


REVIEW

# Probing and manipulating the gut microbiome with chemistry and chemical tools

Pavan K. Mantravadi<sup>1</sup>, Basavaraj S. Kovi<sup>2</sup>, Sabbasani Rajasekhara Reddy<sup>3</sup>,  
Ganesh Pandian Namasivayam<sup>2</sup>, Karunakaran Kalesh<sup>4,5</sup> and Anutthaman Parthasarathy<sup>6</sup> 

<sup>1</sup>CMC and Analytical, Cytokinetics, South San Francisco, CA, USA; <sup>2</sup>Institute for Integrated Cell-Material Sciences (iCeMS), Kyoto University, Kyoto, Japan; <sup>3</sup>Department of Chemistry, School of Advanced Sciences, Vellore Institute of Technology (VIT), Vellore, India; <sup>4</sup>School of Health and Life Sciences, Teesside University, Middlesbrough, UK; <sup>5</sup>National Horizons Centre, Darlington, UK and <sup>6</sup>The School of Chemistry and Biosciences, University of Bradford, Bradford, UK

**Corresponding author:** Anutthaman Parthasarathy; Email: [aparthas@bradford.ac.uk](mailto:aparthas@bradford.ac.uk)

(Received 24 July 2024; revised 03 April 2025; accepted 07 April 2025)

## Abstract

The human gut microbiome represents an extended “second genome” harbouring about  $10^{15}$  microbes containing >100 times the number of genes as the host. States of health and disease are largely mediated by host–microbial metabolic interplay, and the microbiome composition also underlies the differential responses to chemotherapeutic agents between people. Chemical information will be the key to tackle this complexity and discover specific gut microbiome metabolism for creating more personalised interventions. Additionally, rising antibiotic resistance and growing awareness of gut microbiome effects are creating a need for non-microbicidal therapeutic interventions. We classify chemical interventions for the gut microbiome into categories like molecular decoys, bacterial conjugation inhibitors, colonisation resistance-stimulating molecules, “prebiotics” to promote the growth of beneficial microbes, and inhibitors of specific gut microbial enzymes. Moreover, small molecule probes, including click chemistry probes, artificial substrates for assaying gut bacterial enzymes and receptor agonists/antagonists, which engage host receptors interacting with the microbiome, are some other promising developments in the expanding chemical toolkit for probing and modulating the gut microbiome. This review explicitly excludes “biologics” such as probiotics, bacteriophages, and CRISPR to concentrate on chemistry and chemical tools like chemoproteomics in the gut-microbiome context.

**Keywords:** gut microbiome; chemistry; prebiotics; conjugation inhibitors; chemical probes

## Introduction

There are about  $10^{13}$ – $10^{15}$  symbiotic microbes residing inside and on the surface of a human being which collectively constitute the human microbiome (Turnbaugh et al., 2007). The microbiome plays a significant role in lifelong host health (Rackaityte and Lynch, 2020) and underlies a considerable proportion of the individual differences in drug metabolism (Zimmermann et al., 2019). Therefore, modulating the human microbiomes has triggered the interest of both academia and industry, and several interventions have been designed to either preserve or rebuild the function of the microbiome. In the period 2015–2018, over 80 microbiome modulators entered the preclinical phase, while 15 were in phase I trials, 5 in phase II, and 6 in phase III, according to the Pharmaprojects 2018 Microbiome Whitepaper (Ltd., I. U., 2018). The same report details that as of 2018, 10 modulators were in the pipeline for metabolic disorders, 21 for gastrointestinal disorders, and 24 for infectious diseases.

The gut (gastrointestinal system) harbours the most extensive human microbiome, which is critical for host metabolic and immune functions (Shreiner *et al.*, 2015). Further, a healthy microbiome also prevents pathogens from colonising the gut, a phenomenon known as colonisation resistance (CR) (Ducarmon *et al.*, 2019). The gut also contains the largest surface where immune system activity occurs inside the human body (Kraehenbuhl and Neutra, 1992) and the development of the immune system itself is a delicate dance of balancing the host versus the gut microbes (Randall and Mebius, 2014). The gut connects to various distal organs via two-way signalling and therefore, the gut microbiome (GM) maintains far more than just gut health (Schroeder and Bäckhed, 2016). GM dysfunction is implicated in the development of infections, gastrointestinal cancers as well as liver, respiratory, neurological, cardiac, metabolic, and autoimmune diseases (Wang *et al.*, 2017).

