 1998), it is not until the terminal Cambrian that there is a significant correlation between taxonomic diversity origination in reefs (Kiessling et al. 2010). As such, while reefs substantially changed their environment, the evidence that organisms utilized their structures as ecosystem engineers is not in evidence until the terminal Cambrian, so the impact of such reef ecosystem engineering on the ECT diversity patterns is not clear.

In the third type of ecosystem engineering, the substrate was significantly changed through the ECT by increases in bioturbation, also known as the "agronomic revolution" or "Cambrian substrate revolution" (Seilacher 1994; Bottjer et al. 2000). The impact of the first trace makers on the physical environment was initially minimal, with horizontal trails such as Helminthoidichnites and Dickinsonia on or below microbial mats ~555 Ma (Ivantsov and Malakhovskaya 2002; Buatois et al. 2014; Evans et al. 2020; Ivantsov and Zakrevskaya 2021). These horizonal trails increased in density and complexity in the Shibantan and Nama assemblages (Cribb et al. 2019; Linnemann et al. 2019), with increased capacity to bulldoze (e.g., Parapsammichnites), or move sediments (Buatois et al. 2018), and increased depth penetration, cf. treptichnids (Mángano and Buatois 2020). Matground coverage persisted into the Fortunian (Buatois et al. 2014), indicating that sediment disturbance was insufficient to impact mat growth, but crucially, the development of vertical burrows, for example, Treptichnus pedum, and more complex structures, for example, Oldhamia, enabled more efficient feeding on the mats and sediments (Mángano and Buatois 2020). Ichnodiversity and bioturbating organisms increased the most between the terminal Ediacaran and Fortunian within the ECT and Ordovician (Buatois et al. 2020), but the most significant changes in benthic-pelagic coupling occurred during Cambrian Age 2 (529-521 Ma), with the colonization of the substrate by deep suspension-feeding burrowers, cf. Skolithos. Suspension feeders changed the benthic and pelagic environments by increasing organic carbon fluxes and geochemical recycling to the water column, expanding the surface area of the water-substrate interface through their burrows, thus ventilating the substrate, and bringing sediment up from deep to the water interface (Mángano and Buatois 2020), and through the oxygenation of the sediment increasing niche space (Mángano and Buatois 2014), although the realized impacts of these during the ECT may be minimal (Cribb et al. 2023). Detritovores and deposit feeders further churned up the surface substrate, further enhancing geochemical fluxes (McIlroy and Logan 1999; Mazurek 2014; Herbers et al. 2016; Stachacz 2016; Gougeon et al. 2018).

Finally, animals changed the water column by mixing the stratified oceans of the Ediacaran through movement of the water by filter feeding (Lenton et al. 2014; Chen et al. 2022) and swimming (Butterfield 2009, 2018). Biogenically induced aquatic bioturbation would have started with stromatolites and macroalgae changing fluid dynamics through their physical presence as they grew. As yet there is no evidence that these changes in fluid flow led to a feedback inducing further changes. Thus, the first evidence that animals started to contribute to disturbing the fluid flow, leading to changes in the occupation of vertical niches within the water column, comes from the Ediacaran ~572 Ma (Ghisalberti et al. 2014). Mobility in the form of horizontal trace makers and mat stickers is unlikely to have impacted the fluid flow, as the physical changes were limited vertically. Planktonic pelagic macro-animals, such as Attenborites and jellyfish, also likely did not contribute to significant differences of fluid flow, because they only had limited ability to swim against the current. Therefore, the largest impact on ECT water-column fluid dynamics comes from filter feeding, as it increases in abundance both within and top on the substrate, increasing benthic-pelagic coupling (Mángano and Buatois 2014). Today, the largest impact of animals comes from swimming, especially with the marine pump and diel vertical migration, whereby the vertical migration of zooplankton from the depths to the surface at night to feed is the largest (in terms of biomass) migration on this planet, with planktivores following the zooplankton, and predators following them (Barnes 1988; Cisewski et al. 2010). Indeed, the feeding migrations of large marine animals such as whales contribute a substantial amount to oceanic mixing and the biological pump removing carbon from the atmosphere and bringing it into the deep sea (Berger 2007; Cotte et al. 2011; Lavery et al. 2012). However, with the advent of active swimming by macroscopic zooplankton in Cambrian Age 3 (Vannier 2007), nutrients and geochemical mixing would have massively increased vertically, as well as horizontally with larger organisms and larger dispersal ranges, thus increasing the habitability of the water column and opening up new pelagic niches (Butterfield 2011, 2018).

While the four types of different ecosystem engineering processes are not independent, taken together, these animals fundamentally changed the biosphere over the ECT (cf. Butterfield 2011). Importantly, the strong positive feedbacks of these ecosystem engineering processes do not appear to have been fully realized immediately following the respective innovations. Indeed, it may not have been until the Ordovician that full ecosystem engineering feedbacks started, albeit with small effects (Erwin and Tweedt 2012).

## Conclusions

Over the last 50 yr, we have seen a significant change in our understanding of the early evolution of animals with the taxonomic resolution of pre-Cambrian animals in the Ediacaran (Chen et al. 2013; Bobrovskiy et al. 2018; Evans et al. 2020; Dunn et al. 2021, 2022) and the use of phylogenetic analyses to enable to recognition of Ediacaran and Cambrian taxa as stem-group representatives of modern classes and phyla (Daley et al. 2009; Legg et al. 2013; Smith and Caron 2015; Park et al. 2024; Rahman and Zamora 2024). But key for the increase in our capacity to understand the drivers underlying the ECT is our changed approach of putting individual specimens and taxa in the context of the environments in which they lived. After all, it is the interactions that organisms experienced in their daily lives with one another and their environments, that is, their community ecology, that led to the morphological diversification and macroecology and evolutionary patterns that we see over the ECT.

Throughout the ECT, we not only see a diversification in the types of ecological interactions that exist, namely trophic interactions, habitat modifications, and biotic interactions, but also an increase in the number and type of these different interactions. Patterns across different ecological scales are remarkably consistent, with organismal, community, and macroecological patterns all showing a pattern of relative stochasticity, starting in the Ediacaran with low levels of biotic interactions such as competition (Mitchell and Kenchington 2018; Mitchell et al. 2019; Dunn et al. 2022), which then changes with an increase in morphological and ecological diversity (Shen et al. 2008; Bush et al. 2011) at the organism level and increases in niche interactions (Gehling and Droser 2018; Mitchell et al. 2020; Boan et al. 2023) throughout the Ediacaran and into the Cambrian (Bush et al. 2011; Muscente et al. 2019; Buatois et al. 2020; Nanglu et al. 2020b). These patterns are also captured at the macroecological scale (Na and Kiessling 2015; Eden et al. 2022; Meek et al. 2023), demonstrating the buildup of these micro- to macroecological interactions.

By Cambrian Epoch 2, the ecosystem structure is similar in many aspects to that of marine systems today. However, while many of these the key ecological processes, such as predation, habitat modification, facilitation, symbiosis, mutualisms, and commensalism are present (Fig. 4), there are time delays between the origins of such processes and the time when they have an observed impact on other organisms—for example, reef production predates the advent of reef/hard substrate habitation by ~40 Myr, and complex trophic structures emerged around 30 Myr after the advent of predation. As such, while the key building blocks of ecosystem structure were in place by the end of the Cambrian, it takes evolutionary timescales for the impact of them to be realized.

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#### **Literature Cited**

- Anderson, A. J. B. 1971. Ordination methods in ecology. *Journal of Ecology* 59:713–726.
- Anderson, M. J., T. O. Crist, J. M. Chase, M. Vellend, B. D. Inouye, A. L. Freestone, N. J. Sanders, *et al.* 2011. Navigating the multiple meanings of  $\beta$  diversity: a roadmap for the practicing ecologist. *Ecology Letters* 14:19–28.
- Anderson, S. C., P. R. Elsen, B. B. Hughes, R. K. Tonietto, M. C. Bletz, D. A. Gill, M. A. Holgerson, et al. 2021. Trends in ecology and conservation over eight decades. Frontiers in Ecology and the Environment 19:274–282.
- Antcliffe, J. B., and M. D. Brasier. 2007. Charnia and sea pens are poles apart. Journal of the Geological Society 164:49–51.
- Antcliffe, J. B., W. Jessop, and A. C. Daley. 2019. Prey fractionation in the Archaeocyatha and its implication for the ecology of the first animal reef systems. *Paleobiology* 45:652–675.
- Aria, C., and J.-B. Caron. 2017a. Burgess Shale fossils illustrate the origin of the mandibulate body plan. *Nature* 545:89–92.
- Aria, C., and J.-B. Caron. 2017b. Mandibulate convergence in an armoured Cambrian stem chelicerate. *BMC Evolutionary Biology* 17:261.
- Baddeley, A., and R. Turner. 2005. Spatstat: an R package for analyzing spatial point patterns. *Journal of Statistical Software* 12:1–42.
- Bambach, R. K. 1983. Ecospace utilization and guilds in marine communities through the Phanerozoic. Pp. 719–746 in M. J. S. Tevesz and P. L. McCall, eds. Biotic interactions in Recent and fossil benthic communities. Springer US, Boston.
- Bambach, R. K., A. M. Bush, and D. H. Erwin. 2007. Autecology and the filling of ecospace: key metazoan radiations. *Palaeontology* 50:1–22.

- Barnes, H. 1988. Oceanography and marine biology, an annual review. CRC Press, Boca Raton, Fla.
- Bath Enright, O. G., N. J. Minter, E. J. Sumner, M. G. Mángano, and L. A. Buatois. 2021. Flume experiments reveal flows in the Burgess Shale can sample and transport organisms across substantial distances. *Communications Earth and Environment* 2:104.
- Becker-Kerber, B., M. L. A. F. Pacheco, I. D. Rudnitzki, D. Galante, F. Rodrigues, and J. de M. Leme. 2017. Ecological interactions in *Cloudina* from the Ediacaran of Brazil: implications for the rise of animal biomineralization. *Scientific Reports* 7:5482.
- Becker-Kerber, B., P. S. G. Paim, F. Chemale Junior, T. J. Girelli, A. L. Z. da Rosa, A. E. Albani, G. L. Osés, et al. 2020. The oldest record of Ediacaran macrofossils in Gondwana (~563 Ma, Itajaí Basin, Brazil). Gondwana Research 84:211–228.
- Bengtson, S. 1968. The Problematic genus *Mobergella* from the Lower Cambrian of the Baltic area. *Lethaia* 1:325–351.
- Bengtson, S. 2002. Origins and early evolution of predation. Paleontological Society Papers 8:289–318.
- Benus, A., E. Landing, G. M. Narbonne, and P. Myrow. 1988. Sedimentological context of a deep-water Ediacaran fauna (Mistaken Point, Avalon Zone, eastern Newfoundland). Trace fossils, small shelly fossils and the Precambrian–Cambrian boundary. New York State Museum and Geological Survey Bulletin 463:8–9.
- Berger, W. H. 2007. Cenozoic cooling, Antarctic nutrient pump, and the evolution of whales. Deep-Sea Research, part II (Topical Studies in Oceanography) 54:2399–2421.
- Bicknell, R. D. C., and J. R. Paterson. 2018. Reappraising the early evidence of durophagy and drilling predation in the fossil record: implications for escalation and the Cambrian Explosion. *Biological Reviews* 93:754–784.
- Bicknell, R. D. C., J. A. Ledogar, S. Wroe, B. C. Gutzler, W. H. Watson, and J. R. Paterson. 2018. Computational biomechanical analyses demonstrate similar shell-crushing abilities in modern and ancient arthropods. *Proceedings of the Royal Society B* 285:20181935.
- Bicknell, R. D. C., J. D. Holmes, G. D. Edgecombe, S. R. Losso, J. Ortega-Hernández, S. Wroe, and J. R. Paterson. 2021. Biomechanical analyses of Cambrian euarthropod limbs reveal their effectiveness in mastication and durophagy. *Proceedings of the Royal Society B* 288:20202075.
- Bicknell, R. D. C., J. D. Holmes, S. Pates, D. C. García-Bellido, and J. R. Paterson. 2022. Cambrian carnage: trilobite predator-prey interactions in the Emu Bay Shale of South Australia. *Palaeogeography*, *Palaeoclimatology*, *Palaeoecology* 591:110877.
- Billings, E., and E. Billings. 1872. On some fossils from the primordial rocks of Newfoundland. *Canadian Naturalist* 6:465–479.
- Boag, T. H., S. A. F. Darroch, and M. Laflamme. 2016. Ediacaran distributions in space and time: testing assemblage concepts of earliest macroscopic body fossils. *Paleobiology* 42:574–594.
- Boan, P. C., S. D. Evans, C. M. S. Hall, and M. L. Droser. 2023. Spatial distributions of *Tribrachidium*, *Rugoconites*, and *Obamus* from the Ediacara Member (Rawnsley Quartzite), South Australia. *Paleobiology* 49:601–620.
- Bobrovskiy, I., J. M. Hope, A. Ivantsov, B. J. Nettersheim, C. Hallmann, and J. J. Brocks. 2018. Ancient steroids establish the Ediacaran fossil *Dickinsonia* as one of the earliest animals. *Science* **361**:1246–1249.
- Bobrovskiy, I., A. Nagovitsyn, J. M. Hope, E. Luzhnaya, and J. J. Brocks. 2022. Guts, gut contents, and feeding strategies of Ediacaran animals. *Current Biology* **32**:5382–5389.e3.
- Boddy, C., E. G. Mitchell, M. P. Meredith, and A. G. Liu. 2021. The palaeolatitudinal distribution of the Ediacaran macrobiota. *Journal of the Geological Society* 179:jgs2021–030.
- Bottjer, D. J., J. W. Hagadorn, and S. Q. Dornbos. 2000. The Cambrian substrate revolution. GSA Today 10:1–7.
- Bowyer, F., R. A. Wood, and S. W. Poulton. 2017. Controls on the evolution of Ediacaran metazoan ecosystems: a redox perspective. *Geobiology* 15:516–551.
- Bozec, Y.-M., L. Yakob, S. Bejarano, and P. J. Mumby. 2013. Reciprocal facilitation and non-linearity maintain habitat engineering on coral reefs. *Oikos* 122:428–440.
- Brandl, S. J., D. B. Rasher, I. M. Côté, J. M. Casey, E. S. Darling, J. S. Lefcheck, and J. E. Duffy. 2019. Coral reef ecosystem functioning:

eight core processes and the role of biodiversity. *Frontiers in Ecology and the Environment* **17**:445–454.

- Brasier, M. 1979. The Cambrian radiation event. Pp. 253–268 in M. R. House, ed. The origin of major invertebrate groups. Academic Press, London.
- Brasier, M. D., J. B. Antcliffe, and A. G. Liu. 2012. The architecture of Ediacaran Fronds. *Palaeontology* 55:1105–1124.
- Briggs, D. 1976. The arthropod *Branchiocaris* n. gen., Middle Cambrian, Burgess Shale, British Columbia. *Geological Survey of Canada Bulletin* 264:29.
- Briggs, D. E. G. 1979. Anomalocaris, the largest known Cambrian arthropod. Palaeontology 22:631–664.
- Briggs, D. E. G., and R. A. Fortey. 1989. The early radiation and relationships of the major arthropod groups. *Science* 246:241–243.
- Bruton, D. L., and H. B. Whittington. 1983. Emeraldella and Leanchoilia, two arthropods from the Burgess Shale, Middle Cambrian, British Columbia. Philosophical Transactions of the Royal Society B 300:553–582.
- Buatois, L. A., G. M. Narbonne, M. G. Mángano, N. B. Carmona, and P. Myrow. 2014. Ediacaran matground ecology persisted into the earliest Cambrian. *Nature Communications* 5:3544.
- Buatois, L. A., J. Almond, M. G. Mángano, S. Jensen, and G. J. B. Germs. 2018. Sediment disturbance by Ediacaran bulldozers and the roots of the Cambrian explosion. *Scientific Reports* 8:4514.
- Buatois, L. A., M. G. Mángano, N. J. Minter, K. Zhou, M. Wisshak, M. A. Wilson, and R. A. Olea. 2020. Quantifying ecospace utilization and ecosystem engineering during the early Phanerozoic—the role of bioturbation and bioerosion. *Science Advances* 6:eabb0618.
- Budd, G. 1993. A Cambrian gilled lobopod from Greenland. Nature 364:709–711.
- Budd, G. E. 1998. Stem group arthropods from the Lower Cambrian Sirius Passet fauna of North Greenland. Pp. 125–138 in R. A. Fortey and R. H. Thomas, eds. Arthropod relationships. Springer Netherlands, Dordrecht.
- Budd, G. E., and S. Jensen. 2017. The origin of the animals and a "Savannah" hypothesis for early bilaterian evolution. *Biological Reviews* 92:446–473.
- Budd, G. E., and R. P. Mann. 2020. The dynamics of stem and crown groups. Science Advances 6:eaaz1626.
- Budd, G. E., and R. P. Mann. 2023. Two notorious nodes: a critical examination of relaxed molecular clock age estimates of the bilaterian animals and placental mammals. *Systematic Biology* 73:223–234.
- Bush, A. M., R. K. Bambach, and D. H. Erwin. 2011. Ecospace utilization during the Ediacaran radiation and the Cambrian eco-explosion. Pp. 111–133 in M. Laflamme, J. D. Schiffbauer, and S. Q. Dornbos, eds. *Quantifying the evolution of early life: numerical approaches to the evaluation of fossils and ancient ecosystems*. Springer Netherlands, Dordrecht.
- Butterfield, N. J. 2002. Leanchoilia guts and the interpretation of threedimensional structures in Burgess Shale-type fossils. Paleobiology 28:155–171.
- **Butterfield**, N. J. 2009. Oxygen, animals and oceanic ventilation: an alternative view. *Geobiology* 7:1–7.
- Butterfield, N. J. 2011. Animals and the invention of the Phanerozoic Earth system. Trends in Ecology and Evolution 26:81–87.
- Butterfield, N. J. 2018. Oxygen, animals and aquatic bioturbation: an updated account. *Geobiology* 16:3–16.
- Butterfield, N. J. 2022. Constructional and functional anatomy of Ediacaran rangeomorphs. *Geological Magazine* 159:1148–1159.
- Butterfield, N. J., and T. H. P. Harvey. 2012. Small carbonaceous fossils (SCFs): a new measure of early Paleozoic paleobiology. *Geology* 40:71–74.
- Cai, Y., H. Hua, J. D. Schiffbauer, B. Sun, and X. Yuan. 2014. Tube growth patterns and microbial mat-related lifestyles in the Ediacaran fossil *Cloudina*, Gaojiashan Lagerstätte, South China. *Gondwana Research* 25:1008–1018.
- Caron, J.-B., and C. Aria. 2017. Cambrian suspension-feeding lobopodians and the early radiation of panarthropods. BMC Evolutionary Biology 17:29.
- Caron, J.-B., and D. A. Jackson. 2008. Paleoecology of the Greater Phyllopod Bed community, Burgess Shale. Palaeogeography, Palaeoclimatology, Palaeoecology 258:222–256.
- Caron, J.-B., and J. Vannier. 2016. Waptia and the diversification of brood care in early arthropods. *Current Biology* 26:69–74.
- Caron, J.-B., R. R. Gaines, M. G. Mángano, M. Streng, and A. C. Daley. 2010. A new Burgess Shale-type assemblage from the "thin" Stephen Formation of the southern Canadian Rockies. *Geology* 38:811–814.

- Chen, J.-Y., J. Dzik, G. D. Edgecombe, L. Ramsköld, and G.-Q. Zhou. 1995. A possible Early Cambrian chordate. *Nature* **377**:720–722.
- Chen, Z., C. Zhou, M. Meyer, K. Xiang, J. D. Schiffbauer, X. Yuan, and S. Xiao. 2013. Trace fossil evidence for Ediacaran bilaterian animals with complex behaviors. *Precambrian Research* 224:690–701.
- Chen, Z., C. Zhou, X. Yuan, and S. Xiao. 2019. Death march of a segmented and trilobate bilaterian elucidates early animal evolution. *Nature* 573:412–415.
- Chen, F., T. P. Topper, C. B. Skovsted, L. C. Strotz, J. Shen, and Z. Zhang. 2022. Cambrian ecological complexities: perspectives from the earliest brachiopod-supported benthic communities in the early Cambrian Guanshan Lagerstätte. *Gondwana Research* 107:30–41.
- Cisewski, B., V. H. Strass, M. Rhein, and S. Krägefsky. 2010. Seasonal variation of diel vertical migration of zooplankton from ADCP backscatter time series data in the Lazarev Sea, Antarctica. *Deep-Sea Research, part I* (*Oceanographic Research Papers*) 57:78–94.
- Clapham, M. E., G. M. Narbonne, and J. G. Gehling. 2003. Paleoecology of the oldest known animal communities: Ediacaran assemblages at Mistaken Point, Newfoundland. *Paleobiology* 29:527–544.
- Clites, E. C., M. L. Droser, and J. G. Gehling. 2012. The advent of hard-part structural support among the Ediacara biota: Ediacaran harbinger of a Cambrian mode of body construction. *Geology* 40:307–310.
- Collins, D., D. E. G. Briggs, and S. Conway Morris. 1983. New Burgess Shale fossil sites reveal Middle Cambrian faunal complex. *Science* 220:163–167.
- Condon, D., M. Zhu, S. Bowring, W. Wang, A. Yang, and Y. Jin. 2005. U-Pb ages from the Neoproterozoic Doushantuo Formation, China. *Science* 308:95–98.
- Cong, P., X. Ma, X. Hou, G. D. Edgecombe, and N. J. Strausfeld. 2014. Brain structure resolves the segmental affinity of anomalocaridid appendages. *Nature* 513:538–542.
- Cong, P., A. C. Daley, G. D. Edgecombe, and X. Hou. 2017. The functional head of the Cambrian radiodontan (stem-group Euarthropoda) *Amplectobelua symbrachiata. BMC Evolutionary Biology* 17.
- Cong, P., G. D. Edgecombe, A. C. Daley, J. Guo, S. Pates, and X. Hou. 2018. New radiodonts with gnathobase-like structures from the Cambrian Chengjiang biota and implications for the systematics of Radiodonta. *Papers in Palaeontology* 4:605–621.
- **Conway Morris, S.** 1976. *Nectocaris pteryx*, a new organism from the Middle Cambrian Burgess Shale of British Columbia. *Neues Jahrbuch für Geologie und Paläontologie, Monatshefte* **12**:703–713.
- **Conway Morris, S.** 1977. A redescription of the Middle Cambrian worm *Amiskwia sagittiformis* Walcott from the Burgess Shale of British Columbia. *Paläontologische Zeitschrift* **51**:271–287.
- Conway Morris, S., and S. Bengtson. 1994. Cambrian predators: possible evidence from boreholes. *Journal of Paleontology* 68:1–23.
- Conway Morris, S., and J.-B. Caron. 2012. *Pikaia gracilens* Walcott, a stem-group chordate from the Middle Cambrian of British Columbia. *Biological Reviews* 87:480–512.
- Conway Morris, S., and R. A. Robison. 1982. The enigmatic medusoid *Peytoia* and a comparison of some Cambrian biotas. *Journal of Paleontology* 56:116–122.
- Cortijo, I., M. Martí Mus, S. Jensen, and T. Palacios. 2010. A new species of *Cloudina* from the terminal Ediacaran of Spain. *Precambrian Research* 176:1–10.
- Cotte, C., F. d'Ovidio, A. Chaigneau, M. Lèvy, I. Taupier-Letage, B. Mate, and C. Guinet. 2011. Scale-dependent interactions of Mediterranean whales with marine dynamics. *Limnology and Oceanography* 56:219–232.
- Coutts, F. J., C. J. A. Bradshaw, D. C. García-Bellido, and J. G. Gehling. 2018. Evidence of sensory-driven behavior in the Ediacaran organism *Parvancorina*: implications and autecological interpretations. *Gondwana Research* 55:21–29.
- Cribb, A. T., C. G. Kenchington, B. Koester, B. M. Gibson, T. H. Boag, R. A. Racicot, H. Mocke, M. Laflamme, and S. A. F. Darroch. 2019. Increase in metazoan ecosystem engineering prior to the Ediacaran–Cambrian boundary in the Nama Group, Namibia. *Royal Society Open Science* 6:190548.
- Cribb, A. T., S. J. van de Velde, W. M. Berelson, D. J. Bottjer, and F. A. Corsetti. 2023. Ediacaran–Cambrian bioturbation did not extensively oxygenate sediments in shallow marine ecosystems. *Geobiology* 21:435–453.