Antibiotics, in particular, cause deleterious changes to the function of the GM (Becattini *et al.*, 2016) and therefore preserving/restoring those functions is important. The antimicrobial resistance crisis has also led to a search for less indiscriminate therapeutics, which are GM friendly (Patangia *et al.*, 2022). Kang *et al.* showed that gut bacteria such as *Clostridium scindens* and *Clostridium sordellii* which perform 7 $\alpha$ -dehydroxylation of bile salts, also produced endogenous narrow-spectrum antibiotics derived from tryptophan, such as turbomycin A and 1-acetyl- $\beta$ -carboline which inhibit *Clostridioides difficile* (Kang *et al.*, 2019). Indole-3-propionic acid (IPA), another tryptophan metabolite that is produced by *Clostridium sporogenes*, inhibits a variety of mycobacteria, including drug-resistant *Mycobacterium tuberculosis* (Negatu *et al.*, 2020). IPA inhibited *M. tuberculosis* both *in vitro* and when administered in mice models via oral and intravenous routes (where it showed a seven-fold bacterial load reduction in the spleen (Negatu *et al.*, 2018, 2020)). GM-derived IPA can bind and powerfully induce the aryl hydrocarbon receptor (AHR; a major regulator of both innate and adaptive immunity) and therefore modulate the susceptibility to *M. tuberculosis* (Negatu *et al.*, 2020). The recovery of IPA in the serum (Negatu *et al.*, 2020) and the existence of the gut-lung (Schroeder and Bäckhed, 2016) and gut-spleen (Barrea *et al.*, 2017) axes explains how the GM can influence both lung and immune function remotely.

Endogenous narrow-spectrum peptide antibiotics with more complicated structures like bacteriocins also exist (Rea *et al.*, 2010) and could become available for research via solid-phase peptide synthesis since synthetic methods for cyclic peptides are rapidly improving (Bédard and Biron, 2018). Drug delivery targeted to different gut compartments (Hua, 2020) is already a burgeoning field. Therefore, chemically synthesised narrow-spectrum antibiotics could, in the near future, be delivered to specific gut compartments for directly or indirectly influencing the susceptibility and host-colonisation ability of major pathogens such as *M. tuberculosis* (Negatu *et al.*, 2020) and *C. difficile* (Kang *et al.*, 2019) as well as modulating host immunity, to prevent infections or aid recovery from infections.

Direct chemical manipulation of the GM has been the most challenging to perform in the absence of prior knowledge of the targets. However, in a pioneering study, Chen *et al.* devised an *in vitro* screening protocol and were able to use the cyclic D,L- $\alpha$ -peptides they identified via screening to change GM induced by a Western diet into one reflecting a low-fat diet (Chen *et al.*, 2020). This not only ameliorated atherosclerosis in mice, but also adjusted the levels of pro-inflammatory cytokines, short-chain fatty acids (SCFA), and bile acids (BAs) to healthy levels, while improving gut barrier integrity and T-cell function. They described their approach as “directed remodelling,” implying a deliberate manipulation of the GM in a predetermined manner from one state to another.

Research is moving away from largely cataloguing microbial strains to examining and understanding the molecular basis of the GM’s influence on human health (Rackaityte and Lynch, 2020). Therefore, we argue that chemistry and chemical information will play an important part in unravelling GM interactions and manipulating the GM to promote health. With this in mind, we focus on the roles of chemistry and chemoproteomics, while excluding “biologics” strategies such as probiotics, bacteriophages, and CRISPR. Narrow-spectrum antibiotics and directed chemical remodelling are only two recent examples of the potential of chemistry in the GM story. Whether preparing prebiotics, inhibiting bacterial conjugation in the gut, stimulating CR, probing GM-host interactions, or altering the GM composition to promote host health, the versatile toolkit of chemistry offers abundant opportunities to explore and modulate the GM.

## Molecules that preserve/restore the GM

These are classified based on their mode of action as shown in [Figure 1A](#) and some example chemical structures are shown in [Figure 1B](#).