- Daley, A. C., and G. E. Budd. 2010. New anomalocaridid appendages from the Burgess Shale, Canada. *Palaeontology* 53:721–738.
- Daley, A. C., and G. D. Edgecombe. 2014. Morphology of Anomalocaris canadensis from the Burgess Shale. Journal of Paleontology 88:68–91.
- Daley, A. C., G. E. Budd, J.-B. Caron, G. D. Edgecombe, and D. Collins. 2009. The Burgess Shale anomalocaridid *Hurdia* and its significance for early euarthropod evolution. *Science* **323**:1597–1600.
- Daley, A. C., G. E. Budd, and J.-B. Caron. 2013. Morphology and systematics of the anomalocaridid arthropod *Hurdia* from the Middle Cambrian of British Columbia and Utah. *Journal of Systematic Palaeontology* 11:743–787.
- Daley, A. C., J. B. Antcliffe, H. B. Drage, and S. Pates. 2018. Early fossil record of Euarthropoda and the Cambrian Explosion. *Proceedings of the National Academy of Sciences USA* 115:5323–5331.
- Darroch, S., M. Laflamme, J. D. Schiffbauer, and D. E. G. Briggs. 2012. Experimental formation of a microbial death mask. *Palaios* 27:293–303.
- Darroch, S. A. F., M. Laflamme, and M. E. Clapham. 2013. Population structure of the oldest known macroscopic communities from Mistaken Point, Newfoundland. *Paleobiology* 39:591–608.
- Darroch, S. A. F., I. A. Rahman, B. Gibson, R. A. Racicot, and M. Laflamme. 2017. Inference of facultative mobility in the enigmatic Ediacaran organism *Parvancorina*. *Biology Letters* 13:20170033.
- Darroch, S. A. F., M. Laflamme, and P. J. Wagner. 2018a. High ecological complexity in benthic Ediacaran communities. *Nature Ecology and Evolution* 2:1541–1547.
- Darroch, S. A. F., E. F. Smith, M. Laflamme, and D. H. Erwin. 2018b. Ediacaran Extinction and Cambrian Explosion. *Trends in Ecology and Evolution* 33:653–663.
- Darroch, S. A. F., S. Gutarra, H. Masaki, A. Olaru, B. M. Gibson, F. S. Dunn, E. G. Mitchell, R. A. Racicot, G. Burzynski, and I. A. Rahman. 2023a. The rangeomorph *Pectinifrons abyssalis*: hydrodynamic function at the dawn of animal life. *iScience* 26:105989.
- Darroch, S. A. F., E. F. Smith, L. L. Nelson, M. Craffey, J. D. Schiffbauer, and M. Laflamme. 2023b. Causes and consequences of end-Ediacaran extinction: an update. *Cambridge Prisms: Extinction* 1:e15.
- Darroch, S. A. F., E. A. Sperling, T. H. Boag, R. A. Racicot, S. J. Mason, A. S. Morgan, S. Tweedt, et al. 2015. Biotic replacement and mass extinction of the Ediacara biota. Proceedings of the Royal Society B 282:20151003.
- De Baets, K., J. W. Huntley, D. Scarponi, A. A. Klompmaker, and A. Skawina. 2021. Phanerozoic parasitism and marine metazoan diversity: dilution versus amplification. *Philosophical Transactions of the Royal Society B* 376:20200366.
- Droser, M. L., and J. G. Gehling. 2008. Synchronous aggregate growth in an abundant new Ediacaran tubular organism. *Science* 319:1660–1662.
- Droser, M. L., and J. G. Gehling. 2015. The advent of animals: the view from the Ediacaran. Proceedings of the National Academy of Sciences USA 112:4865–4870.
- **Droser, M. L., L. G. Tarhan, and J. G. Gehling**. 2017. The rise of animals in a changing environment: global ecological innovation in the late Ediacaran. *Annual Review of Earth and Planetary Sciences* **45**:593–617.
- Droser, M. L., S. D. Evans, P. W. Dzaugis, E. B. Hughes, and J. G. Gehling. 2020. Attenborites janeae: a new enigmatic organism from the Ediacara Member (Rawnsley Quartzite), South Australia. Australian Journal of Earth Sciences 67:915–921.
- Droser, M. L., S. D. Evans, L. G. Tarhan, R. L. Surprenant, I. V. Hughes, E. B. Hughes, and J. G. Gehling. 2022. What happens between depositional events, stays between depositional events: the significance of organic mat surfaces in the capture of Ediacara communities and the sedimentary rocks that preserve them. *Frontiers in Earth Science* 10:826353. https://doi.org/10.3389/feart.2022.826353.
- Droser, M. L., J. G. Gehling, L. G. Tarhan, S. D. Evans, C. M. S. Hall, I. V. Hughes, E. B. Hughes, et al. 2019. Piecing together the puzzle of the Ediacara Biota: excavation and reconstruction at the Ediacara National Heritage site Nilpena (South Australia). Palaeogeography, Palaeoclimatology, Palaeoecology 513:132–145.
- Dunn, F. S., A. G. Liu, D. V. Grazhdankin, P. Vixseboxse, J. Flannery-Sutherland, E. Green, S. Harris, P. R. Wilby, and P. C. J. Donoghue. 2021. The developmental biology of *Charnia* and the

eumetazoan affinity of the Ediacaran rangeomorphs. Science Advances 7: eabe0291.

- Dunn, F. S., C. G. Kenchington, L. A. Parry, J. W. Clark, R. S. Kendall, and P. R. Wilby. 2022. A crown-group cnidarian from the Ediacaran of Charnwood Forest, UK. *Nature Ecology and Evolution* 6:1095–1104.
- Dunne, J. A., R. J. Williams, N. D. Martinez, R. A. Wood, and D. H. Erwin. 2008. Compilation and network analyses of Cambrian food webs. *PLoS Biology* 6:e102.
- Eden, R., A. Manica, and E. G. Mitchell. 2022. Metacommunity analyses show an increase in ecological specialisation throughout the Ediacaran period. *PLoS Biology* 20:e3001289.
- Eibye-Jacobsen, D. 2004. A reevaluation of *Wiwaxia* and the polychaetes of the Burgess Shale. *Lethaia* 37:317–335.
- Erwin, D. H., and S. Tweedt. 2012. Ecological drivers of the Ediacaran–Cambrian diversification of Metazoa. *Evolutionary Ecology* 26:417–433.
- Erwin, D. H., M. Laflamme, S. M. Tweedt, E. A. Sperling, D. Pisani, and K. J. Peterson. 2011. The Cambrian conundrum: early divergence and later ecological success in the early history of animals. *Science* 334:1091–1097.
- Evans, S. D., C. W. Diamond, M. L. Droser, and T. W. Lyons. 2018. Dynamic oxygen and coupled biological and ecological innovation during the second wave of the Ediacara Biota. *Emerging Topics in Life Sciences* 2:223–233.
- Evans, S. D., J. G. Gehling, and M. L. Droser. 2019. Slime travelers: early evidence of animal mobility and feeding in an organic mat world. *Geobiology* 17:490–509.
- Evans, S. D., I. V. Hughes, J. G. Gehling, and M. L. Droser. 2020. Discovery of the oldest bilaterian from the Ediacaran of South Australia. *Proceedings of* the National Academy of Sciences USA 117:7845–7850.
- Evans, S. D., C. Tu, A. Rizzo, R. L. Surprenant, P. C. Boan, H. McCandless, N. Marshall, S. Xiao, and M. L. Droser. 2022. Environmental drivers of the first major animal extinction across the Ediacaran White Sea–Nama transition. *Proceedings of the National Academy of Sciences USA* 119: e2207475119.
- Fedonkin, M. A., and B. M. Waggoner. 1997. The Late Precambrian fossil Kimberella is a mollusc-like bilaterian organism. Nature 388:868–871.
- Fedonkin, M. A., A. Simonetta, and A. Y. Ivantsov. 2007. New data on *Kimberella*, the Vendian mollusc-like organism (White Sea region, Russia): palaeoecological and evolutionary implications. *Geological Society* of London Special Publication 286:157–179.
- Finnegan, S., J. G. Gehling, and M. L. Droser. 2019. Unusually variable paleocommunity composition in the oldest metazoan fossil assemblages. *Paleobiology* 45:235–245.
- Ford, T. D. 1958. Pre-Cambrian fossils from Charnwood Forest. Proceedings of the Yorkshire Geological Society 31:211–217.
- Fu, D., J. Ortega-Hernández, A. C. Daley, X. Zhang, and D. Shu. 2018. Anamorphic development and extended parental care in a 520 million-year-old stem-group euarthropod from China. *BMC Evolutionary Biology* 18:147.
- Fu. D., G. Tong, T. Dai, W. Liu, Y. Yang, Y. Zhang, L. Cui, L. Li, H. Yun, Y. Wu, A. Sun, C. Liu, W. Pei, R. R. Gaines, and X. Zhang. 2019. The Qingjiang biota—a Burgess Shale-type fossil Lagerstätte from the early Cambrian of South China. *Science* 363:1338–1342.
- Gaines, R. R. 2014. Burgess Shale-type preservation and its distribution in space and time. *The Paleontological Society Papers* 20:1.
- Gandin, A., and F. Debrenne. 2010. Distribution of the archaeocyath–calcimicrobial bioconstructions on the Early Cambrian shelves. *Palaeoworld* 19:222–241.
- Gehling, J. G. 1999. Microbial mats in terminal Proterozoic siliciclastics; Ediacaran death masks. *Palaios* 14:40–57.
- Gehling, J. G., and M. L. Droser. 2018. Ediacaran scavenging as a prelude to predation. *Emerging Topics in Life Sciences* 2:213–222.
- Gehling, J. G., M. Droser, and S. R. Jensen. 2005. Ediacaran organisms: relating form and function. Pp. 43–67 in D. E. G. Briggs, ed. Evolving form and function: fossils and development: proceedings of a symposium honoring Adolf Seilacher for his contributions to paleontology, in celebration of his 80th birthday. Yale University Press, New Haven, Conn.
- Ghisalberti, M., D. A. Gold, M. Laflamme, M. E. Clapham, G. M. Narbonne, R. E. Summons, D. T. Johnston, and D. K. Jacobs. 2014.

Canopy flow analysis reveals the advantage of size in the oldest communities of multicellular eukaryotes. *Current Biology* **24**:305–309.

- Gibson, B. M., J. D. Schiffbauer, and S. A. F. Darroch. 2018. Ediacaran-style decay experiments using mollusks and sea anemones. *Palaios* 33:185–203.
- Gibson, B. M., D. J. Furbish, I. A. Rahman, M. W. Schmeeckle, M. Laflamme, and S. A. F. Darroch. 2020. Ancient life and moving fluids. *Biological Reviews* 96:129–152.
- Gibson, B. M., S. A. F. Darroch, K. M. Maloney, and M. Laflamme. 2021. The importance of size and location within gregarious populations of *Ernietta plateauensis. Frontiers in Earth Science* 9:749150. https://doi.org/ 10.3389/feart.2021.749150.
- Gibson, B. M., M. Chipman, P. Attanasio, Z. Qureshi, S. A. F. Darroch, I. A. Rahman, and M. Laflamme. 2023. Reconstructing the feeding ecology of Cambrian sponge reefs: the case for active suspension feeding in Archaeocyatha. *Royal Society Open Science* 10:230766.
- Giguère, T. N., and V. Tunnicliffe. 2021. Beta diversity differs among hydrothermal vent systems: implications for conservation. *PLoS ONE* 16: e0256637.
- Glaessner, M. F. 1959. The oldest fossil faunas of South Australia. Geologische Rundschau 47:522–531.
- Gougeon, R. C., M. G. Mángano, L. A. Buatois, G. M. Narbonne, and B. A. Laing. 2018. Early Cambrian origin of the shelf sediment mixed layer. *Nature Communications* 9:1909.
- Grazhdankin, D. 2004. Patterns of distribution in the Ediacaran biotas: facies versus biogeography and evolution. *Paleobiology* **30**:203–221.
- Grazhdankin, D., and A. Seilacher. 2002. Underground Vendobionta from Namibia. Palaeontology 45:57–78.
- Griffiths, H. J., R. J. Whittle, and E. G. Mitchell. 2023. Animal survival strategies in Neoproterozoic ice worlds. *Global Change Biology* 29:10–20.
- Grotzinger, J. P., E. W. Adams, and S. Schroder. 2005. Microbial-metazoan reefs of the terminal Proterozoic Nama Group (c. 550–543 Ma), Namibia. *Geological Magazine* 142:499–517.
- Gürich, G. 1933. Die Kuibis-Fossilien der Nama-Formation von Südwestafrika. Palaeontologische Zeitschrift 15:137–154.
- Hall, M., A. J. Kaufman, P. Vickers-Rich, A. Ivantsov, P. Trusler, U. Linnemann, M. Hofmann, et al. 2013. Stratigraphy, palaeontology and geochemistry of the late Neoproterozoic Aar Member, southwest Namibia: reflecting environmental controls on Ediacara fossil preservation during the terminal Proterozoic in African Gondwana. Precambrian Research 238:214–232.
- Harborne, A. R., P. J. Mumby, K. ZŻychaluk, J. D. Hedley, and P. G. Blackwell. 2006. Modeling the beta diversity of coral reefs. *Ecology* 87:2871–2881.
- Harper, D. A. T., E. U. Hammarlund, T. P. Topper, A. T. Nielsen, J. A. Rasmussen, T.-Y. S. Park, and M. P. Smith. 2019. The Sirius Passet Lagerstätte of North Greenland: a remote window on the Cambrian Explosion. *Journal of the Geological Society* 176:1023–1037.
- Harvey, T. H. P., and N. J. Butterfield. 2008. Sophisticated particle-feeding in a large Early Cambrian crustacean. *Nature* 452:868–871.
- Harvey, T. H. P., and N. J. Butterfield. 2022. A new species of early Cambrian arthropod reconstructed from exceptionally preserved mandibles and associated small carbonaceous fossils (SCFs). *Papers in Palaeontology* 8:e1458.
- Hastings, A., J. E. Byers, J. A. Crooks, K. Cuddington, C. G. Jones, J. G. Lambrinos, T. S. Talley, and W. G. Wilson. 2007. Ecosystem engineering in space and time. *Ecology Letters* 10:153–164.
- Haug, J. T., D. Walosek, A. Maas, Y. Liu, and C. Haug. 2012. Functional morphology, ontogeny and evolution of mantis shrimp-like predators in the Cambrian. *Palaeontology* 55:369–399.
- Hautmann, M. 2014. Diversification and diversity partitioning. *Paleobiology* 40:162–176.
- Hendricks, J. R., B. S. Lieberman, and A. L. Stigall. 2008. Using GIS to study palaeobiogeographic and macroevolutionary patterns in soft-bodied Cambrian arthropods. *Palaeogeography, Palaeoclimatology, Palaeoecology* 264:163–175.
- Herbers, D. S., R. B. MacNaughton, E. R. Timmer, and M. K. Gingras. 2016. Sedimentology and ichnology of an Early–Middle Cambrian storm-influenced barred shoreface succession, Colville Hills, Northwest Territories. *Bulletin of Canadian Petroleum Geology* 64:538–554.

- Hillebrand, H., D. M. Bennett, and M. W. Cadotte. 2008. Consequences of dominance: a review of evenness effects on local and regional ecosystem processes. *Ecology* 89:1510–1520.
- Hofmann, H. J., and E. W. Mountjoy. 2001. Namacalathus–Cloudina assemblage in Neoproterozoic Miette Group (Byng Formation), British Columbia: Canada's oldest shelly fossils. *Geology* 29:1091–1094.
- Hou, X.-G., D. J. Siveter, D. J. Siveter, R. J. Aldridge, P.-Y. Cong, S. E. Gabbot, X. Ma, M. A. Purnell, and M. Williams. 2017. The Cambrian fossils of Chengjiang, China: the flowering of early animal life, 2nd ed. Wiley, Oxford.
- Howard, R. J., X. Hou, G. D. Edgecombe, T. Salge, X. Shi, and X. Ma. 2020. A tube-dwelling Early Cambrian lobopodian. *Current Biology* 30:1529–1536.e2.
- Hoyal Cuthill, J. F., and S. Conway Morris. 2014. Fractal branching organizations of Ediacaran rangeomorph fronds reveal a lost Proterozoic body plan. Proceedings of the National Academy of Sciences USA 111:13122–13126.
- Hu, S., M. Steiner, M. Zhu, B.-D. Erdtmann, H. Luo, L. Chen, and B. Weber. 2007. Diverse pelagic predators from the Chengjiang Lagerstätte and the establishment of modern-style pelagic ecosystems in the early Cambrian. *Palaeogeography, Palaeoclimatology, Palaeoecology* 254:307–316.
- Hua, H., B. R. Pratt, and L.-Y. Zhang. 2003. Borings in *Cloudina* Shells: complex predator-prey dynamics in the terminal Neoproterozoic. *Palaios* 18:454–459.
- Huang, T., D. Chen, Y. Ding, X. Zhou, and G. Zhang. 2020. SIMS U-Pb zircon geochronological and carbon isotope chemostratigraphic constraints on the Ediacaran-Cambrian boundary succession in the Three Gorges area, South China. *Journal of Earth Science* 31:69–78.
- Hughes, C. P. 1975. Redescription of *Burgessia bella* from the Middle Cambrian Burgess Shale, British Columbia. Pp. 415–435 in Evolution and morphology of the Trilobita, Trilobitoidea and Merostomata. Universitetsforlaget, Oslo, Norway.
- Illian, D. J., P. A. Penttinen, D. H. Stoyan, and D. D. Stoyan. 2008. Statistical analysis and modelling of spatial point patterns. Wiley, Hoboken, N.J.
- Ivantsov, A. Yu. 2011. Feeding traces of proarticulata—the Vendian metazoa. Paleontological Journal 45:237–248.
- Ivantsov, A. Y., and Y. E. Malakhovskaya. 2002. Giant traces of Vendian animals. Doklady Earth Sciences 385:6.
- Ivantsov, A., and M. Zakrevskaya. 2021. Dickinsonia: mobile and adhered. Geological Magazine 159:1118–1133.
- Ivantsov, A., G. M. Narbonne, P. W. Dzaugis, C. Greentree, and P. Vickers-Rich. 2016. Elucidating *Ernietta*: new insights from exceptional specimens in the Ediacaran of Namibia. *Lethaia* 49:539–554.
- Ivantsov, A., A. Nagovitsyn, and M. Zakrevskaya. 2019. Traces of locomotion of Ediacaran macroorganisms. *Geosciences* 9:395.
- Ives, A. R., and S. R. Carpenter. 2007. Stability and diversity of ecosystems. *Science* 317:58–62.
- Izquierdo-López, A., and J.-B. Caron. 2019. A possible case of inverted lifestyle in a new bivalved arthropod from the Burgess Shale. *Royal Society Open Science* 6:191350.
- Jain, S., and S. Krishna. 2002. Large extinctions in an evolutionary model: the role of innovation and keystone species. *Proceedings of the National Academy of Sciences USA* 99:2055–2060.
- Jensen, S., B. Z. Saylor, J. G. Gehling, and G. J. B. Germs. 2000. Complex trace fossils from the terminal Proterozoic of Namibia. *Geology* 28:143–146.
- Jensen, S., M. L. Droser, and J. G. Gehling. 2006. A critical look at the Ediacaran trace fossil record. Pp. 115–157 in S. Xiao and A. J. Kaufman, eds. Neoproterozoic geobiology and paleobiology. Springer Netherlands, Dordrecht.
- Jiao, D.-G., S. Pates, R. Lerosey-Aubril, J. Ortega-Hernández, J. Yang, T. Lan, and X.-G. Zhang. 2021. New multipodomerous appendages of stem-group euarthropods from the Cambrian (Stage 4) Guanshan Konservat-Lagerstätte. *Royal Society Open Science* 8:211134.
- Jones, C. G., J. H. Lawton, and M. Shachak. 1994. Organisms as ecosystem engineers. Oikos 69:373–386.
- Kiessling, W., C. Simpson, and M. Foote. 2010. Reefs as cradles of evolution and sources of biodiversity in the Phanerozoic. *Science* 327:196–198.