### Prebiotics

Prebiotics are selectively fermented ingredients that trigger specific changes in the microbiome composition and activity to promote host health (Gibson et al., 2004). Safely administering live microbes and establishing their colonisation in the gut is difficult and faces regulatory hurdles, making small molecule interventions more attractive (Cully, 2019). Small molecules, especially endogenous metabolites can accumulate to high concentrations with negligible toxicity, remain stable in the systemic circulation, and obey the principles of pharmacokinetics. The major prebiotics are human milk oligosaccharides (HMOs), inulins (1 in [Figure 1B](#)), fructose oligosaccharides (FOS), xylooligosaccharides (XOS), mannan oligosaccharides (MOS), and galactooligosaccharides, which are polymers/oligomers of glucose, fructose, mannose, fucose, galactose, sialic acid, xylose, uronic acid, and arabinofuranose units linked together with  $\beta$ 2,  $\beta$ 3, and  $\beta$ 4 linkages (Enam and Mansell, 2019).

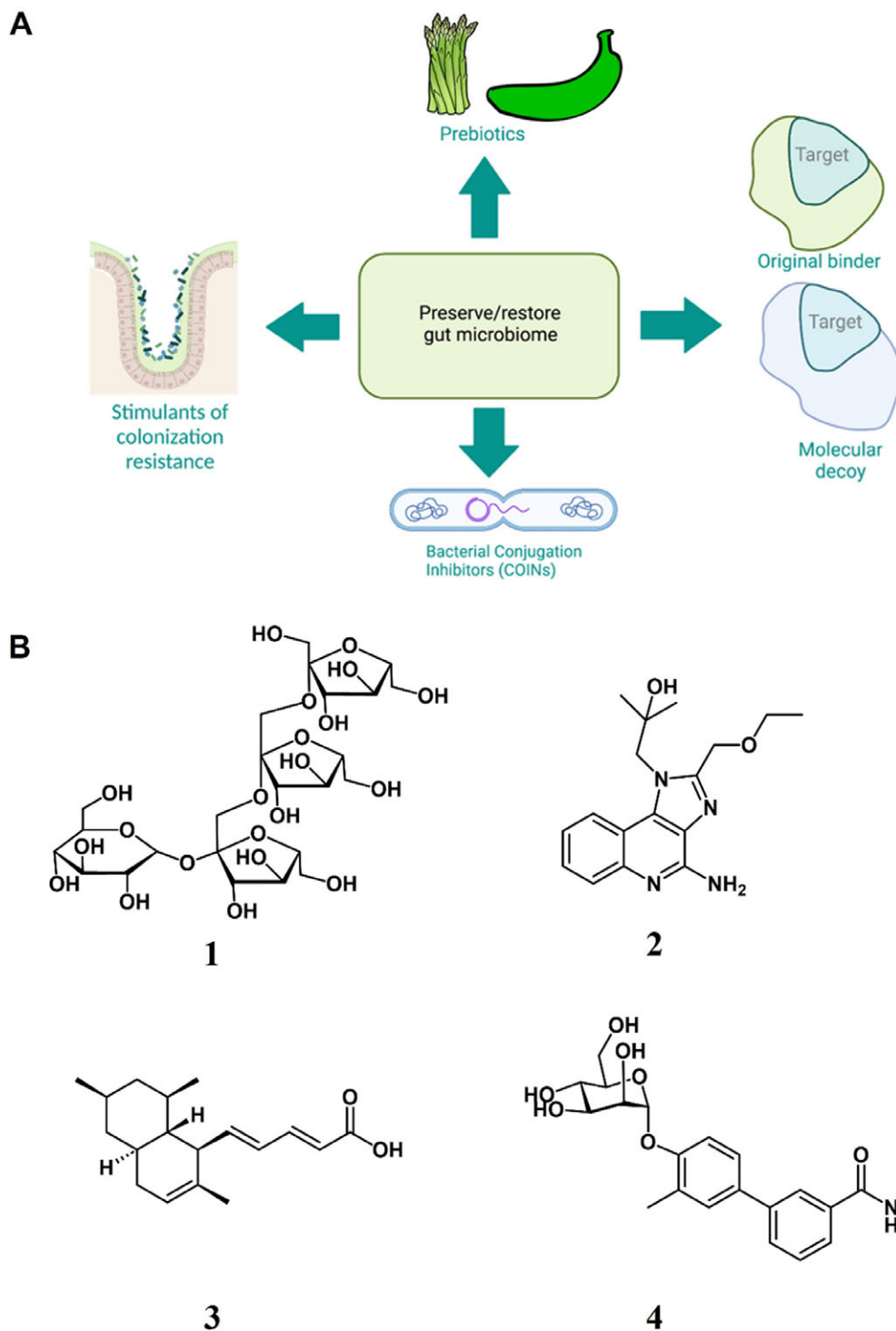
Developments in chemical synthesis are bringing the goal of complex carbohydrate assembly closer. Difficulties arise mainly from (1) the need to selectively protect and deprotect monosaccharides and (2) regioselectivity and stereoselectivity. Improved glycosylation strategies have been reported, which enables glycosyl donors to react in a specific order, yielding a single oligosaccharide product (Vohra et al., 2008). Automated glycan assembly (AGA) currently enables access to a maximum length of 100, while convergent block coupling of 30- and 31-mer oligosaccharide fragments made by AGA was used to make a multiple-branched 151-mer polymannoside (Joseph et al., 2020).

Enzymatic and chemoenzymatic processes offer better regioselectivity and stereoselectivity, along with fewer steps in the synthesis, which makes them faster and more cost effective (Li et al., 2019). For example, the HMO 2'-fucosyllactose (2'FL) has been synthesised in engineered *Escherichia coli* strains (Baumgärtner et al., 2013). One-pot multi-enzyme synthesis has been reported which employs glycosyltransferases to synthesise sialyl- and fucosyl-derivatives (Chen et al., 2015). Sialylated HMOs with high regioselectivity and stereoselectivity have been synthesised using a chemoenzymatic strategy, whereby automated solid-phase synthesis of the glycan backbone was followed by  $\alpha$ -(2,3)-sialyltransferase treatment (Fair et al., 2015). Interest in sustainable chemical feedstocks has led to method development for the conversion of lignocellulose biomass into valuable prebiotics such as XOS (Poletto et al., 2020).

Prebiotics can have synergistic interactions with approved drugs. Konjac MOS from the plant *Amorphophallus konjac* are prebiotics containing  $\beta$ -D-mannose and  $\beta$ -D-glucose residues linked by one to four linkages (Liu et al., 2015). The combined administration of the drug metformin and konjac MOS mitigates insulin resistance and glucose tolerance, while also improving islet and hepatic tissue function (Zheng et al., 2018). The beneficial effects were correlated with the reduced abundance of the Rikenellaceae family and the Clostridiales order, with an increased relative abundance of *Bifidobacterium pseudolongum*, *Akkermansia muciniphila*, and OTU05945 of family S24-7 (Zheng et al., 2018). Further studies focussing on prebiotic-drug interactions could lead to more targeted application of prebiotics in combination with approved drugs to mitigate the impact of specific diseases.

### Stimulants of CR

CR is a mechanism by which the gut microbiota protects itself against the incursion and establishment of largely harmful microorganisms. This protection can be accomplished by several routes, such as antimicrobial secretion, nutrient limitation, and stimulation of gut barrier integrity and the action of bacteriophages (Ducarmon et al., 2019). Disturbances to the gut resulting from the use of antibiotics, other drugs, or inflammation can reduce CR, allowing enteric pathogens such as *C. difficile*, *Salmonella*



**Figure 1.** (A) Functional classification of molecules to preserve/restore the gut microbiome. (B) Chemical diversity of molecules with microbiome preserving/restoring functions; 1 = General structure of inulins (endogenous prebiotic), 2 = resiquimod or R848 (synthetic stimulant of colonisation resistance); 3 = tanzawaic acid B or TZA-B (natural product colonisation inhibitor); and 4 = a mannose (mannose-containing decoy for urinary pathogens which preserves the gut microbiota).

*enterica* serovar Typhimurium, *E. coli*, *Shigella flexneri*, *Campylobacter jejuni*, *Vibrio cholerae*, *Yersinia enterocolitica*, and *Listeria monocytogenes*, to colonise the niches vacated by microbiome disruption (Sorbara and Pamer, 2019). Both endogenous molecules such as SCFA and tryptophan metabolites produced by the GM and exogenous synthetic small molecules can restore CR function. Synthetic molecules are beginning to be used in efforts to stimulate CR following disturbances to the GM, for example, after antibiotic administration. For example, vancomycin-resistant enterococci (VRE) flourish when CR is compromised following antibiotic treatment. A synthetic molecule, resiquimod or R848 (2 in Figure 1B), binds to a Toll-like receptor 7 that stimulates innate immune defences, leading to the restoration of CR against VRE by triggering the expression of the antimicrobial peptide Reg3 $\gamma$  (Abt et al., 2016). R848 can be taken orally and induces the secretion of interleukins IL-23 and IL-22.