- Kihm, J.-H., F. W. Smith, S. Kim, H. S. Rho, X. Zhang, J. Liu, and T.-Y. S. Park. 2023. Cambrian lobopodians shed light on the origin of the tardigrade body plan. *Proceedings of the National Academy of Sciences USA* 120: e2211251120.
- Kimmig, J., L. C. Strotz, S. R. Kimmig, S. O. Egenhoff, and B. S. Lieberman. 2019. The Spence Shale Lagerstätte: an important window into Cambrian biodiversity. *Journal of the Geological Society* 176:609–619.
- Klompmaker, A. A., and S. Finnegan. 2018. Extreme rarity of competitive exclusion in modern and fossil marine benthic ecosystems. *Geology* 46:723–726.
- Kouchinsky, A., S. Bengtson, B. Runnegar, C. Skovsted, M. Steiner, and M. Vendrasco. 2012. Chronology of early Cambrian biomineralization. *Geological Magazine* 149:221–251.
- Laflamme, M., and G. M. Narbonne. 2008. Ediacaran fronds. Palaeogeography, Palaeoclimatology, Palaeoecology 258:162–179.
- Laflamme, M., S. Xiao, and M. Kowalewski. 2009. Osmotrophy in modular Ediacara organisms. *Proceedings of the National Academy of Sciences USA* 106:14438–14443.
- Laflamme, M., J. D. Schiffbauer, G. M. Narbonne, and D. E. G. Briggs. 2011. Microbial biofilms and the preservation of the Ediacara biota. *Lethaia* 44:203–213.
- Laflamme, M., S. A. F. Darroch, S. M. Tweedt, K. J. Peterson, and D. H. Erwin. 2013. The end of the Ediacara biota: extinction, biotic replacement, or Cheshire Cat? *Gondwana Research* 23:558–573.
- Laundré, J. W., L. Hernández, and K. B. Altendorf. 2001. Wolves, elk, and bison: reestablishing the "landscape of fear" in Yellowstone National Park, U.S.A. *Canadian Journal of Zoology* 79:1401–1409.
- Lavery, T. J., B. Roudnew, L. Seuront, J. G. Mitchell, and J. Middleton. 2012. Can whales mix the ocean? *Biogeosciences Discussions* 9:8387–8403.
- Legg, D. A., M. D. Sutton, and G. D. Edgecombe. 2013. Arthropod fossil data increase congruence of morphological and molecular phylogenies. *Nature Communications* 4:1–7.
- Leibold, M. A., M. Holyoak, N. Mouquet, P. Amarasekare, J. M. Chase, M. F. Hoopes, R. D. Holt, *et al.* 2004. The metacommunity concept: a framework for multi-scale community ecology. *Ecology Letters* 7:601–613.
- Lenton, T. M., R. A. Boyle, S. W. Poulton, G. A. Shields-Zhou, and N. J. Butterfield. 2014. Co-evolution of eukaryotes and ocean oxygenation in the Neoproterozoic era. *Nature Geoscience* 7:257–265.
- Lerosey-Aubril, R., and J. Ortega-Hernández. 2022. A new lobopodian from the middle Cambrian of Utah: did swimming body flaps convergently evolve in stem-group arthropods? *Papers in Palaeontology* 8:e1450.
- Lerosey-Aubril, R., and S. Pates. 2018. New suspension-feeding radiodont suggests evolution of microplanktivory in Cambrian macronekton. *Nature Communications* 9:377..
- Lerosey-Aubril, R., R. R. Gaines, T. A. Hegna, J. Ortega-Hernández, P. Van Roy, C. Kier, and E. Bonino. 2018. The Weeks Formation Konservat-Lagerstätte and the evolutionary transition of Cambrian marine life. *Journal of the Geological Society* **175**:705–715.
- Lerosey-Aubril, R., J. Kimmig, S. Pates, J. Skabelund, A. Weug, and J. Ortega-Hernández. 2020. New exceptionally preserved panarthropods from the Drumian Wheeler Konservat-Lagerstätte of the House Range of Utah. *Papers in Palaeontology* 4:501–531.
- Li, Y., M. Williams, T. H. P. Harvey, F. Wei, Y. Zhao, J. Guo, S. Gabbott, T. Fletcher, X. Hou, and P. Cong. 2020. Symbiotic fouling of Vetulicola, an early Cambrian nektonic animal. *Communications Biology* 3:517.
- Lieberman, B. S. 2002. Phylogenetic analysis of some basal early Cambrian trilobites, the biogeographic origins of the Eutrilobita, and the timing of the Cambrian radiation. *Journal of Paleontology* **76**:692–708.
- Lieberman, B. S. 2003. A new soft-bodied fauna: the Pioche Formation of Nevada. *Journal of Paleontology* 77:674–690.
- Lieberman, B. S. 2008. The Cambrian radiation of bilaterians: evolutionary origins and palaeontological emergence; earth history change and biotic factors. *Palaeogeography, Palaeoclimatology, Palaeoecology* 258:180–188.
- Lieberman, B. S., and J. G. Meert. 2004. Biogeography and the nature and timing of the Cambrian radiation. *Paleontological Society Papers* 10:79–92.
- Linnemann, U., M. Ovtcharova, U. Schaltegger, A. Gärtner, M. Hautmann, G. Geyer, P. Vickers-Rich, et al. 2019. New high-resolution age data from

the Ediacaran–Cambrian boundary indicate rapid, ecologically driven onset of the Cambrian explosion. *Terra Nova* **31**:49–58.

- Liu, A. G., D. Mcllroy, and M. D. Brasier. 2010. First evidence for locomotion in the Ediacara biota from the 565 Ma Mistaken Point Formation, Newfoundland. *Geology* 38:123–126.
- Liu, A. G., J. J. Matthews, L. R. Menon, D. McIlroy, and M. D. Brasier. 2014. Haootia quadriformis n. gen., n. sp., interpreted as a muscular cnidarian impression from the Late Ediacaran period (approx. 560 Ma). Proceedings of the Royal Society B 281:20141202.
- Liu, J., Q. Ou, J. Han, J. Li, Y. Wu, G. Jiao, and T. He. 2015. Lower Cambrian polychaete from China sheds light on early annelid evolution. *Die Naturwissenschaften* 102:34.
- Liu, Y., G. Scholtz, and X. Hou. 2015. When a 520 million-year-old Chengjiang fossil meets a modern micro-CT—a case study. *Scientific Reports* 5:112802.
- Ma, J., S. Pates, Y. Wu, W. Lin, C. Liu, Y. Wu, M. Zhang, and D. Fu. 2023. Ontogeny and brooding strategy of the early Cambrian arthropod *Isoxys* minor from the Qingjiang biota. Frontiers in Ecology and Evolution 11:1–10.
- Ma, X., X. Hou, and J. Bergström. 2009. Morphology of Luolishania longicruris (Lower Cambrian, Chengjiang Lagerstätte, SW China) and the phylogenetic relationships within lobopodians. Arthropod Structure and Development 38:271–291.
- Ma, X., X. Hou, R. J. Aldridge, D. J. Siveter, D. J. Siveter, S. E. Gabbott, M. A. Purnell, A. R. Parker, and G. D. Edgecombe. 2012a. Morphology of Cambrian lobopodian eyes from the Chengjiang Lagerstätte and their evolutionary significance. Arthropod Structure and Development 41:495–504.
- Ma, X., X. Hou, G. D. Edgecombe, and N. J. Strausfeld. 2012b. Complex brain and optic lobes in an early Cambrian arthropod. *Nature* 490:258–261.
- Mángano, M. G., and L. A. Buatois. 2014. Decoupling of body-plan diversification and ecological structuring during the Ediacaran–Cambrian transition: evolutionary and geobiological feedbacks. *Proceedings of the Royal Society B* 281:20140038.
- Mángano, M. G., and L. A. Buatois. 2017. The Cambrian revolutions: tracefossil record, timing, links and geobiological impact. *Earth-Science Reviews* 173:96–108.
- Mángano, M. G., and L. A. Buatois. 2020. The rise and early evolution of animals: where do we stand from a trace-fossil perspective? *Interface Focus* 10:20190103.
- Manzuk, R. A., A. C. Maloof, J. A. Kaandorp, and M. Webster. 2023. Branching archaeocyaths as ecosystem engineers during the Cambrian radiation. *Geobiology* 21:66–85.
- Mariotti, G., S. B. Pruss, X. Ai, J. T. Perron, and T. Bosak. 2016. Microbial Origin of Early Animal Trace Fossils? *Journal of Sedimentary Research* 86:287–293.
- Matthews, J. J., A. G. Liu, C. Yang, D. McIlroy, B. Levell, and D. J. Condon. 2021. A chronostratigraphic framework for the rise of the Ediacaran macrobiota: new constraints from Mistaken Point Ecological Reserve, Newfoundland. GSA Bulletin.
- May, R. M. 1974. Stability and Complexity in Model Ecosystems. Princeton University Press, p. 266
- May, R. M. 2019. Stability and Complexity in Model Ecosystems. Princeton University Press, p.
- Mazurek, D. 2014. The Tommotian phase of the Early Cambrian Agronomic Revolution in the carbonate mud environment of central Siberia. *Lethaia* 47:133–150.
- McCann, K. S. 2000. The diversity-stability debate. Nature 405:228-233.
- McIlroy, D., and G. A. Logan. 1999. The impact of bioturbation on infaunal ecology and evolution during the Proterozoic-Cambrian transition. *PALAIOS* 14:58–72.
- Meek, D. M., L. A. Buatois, M. G. Mángano, and B. M. Eglington. 2023. Increased habitat segregation at the dawn of the Phanerozoic revealed by correspondence analysis of bioturbation. *Scientific Reports* 13:22328.
- Meyer, M., D. Elliott, J. D. Schiffbauer, M. Hall, K. H. Hoffman, G. Schneider, P. Vickers-Rich, and S. Xiao. 2014. Taphonomy of the Ediacaran fossil *Pteridinium simplex* preserved three-dimensionally in mass flow deposits, Nama Group, Namibia. *Journal of Paleontology* 88:240–252.

- Minter, N. J., M. G. Mángano, and J.-B. Caron. 2011. Skimming the surface with Burgess Shale arthropod locomotion. *Proceedings of the Royal Society B* 279:1613–1620.
- Misra, S. B. 1969. Late Precambrian (?) fossils from southeastern Newfoundland. GSA Bulletin 80:2133–2140.
- Mitchell, E. G., and N. J. Butterfield. 2018. Spatial analyses of Ediacaran communities at Mistaken Point. *Paleobiology* 44:40–57.
- Mitchell, E. G., and C. G. Kenchington. 2018. The utility of height for the Ediacaran organisms of Mistaken Point. *Nature Ecology and Evolution* 2:1218–1222.
- Mitchell, E. G., C. G. Kenchington, A. G. Liu, J. J. Matthews, and N. J. Butterfield. 2015. Reconstructing the reproductive mode of an Ediacaran macro-organism. *Nature* 524:343–346.
- Mitchell, E. G., S. Harris, C. G. Kenchington, P. Vixseboxse, L. Roberts, C. Clark, A. Dennis, A. G. Liu, and P. R. Wilby. 2019. The importance of neutral over niche processes in structuring Ediacaran early animal communities. *Ecology Letters* 22:2028–2038.
- Mitchell, E. G., N. Bobkov, N. Bykova, A. Dhungana, A. Kolesnikov, I. R. P. Hogarth, A. G. Liu, *et al.* 2020. The influence of environmental setting on the community ecology of Ediacaran organisms. *Interface Focus* 10.
- Mitchell, E. G., S. D. Evans, Z. Chen, and S. Xiao. 2022. A new approach for investigating spatial relationships of ichnofossils: a case study of Ediacaran–Cambrian animal traces. *Paleobiology* 48:557–575.
- Mittelbach, G. G., D. W. Schemske, H. V. Cornell, A. P. Allen, J. M. Brown, M. B. Bush, S. P. Harrison, *et al.* 2007. Evolution and the latitudinal diversity gradient: speciation, extinction and biogeography. *Ecology Letters* 10:315–331.
- Moody, E. R., S. Álvarez-Carretero, T. A. Mahendrarajah, J. W. Clark, H. C. Betts, N. Dombrowski, L. L. Szánthó, R. A. Boyle, S. Daines, X. Chen, and N. Lane. 2024. The nature of the last universal common ancestor and its impact on the early Earth system. *Nature Ecology and Evolution* 12:1–3.
- Moysiuk, J., and J.-B. Caron. 2019. A new hurdiid radiodont from the Burgess Shale evinces the exploitation of Cambrian infaunal food sources. *Proceedings of the Royal Society B* 286.
- Moysiuk, J., and J.-B. Caron. 2021. Exceptional multifunctionality in the feeding apparatus of a mid-Cambrian radiodont. *Paleobiology* 47:704–724.
- Moysiuk, J., M. R. Smith, and J.-B. Caron. 2017. Hyoliths are Palaeozoic lophophorates. *Nature* 541:394–397.
- Muscente, A. D., J. D. Schiffbauer, J. Broce, M. Laflamme, K. O'Donnell, T. H. Boag, M. Meyer, A., et al. 2017. Exceptionally preserved fossil assemblages through geologic time and space. *Gondwana Research* 48:164–188.
- Muscente, A. D., N. Bykova, T. H. Boag, L. A. Buatois, M. G. Mángano, A. Eleish, A. Prabhu, *et al.* 2019. Ediacaran biozones identified with network analysis provide evidence for pulsed extinctions of early complex life. *Nature Communications* 10:911.
- Na, L., and W. Kiessling. 2015. Diversity partitioning during the Cambrian radiation. Proceedings of the National Academy of Sciences USA 112:4702–4706.
- Na, L., Á. T. Kocsis, Q. Li, and W. Kiessling. 2023. Coupling of geographic range and provincialism in Cambrian marine invertebrates. *Paleobiology* 49:284–295.
- Nanglu, K., and J. B. Caron. 2018. A new Burgess Shale polychaete and the origin of the annelid head revisited. *Current Biology* 28:319–326.
- Nanglu, K., and J.-B. Caron. 2021. Symbiosis in the Cambrian: enteropneust tubes from the Burgess Shale co-inhabited by commensal polychaetes. *Proceedings of the Royal Society B* 288:20210061.
- Nanglu, K., and T. M. Cullen. 2023. Across space and time: a review of sampling, preservational, analytical, and anthropogenic biases in fossil data across macroecological scales. *Earth-Science Reviews* 244:104537.
- Nanglu, K., J.-B. Caron, S. Conway Morris, and C. B. Cameron. 2016. Cambrian suspension-feeding tubicolous hemichordates. *BMC Biology* 14:56.
- Nanglu, K., J.-B. Caron, and C. B. Cameron. 2020a. Cambrian tentaculate worms and the origin of the hemichordate body plan. *Current Biology* 30:4238–4244.e1.

- Nanglu, K., J.-B. Caron, and R. R. Gaines. 2020b. The Burgess Shale paleocommunity with new insights from Marble Canyon, British Columbia. *Paleobiology* 46:58–81.
- Nanglu, K., R. Lerosey-Aubril, J. C. Weaver, and J. Ortega-Hernández. 2023. A mid-Cambrian tunicate and the deep origin of the ascidiacean body plan. *Nature Communications* 14:3832.
- Narbonne, G. M. 2004. Modular construction of early Ediacaran complex life forms. *Science* 305:1141–1144.
- Narbonne, G. M. 2005. The Ediacara biota: Neoproterozoic origin of animals and their ecosystems. Annual Review of Earth and Planetary Sciences 33:421–442.
- Narbonne, G. M., P. M. Myrow, E. Landing, and M. M. Anderson. 1987. A candidate stratotype for the Precambrian–Cambrian boundary, Fortune Head, Burin Peninsula, southeastern Newfoundland. *Canadian Journal of Earth Sciences* 24:1277–1293.
- Narbonne, G. M., B. Z. Saylor, and J. P. Grotzinger. 1997. The youngest Ediacaran fossils from Southern Africa. *Journal of Paleontology* 71:953–967.
- Neutel, A.-M., J. A. P. Heesterbeek, J. van de Koppel, G. Hoenderboom, A. Vos, C. Kaldeway, F. Berendse, and P. C. de Ruiter. 2007. Reconciling complexity with stability in naturally assembling food webs. *Nature* 449:599–602.
- Noble, S. R., D. J. Condon, J. N. Carney, P. R. Wilby, T. C. Pharaoh, and T. D. Ford. 2015. U-Pb geochronology and global context of the Charnian Supergroup, UK: constraints on the age of key Ediacaran fossil assemblages. *GSA Bulletin* 127:250–265.
- O'Brien, L. J., and J.-B. Caron. 2012. A new stalked filter-feeder from the Middle Cambrian Burgess Shale, British Columbia, Canada. PLoS ONE 7:e29233.
- O'Brien, L. J., and J.-B. Caron. 2016. Paleocommunity analysis of the Burgess Shale Tulip Beds, Mount Stephen, British Columbia: comparison with the Walcott Quarry and implications for community variation in the Burgess Shale. *Paleobiology* 42:27–53.
- Ortega-Hernández, J., J. Yang, and X.-G. Zhang. 2018. Fuxianhuiids. Current Biology 28:719–736.
- Ortega-Hernández, J., R. Lerosey-Aubril, and S. Pates. 2019. Proclivity of nervous system preservation in Cambrian Burgess Shale-type deposits. *Proceedings of the Royal Society B* 286:20192370.
- Ortega-Hernández, J., R. Lerosey-Aubril, S. R. Losso, and J. C. Weaver. 2022. Neuroanatomy in a middle Cambrian mollisoniid and the ancestral nervous system organization of chelicerates. *Nature Communications* 13:1–11.
- Ou, Q., J. Han, Z. Zhang, D. Shu, G. Sun, and G. Mayer. 2017. Three Cambrian fossils assembled into an extinct body plan of cnidarian affinity. *Proceedings of the National Academy of Sciences USA* 114:8835–8840.
- Ou, Q., J. Vannier, X. Yang, A. Chen, H. Mai, D. Shu, J. Han, D. Fu, R. Wang, and G. Mayer. 2020. Evolutionary trade-off in reproduction of Cambrian arthropods. *Science Advances* 6.
- Paine, R. T. 1969. A note on trophic complexity and community stability. American Naturalist 103:91–93.
- Park, T.-Y. S., M. L. Nielsen, L. A. Parry, M. V. Sørensen, M. Lee, J. H. Kihm, I. Ahn, et al. 2024. A giant stem-group chaetognath. Science Advances 10: eaax6678.
- Parry, L. A., and J.-B. Caron. 2019. Canadia spinosa and the early evolution of the annelid nervous system. Science Advances 5:eaax5858.
- Parry, L. A., R. Lerosey-Aubril, J. C. Weaver, and J. Ortega-Hernández. 2021. Cambrian comb jellies from Utah illuminate the early evolution of nervous and sensory systems in ctenophores. *iScience* 24:102943.
- Paterson, J. R., G. D. Edgecombe, and M. S. Y. Lee. 2019. Trilobite evolutionary rates constrain the duration of the Cambrian explosion. *Proceedings of* the National Academy of Sciences USA 116:4394–4399.
- Paterson, J. R., G. D. Edgecombe, and D. C. García-Bellido. 2020. Disparate compound eyes of Cambrian radiodonts reveal their developmental growth mode and diverse visual ecology. *Science Advances* 6:eabc6721.
- Pates, S., A. C. Daley, D. A. Legg, and I. A. Rahman. 2021a. Vertically migrating *Isoxys* and the early Cambrian biological pump. *Proceedings of the Royal Society B* 288:20210464.
- Pates, S., R. Lerosey-Aubril, A. C. Daley, C. Kier, E. Bonino, and J. Ortega-Hernández. 2021b. The diverse radiodont fauna from the Marjum Formation of Utah, USA (Cambrian: Drumian). *PeerJ* 9:1–43.

- Peel, J. S. 2015. Failed predation, commensalism and parasitism on lower Cambrian linguliformean brachiopods. *Alcheringa* 39:149–163.
- Peel, J. S. 2017. Feeding behaviour of a new worm (Priapulida) from the Sirius Passet Lagerstätte (Cambrian Series 2, Stage 3) of North Greenland (Laurentia). *Palaeontology* **60**:795–805.
- Penny, A., and B. Kröger. 2019. Impacts of spatial and environmental differentiation on early Palaeozoic marine biodiversity. *Nature Ecology and Evolution* 3:1655–1660.
- Penny, A. M., R. Wood, A. Curtis, F. Bowyer, R. Tostevin, and K.-H. Hoffman. 2014. Ediacaran metazoan reefs from the Nama Group, Namibia. *Science* 344:1504–1506.
- Petchey, O. L., and K. J. Gaston. 2006. Functional diversity: back to basics and looking forward. *Ecology Letters* 9:741-758.
- Pflug, H. D. 1966. Neue Fossilreste aus den Nama-Schichten in Südwest-Afrika. Paläontologische Zeitschrift 40:14–25.
- Plotnick, R. E., S. Q. Dornbos, and J. Chen. 2010. Information landscapes and sensory ecology of the Cambrian Radiation. *Paleobiology* 36:303–317.
- Potin, G. J.-M., and A. C. Daley. 2023. The significance of Anomalocaris and other Radiodonta for understanding paleoecology and evolution during the Cambrian explosion. Frontiers in Earth Science 11:1160285. https://doi.org/ 10.3389/feart.2023.1160285.
- Pratt, B. R. 1998. Probable predation on Upper Cambrian trilobites and its relevance for the extinction of soft-bodied Burgess Shale-type animals. *Lethaia* 31:73–88.
- Rahman, I. A., and S. Zamora. 2024. Origin and early evolution of echinoderms. Annual Review of Earth and Planetary Sciences 52:295–320.
- Rahman, I. A., S. A. F. Darroch, R. A. Racicot, and M. Laflamme. 2015a. Suspension feeding in the enigmatic Ediacaran organism *Tribrachidium* demonstrates complexity of Neoproterozoic ecosystems. *Science Advances* 1:e1500800.
- Rahman, I. A., S. Zamora, P. L. Falkingham, and J. C. Phillips. 2015b. Cambrian cinctan echinoderms shed light on feeding in the ancestral deuterostome. *Proceedings of the Royal Society B* 282:20151964.
- Rahman, I. A., J. O'Shea, S. Lautenschlager, and S. Zamora. 2020. Potential evolutionary trade-off between feeding and stability in Cambrian cinctan echinoderms. *Palaeontology* 63:689–701.
- Reaka-Kudla, M. L., D. E. Wilson, and E. O. Wilson. 1996. Biodiversity II: understanding and protecting our biological resources. Joseph Henry Press, Washington, D.C.
- Reichman, O. J., and E. W. Seabloom. 2002. Ecosystem engineering: a trivialized concept? Response from Reichman and Seabloom. *Trends in Ecology* and Evolution 17:308.
- Riisgård, H., and P. Manríquez. 1997. Filter-feeding in fifteen marine ectoprocts (Bryozoa): particle capture and water pumping. *Marine Ecology Progress Series* 154:223–239.
- Ruiter, P. C. de, A.-M. Neutel, and J. C. Moore. 1995. Energetics, patterns of interaction strengths, and stability in real ecosystems. *Science* 269:1257–1260.
- Schiffbauer, J. D., J. W. Huntley, G. R. O'Neil, S. A. F. Darroch, M. Laflamme, and Y. Cai. 2016. The latest Ediacaran wormworld fauna: setting the ecological stage for the Cambrian Explosion. GSA Today 26:4–11.
- Schoenemann, B. 2006. Cambrian view. Palaeoworld 15:307-314.
- Schoenemann, B., and E. N. K. Clarkson. 2011. Eyes and vision in the Chengjiang arthropod *Isoxys* indicating adaptation to habitat. *Lethaia* 44:223–230.
- Schoenemann, B., H. Pärnaste, and E. N. K. Clarkson. 2017. Structure and function of a compound eye, more than half a billion years old. *Proceedings* of the National Academy of Sciences USA 114:13489–13494.
- Seilacher, A. 1994. From biomats to benthic agriculture: a biohistoric revolution. Pp. 97–105 in W. S. Krumbein, D. M. Paterson, and L. S. Stal, eds. *Biostabilization of Sediments*. Biblioteks und Informationsystem der Universität Oldenburg, Oldenburg, Germany.
- Seilacher, A., L. A. Buatois, and M. Gabriela Mángano. 2005. Trace fossils in the Ediacaran–Cambrian transition: behavioral diversification, ecological turnover and environmental shift. *Palaeogeography, Palaeoclimatology, Palaeoecology* 227:323–356.

- Sepkoski, J. J. 1978. A kinetic model of Phanerozoic taxonomic diversity I. Analysis of marine orders. *Paleobiology* 4:223–251.
- Serezhnikova, E., and A. Y. Ivantsov. 2007. *Fedomia mikhaili*—a new spiculebearing organism of sponge grade from the Vendian (Ediacaran) of the White Sea, Russia. *Palaeoworld* 16:319–324.
- Servais, T., B. Cascales-Miñana, D. A. T. Harper, B. Lefebvre, A. Munnecke, W. Wang, and Y. Zhang. 2023. No (Cambrian) explosion and no (Ordovician) event: a single long-term radiation in the early Palaeozoic. *Palaeogeography, Palaeoclimatology, Palaeoecology* 623:111592.
- Shannon, C. E. 1948. A mathematical theory of communication. Bell System Technical Journal 27:379–423.
- Shen, B., L. Dong, S. Xiao, and M. Kowalewski. 2008. The Avalon Explosion: evolution of Ediacara morphospace. *Science* 319:81–84.
- Shu, D.-G., H.-L. Luo, S. Conway Morris, X.-L. Zhang, S.-X. Hu, L. Chen, J. Han, M. Zhu, Y. Li, and L.-Z. Chen. 1999. Lower Cambrian vertebrates from south China. *Nature* 402:42–46.
- Shu, D., S. Conway Morris, J. Han, Z.-F. Zhang, K. Yasui, P. Janvier, L. Chen, et al. 2003. Head and backbone of the Early Cambrian vertebrate Haikouichthys. Nature 421:526–529.
- Small, A., W. H. Adey, and D. Spoon. 1998. Are current estimates of coral reef biodiversity too low? The view through the window of a microcosm. *Atoll Research Bulletin* 458:1–20.
- Smith, M. R. 2013. Nectocaridid ecology, diversity, and affinity: early origin of a cephalopod-like body plan. *Paleobiology* 39:297–321.
- Smith, M. R., and J.-B. Caron. 2015. *Hallucigenia*'s head and the pharyngeal armature of early ecdysozoans. *Nature* **523**:75–78.
- Sokolov, B. 1972. The Vendian stage in Earth history. XXIV IGC Session. Reports of Soviet Geologists. International Geological Congress, Abstracts—Congres Geologique Internationale, Resumes 24:243
- Sperling, E. A., and J. Vinther. 2010. A placozoan affinity for *Dickinsonia* and the evolution of late Proterozoic metazoan feeding modes. *Evolution and Development* 12:201–209.
- Sperling, E. A., K. J. Peterson, and M. Laflamme. 2011. Rangeomorphs, *Thectardis* (Porifera?) and dissolved organic carbon in the Ediacaran oceans. *Geobiology* 9:24–33.
- Sprigg, R. 1947. Early Cambrian (?) jellyfishes from the flinders ranges, South Australia. Transactions of the Royal Society of South Australia 71:212–224.
- Stachacz, M. 2016. Ichnology of the Cambrian Ociesęki Sandstone Formation (Holy Cross Mountains, Poland). Annales Societatis Geologorum Poloniae 86:291–328.
- Stanley, S. M. 2008. Predation defeats competition on the seafloor. *Paleobiology* 34:1-21.
- Steiner, M., M. Zhu, Y. Zhao, and B.-D. Erdtmann. 2005. Lower Cambrian Burgess Shale-type fossil associations of South China. *Palaeogeography*, *Palaeoclimatology*, *Palaeoecology* 220:129–152.
- Strang, K. M., H. A. Armstrong, and D. A. T. Harper. 2016. Minerals in the gut: scoping a Cambrian digestive system. *Royal Society Open Science* 3:160420.
- Suarez, P. A., and S. P. Leys. 2022. The sponge pump as a morphological character in the fossil record. *Paleobiology* **48**:446–461.
- Sun, H., F. Zhao, M. Steiner, G. Li, L. Na, B. Pan, Z. Yin, H. Zeng, H. Van Iten, and M. Zhu. 2020. Skeletal faunas of the lower Cambrian Yu'anshan Formation, eastern Yunnan, China: Metazoan diversity and community structure during the Cambrian Age 3. *Palaeogeography, Palaeoclimatology, Palaeoecology* 542:109580.
- Suprenaut, R. L., J. G. Gehling, and M. L. Droser. 2020. Biological and ecological insights from the preservational variability of *Funisia dorothea*, Ediacara Member, South Australia. *Palaios* 35:359–376.
- Szaniawski, H. 1983. Structure of protoconodont elements. *Fossils and Strata* 15:21–27.
- Szaniawski, H. 2002. New evidence for the protoconodont origin of chaetognaths. Acta Palaeontologica Polonica 47:405–419.
- Tanaka, G., X. Hou, X. Ma, G. D. Edgecombe, and N. J. Strausfeld. 2013. Chelicerate neural ground pattern in a Cambrian great appendage arthropod. *Nature* 502:364–367.
- Tarhan, L. G., M. L. Droser, and J. G. Gehling. 2015. Depositional and preservational environments of the Ediacara Member, Rawnsley Quartzite (South Australia): assessment of paleoenvironmental proxies and the timing

of "ferruginization." *Palaeogeography, Palaeoclimatology, Palaeoecology* **434**:4–13.

- Tarhan, L. G., M. L. Droser, J. G. Gehling, and M. P. Dzaugis. 2017. Microbial mat sandwiches and other anactualistic sedimentary features of the Ediacara Member (Rawnsley Quartzite, South Australia): implications for interpretation of the Ediacaran sedimentary record. *Palaios* 32:181–194.
- Tilman, D. 1994. Competition and biodiversity in spatially structured habitats. *Ecology* 75:2–16.
- Topper, T. P., L. E. Holmer, and J.-B. Caron. 2014. Brachiopods hitching a ride: an early case of commensalism in the middle Cambrian Burgess Shale. *Scientific Reports* 4:6704.
- Topper, T. P., L. C. Strotz, L. E. Holmer, and J.-B. Caron. 2015. Survival on a soft seafloor: life strategies of brachiopods from the Cambrian Burgess Shale. *Earth-Science Reviews* 151:266–287.
- Usami, Y. 2006. Theoretical study on the body form and swimming pattern of Anomalocaris based on hydrodynamic simulation. Journal of Theoretical Biology 238:11–17.
- Vannier, J. 2007. Early Cambrian origin of complex marine ecosystems. Pp. 81–100 in M. Williams, A. M. Haywood, F. J. Gregory, and D. N. Schmidt, eds. Deep-time perspectives on climate change: marrying the signal from computer models and biological proxies. Geological Society of London, London.
- Vannier, J. 2012. Gut contents as direct indicators for trophic relationships in the Cambrian marine ecosystem. *PLoS ONE* 7:e52200.
- Vannier, J., and J. Chen. 2000. The Early Cambrian colonization of pelagic niches exemplified by *Isoxys* (Arthropoda). *Lethaia* 33:295–311.
- Vannier, J. M. C., and J. Chen. 2005. Early Cambrian food chain: new evidence from fossil aggregates in the Maotianshan Shale Biota, SW China. *Palaios* 20:3–26.
- Vannier, J., D. C. García-Bellido, S. X. Hu, and A. L. Chen. 2009. Arthropod visual predators in the early pelagic ecosystem: evidence from the Burgess Shale and Chengjiang biotas. *Proceedings of the Royal Society B* 276:2567–2574.
- Vannier, J., J. Liu, R. Lerosey-Aubril, J. Vinther, and A. C. Daley. 2014. Sophisticated digestive systems in early arthropods. *Nature Communications* 5:1–9.
- Vannier, J., C. Aria, R. S. Taylor, and J.-B. Caron. 2018. Waptia fieldensis Walcott, a mandibulate arthropod from the middle Cambrian Burgess Shale. Royal Society Open Science 5:172206.
- Vinn, O. 2017. Early symbiotic interactions in the Cambrian. Palaios 32:231–237.
- Vinn, O. 2018. Traces of predation in the Cambrian. *Historical Biology* 30:1043–1049.
- Vinther, J., M. Stein, N. R. Longrich, and D. A. T. Harper. 2014. A suspension-feeding anomalocarid from the Early Cambrian. *Nature* 507:496–499.
- Waggoner, B. 2003. The Ediacaran Biotas in space and time. Integrative and Comparative Biology 43:104–113.
- Walcott, C. D. 1911a. Cambrian geology and paleontology II: Middle Cambrian Merostomata. Smithsonian Miscellaneous Collections 57:18–40.
- Walcott, C. D. 1911b. Cambrian geology and paleontology II: Middle Cambrian Holothurians and Medusae. Smithsonian Miscellaneous Collections 57:42–68.
- Walcott, C. D. 1911c. Cambrian geology and paleontology II: Middle Cambrian annelids. Smithsonian Miscellaneous Collections 57:110–144.
- Walcott, C. D. 1912. Cambrian geology and paleontology II: Middle Cambrian Branchiopoda, Malacostraca, Trilobita and Merostomata. *Smithsonian Miscellaneous Collections* 57:145–228.
- Walcott, C. D. 1918. Cambrian geology and paleontology IV: appendages of trilobites. Smithsonian Miscellaneous Collections 67:116–217.
- Walcott, C. D. 1920. Cambrian geology and paleontology IV: Middle Cambrian Spongiae. *Smithsonian Miscellaneous Collections* 67:261–324.
- Wan, B., X. Yuan, Z. Chen, C. Guan, K. Pang, Q. Tang, and S. Xiao. 2016. Systematic description of putative animal fossils from the early Ediacaran Lantian Formation of South China. *Palaeontology* 59:515–532.
- Wang, Y., Y. Wang, W. Du, and X. Wang. 2014. The correlation between macroscopic algae and metazoans in the Ediacaran: a case study on the Wenghui biota in northeastern Guizhou, South China. Australian Journal of Earth Sciences 61:967–977.

- Warren, L. V., T. R. Fairchild, C. Gaucher, P. C. Boggiani, D. G. Poiré, L. E. Anelli, and J. C. G. Inchausti. 2011. Corumbella and in situ Cloudina in association with thrombolites in the Ediacaran Itapucumi Group, Paraguay. Terra Nova 23:382–389.
- Warren, L. V., F. Quaglio, M. G. Simões, C. Gaucher, C. Riccomini, D. G. Poiré, B. T. Freitas, P. C. Boggiani, and A. N. Sial. 2017. Cloudina-Corumbella-Namacalathus association from the Itapucumi Group, Paraguay: increasing ecosystem complexity and tiering at the end of the Ediacaran. Precambrian Research 298:79–87.
- Warren, L. V., L. A. Buatois, M. G. Mángano, M. G. Simões, M. G. M. Santos, D. G. Poiré, C. Riccomini, and M. L. Assine. 2020. Microbially induced pseudotraces from a Pantanal soda lake, Brazil: alternative interpretations for Ediacaran simple trails and their limits. *Geology* 48:857–861.
- Warren, L. V., L. Inglez, S. Xiao, L. A. Buatois, M. G. Mángano, J. Okubo, L. Alessandretti, *et al.* 2023. Evolutionary, paleoecological, and biostratigraphic implications of the Ediacaran–Cambrian interval in West Gondwana. *GSA Bulletin* 135:3102–3111.
- Whittington, H. B. 1971. Redescription of Marrella splendens (Trilobitoidea) from the Burgess Shale, Middle Cambrian, British Columbia. Geological Survey of Canada, Bulletin 209:1–24.
- Whittington, H. B. 1975. The enigmatic animal Opabinia regalis, middle Cambrian, Burgess Shale, British Columbia. *Philosophical Transactions of* the Royal Society B 271:1–43.
- Whittington, H. B., and D. E. G. Briggs. 1985. The largest Cambrian animal, Anomalocaris, Burgess Shale, British Columbia. *Philosophical Transactions* of the Royal Society B 309:569–609.
- Wiegand, T., and K. A. Moloney. 2013. Handbook of spatial point-pattern analysis in ecology. CRC Press, Boca Raton, Fla.
- Wilby, P. R., J. N. Carney, and M. P. A. Howe. 2011. A rich Ediacaran assemblage from eastern Avalonia: evidence of early widespread diversity in the deep ocean. *Geology* 39:655–658.
- Wolfe, J. M., and J. Ortega-Hernández. 2021. Evolution: Oh, my Cambrian nerves. Current Biology 31:R1132–R1135.
- Wood, D. A., R. W. Dalrymple, G. M. Narbonne, J. G. Gehling, and M. E. Clapham. 2003. Paleoenvironmental analysis of the late Neoproterozoic Mistaken Point and Trepassey formations, southeastern Newfoundland. *Canadian Journal of Earth Sciences* 40:1375–1391.
- Wood, R., A. Curtis, A. Penny, A. Yu. Zhuravlev, S. Curtis-Walcott, S. Iipinge, and F. Bowyer. 2017. Flexible and responsive growth strategy of the Ediacaran skeletal *Cloudina* from the Nama Group, Namibia. *Geology* 45:259–262.
- Wood, R., A. G. Liu, F. Bowyer, P. R. Wilby, F. S. Dunn, C. G. Kenchington, J. F. H. Cuthill, E. G. Mitchell, and A. Penny. 2019. Integrated records of environmental change and evolution challenge the Cambrian Explosion. *Nature Ecology and Evolution* 3:528–538.
- Wu, Y., S. Pates, J. Ma, W. Lin, Y. Wu, X. Zhang, and D. Fu. 2022. Addressing the Chengjiang conundrum: a palaeoecological view on the rarity of hurdiid radiodonts in this most diverse early Cambrian Lagerstätte. *Geoscience Frontiers* 13:101430.
- Wu, Y., S. Pates, D. Pauly, X. Zhang, and D. Fu. 2024. Rapid growth in a large Cambrian apex predator. *National Science Review* 11:nwad284.
- Xiao, S., and M. Laflamme. 2009. On the eve of animal radiation: phylogeny, ecology and evolution of the Ediacara biota. *Trends in Ecology and Evolution* 24:31–40.
- Xiao, S., Z. Chen, C. Zhou, and X. Yuan. 2019. Surfing in and on microbial mats: oxygen-related behavior of a terminal Ediacaran bilaterian animal. *Geology* 47:1054–1058.
- Xiao, S., J. G. Gehling, S. D. Evans, I. V. Hughes, and M. L. Droser. 2020. Probable benthic macroalgae from the Ediacara Member, South Australia. *Precambrian Research* 350:105903.
- Xiao, S., Z. Chen, K. Pang, C. Zhou, and X. Yuan. 2021. The Shibantan Lagerstätte: insights into the Proterozoic–Phanerozoic transition. *Journal* of the Geological Society 178;jgs2020–2135.
- Yang, J., J. Ortega-Hernández, N. J. Butterfield, and X. G. Zhang. 2013. Specialized appendages in fuxianhuiids and the head organization of early euarthropods. *Nature* 494:468–471.

- Yang, J., J. Ortega-Hernández, S. Gerber, N. J. Butterfield, J. Hou, T. Lan, and X. Zhang. 2015. A superarmored lobopodian from the Cambrian of China and early disparity in the evolution of Onychophora. *Proceedings* of the National Academy of Sciences USA 112:8678–8683.
- Yang, J., J. Ortega-Hernández, D. A. Legg, T. Lan, J. B. Hou, and X. G. Zhang. 2018. Early Cambrian fuxianhuiids from China reveal origin of the gnathobasic protopodite in euarthropods. *Nature Communications* 9:470.
- Yuan, X., Z. Chen, S. Xiao, B. Wan, C. Guan, W. Wang, C. Zhou, and H. Hua. 2013. The Lantian biota: a new window onto the origin and early evolution of multicellular organisms. *Chinese Science Bulletin* 58:701–707.
- Zacaï, A., J. Vannier, and R. Lerosey-Aubril. 2016. Reconstructing the diet of a 505-million-year-old arthropod: *Sidneyia inexpectans* from the Burgess Shale fauna. *Arthropod Structure and Development* **45**:200–220.
- Zakrevskaya, M. 2014. Paleoecological reconstruction of the Ediacaran benthic macroscopic communities of the White Sea (Russia). *Palaeogeography, Palaeoclimatology, Palaeoecology* **410**:27–38.
- Zhai, D., J. Ortega-Hernández, J. M. Wolfe, X. Hou, C. Cao, and Y. Liu. 2019a. Three-dimensionally preserved appendages in an Early Cambrian stem-group Pancrustacean. *Current Biology* 29:171–177.
- Zhai, D., M. Williams, D. J. Siveter, T. H. P. Harvey, R. S. Sansom, S. E. Gabbott, D. J. Siveter, et al. 2019b. Variation in appendages in early Cambrian bradoriids reveals a wide range of body plans in stem-euarthropods. *Communications Biology* 2:329.
- Zhang, X., W. Liu, and Y. Zhao. 2008. Cambrian Burgess Shale-type Lagerstätten in South China: distribution and significance. Gondwana Research 14:255–262.
- Zhang, Z., S. P. Robson, C. Emig, and D. Shu. 2008. Early Cambrian radiation of brachiopods: a perspective from South China. *Gondwana Research* 14:241–254.
- Zhang, Z., J. Han, Y. Wang, C. C. Emig, and D. Shu. 2009. Epibionts on the lingulate brachiopod Diandongia from the Early Cambrian Chengjiang Lagerstätte, South China. *Proceedings of the Royal Society B* 277:175–181.
- Zhang, X., C. Chang, L. Cui, and Y. Qiao. 2021. Ecosystem reconstruction during the Cambrian Explosion. *Paleontological Research* 25:305–314.

- Zhang, Z., Z. Zhang, J. Ma, P. D. Taylor, L. C. Strotz, S. M. Jacquet, C. B. Skovsted, F. Chen, J. Han, and G. A. Brock. 2021. Fossil evidence unveils an early Cambrian origin for Bryozoa. *Nature* 599:251–255.
- Zhao, F., M. Zhu, and S. Hu. 2010. Community structure and composition of the Cambrian Chengjiang biota. *Science China Earth Sciences* 53:1784–1799.
- Zhao, F., S. Hu, J.-B. Caron, M. Zhu, Z. Yin, and M. Lu. 2012. Spatial variation in the diversity and composition of the Lower Cambrian (Series 2, Stage 3) Chengjiang Biota, Southwest China. *Palaeogeography*, *Palaeoclimatology*, *Palaeoecology* 346–347:54–65.
- Zhao, F., D. J. Bottjer, S. Hu, Z. Yin, and M. Zhu. 2013. Complexity and diversity of eyes in Early Cambrian ecosystems. *Scientific Reports* 3:2751.
- Zhao, F., J.-B. Caron, D. J. Bottjer, S. Hu, Z. Yin, and M. Zhu. 2014. Diversity and species abundance patterns of the Early Cambrian (Series 2, Stage 3) Chengjiang Biota from China. *Paleobiology* 40:50–69.
- Zhao, Y., J. Vinther, L. A. Parry, F. Wei, E. Green, D. Pisani, X. Hou, G. D. Edgecombe, and P. Cong. 2019. Cambrian sessile, suspension feeding stem-group ctenophores and evolution of the comb jelly body plan. *Current Biology* 29:1112–1125.e2.
- Zhao, J., Y. Li, and P. A. Selden. 2023. A new primitive polychaete with eyes from the lower Cambrian Guanshan biota of Yunnan Province, China. *Frontiers in Ecology and Evolution* 11:1128070.
- Zhao, Y., X. Hou, and P. Cong. 2023. Tentacular nature of the "column" of the Cambrian diploblastic Xianguangia sinica. Journal of Systematic Palaeontology 21:2215787.
- Zhuravlev, A. Y., and R. A. Wood. 1995. Lower Cambrian reefal cryptic communities. *Palaeontology* 38:443–470.
- Zhuravlev, A. Y., and R. A. Wood. 2018. The two phases of the Cambrian Explosion. *Scientific Reports* 8:16656.
- Zhuravlev, A. Yu., E. B. Naimark, and R. A. Wood. 2015. Controls on the diversity and structure of earliest metazoan communities: Early Cambrian reefs from Siberia. *Earth-Science Reviews* 147:18–29.
- Zhuravlev, A. Y., E. G. Mitchell, F. Bowyer, R. Wood, and A. Penny. 2022. Increases in reef size, habitat and metacommunity complexity associated with Cambrian radiation oxygenation pulses. *Nature Communications* 13:7523.