### Bacterial conjugation inhibitors

Antibiotic resistance is spread by several mechanisms, including horizontal gene transfer mediated by plasmids. Analysis of *Bacteroidetes* strains sharing the intestinal niches of specific individual humans, demonstrated the extensive occurrence of horizontal gene transfer among those strains. In this case, the genetic elements exchanged coded for orphan DNA methylases, fimbriae synthesis proteins, novel metabolic enzymes, antibiotics, and proposed type VI secretion systems (T6SS) (Coyne et al., 2014). More recent studies have recorded extensive plasmid exchange in the gut environment using CRISPR-Cas spacer acquisition analysis in an *E. coli* strain (Munck et al., 2020). Unlike earlier studies that relied on phenotypic markers or post-transfer replication to detect mobile genetic elements, the spacer acquisition analysis reveals plasmid transfer in real time, and the results showed that the IncX plasmid type was most frequently transferred (Munck et al., 2020). Therefore, inhibiting bacterial conjugation in a bacteria-dense environment could enable the host to mitigate antibiotic-resistant infections. In general, resident bacteria in the healthy GM may be able to suppress the evolution of antibiotic resistance *in vivo*. However, the wide distribution of plasmid-borne resistance in the environment is well known and exposure to them might be common. Moreover, gut inflammation boosts plasmid transfer between pathogenic and commensal Enterobacteriaceae (Stecher et al., 2012). Therefore, inhibiting plasmid transfer in the gut is expected to promote host health, and conjugation inhibitors (COINs) are unlikely to disturb the GM composition, unlike conventional antibiotics. We describe a few known COINs, but some need to be further specifically tested in the gut environment.

Early studies to identify COINs unearthed many unspecific molecules that affected DNA replication or growth (Cabezón et al., 2017). Plant phenolics seems to be a good source of COINs and have yielded two molecules that specifically inhibited bacterial conjugation, namely rottlerin and 8-cinnamoyl-5,7-dihydroxy-2,2,6-trimethylchromene (Oyedemi et al., 2016). Screening of a library of over 12,000 NPs (NatChem library) based on high-throughput whole-cell-based assays enabled the discrimination between true COINs and false “hits” which merely affected cell growth, leading to the discovery of the COIN dehydrocrepnynic acid (DHCA) (Fernandez-Lopez et al., 2005). DHCA belongs to the chemical family of unsaturated fatty acids (UFAs), which is generally a good source of COINs. DHCA is derived from a tropical seed, and its supply is limited. However, it was used as the starting point for the synthesis of other COINs, particularly 2-hexadecynoic acid (2-HDA) and other 2-alkynoic fatty acids (2-AFAs), which specifically inhibited the transfer of a range of plasmids, including the common and highly infective IncF, in various bacteria (Getino et al., 2015). 2-HAD was later reported to prevent bacterial conjugation in the mouse gut (Palencia-Gándara et al., 2021). A series of UFA NPs called tanzawaic acids were discovered (tanzawaic acid B or TZA-B is depicted as (3 in Figure 1B); they mainly inhibited conjugation by the IncW- and IncFII-based plasmids. Other plasmids classified under the IncFI, IncI, IncL/M, IncX, and IncH incompatibility groups were less affected, whereas the IncN and IncP plasmids were unaffected (Getino et al., 2016).

Conjugation is driven by the type 4 secretion system, whose architecture is conserved in most bacteria, and contains the pilus, the core channel complex, the inner membrane platform, and the ATPases that

provide energy for substrate transport and pilus biogenesis (Cabezón *et al.*, 2015). Nicking the DNA to relax the plasmid, DNA transfer to the secretion channel, the transfer of pilin molecules during pilus biogenesis, and pilus biogenesis are performed by four distinct ATP-ase enzymes, among which carboxylic acid COINS were shown to target the last step (TrwD protein). Based on structural and computational data, the UFAs and AFAs were suggested to bind at the end of the N-terminal domain as well as at the beginning of the linker region that connects the N-terminal and C-terminal domains, likely hindering the swapping movements of the domains needed for the catalytic cycle (Ripoll-Rozada *et al.*, 2016).

### **Molecular decoys**

These molecules bind enteric pathogens and stimulate their elimination from the gastrointestinal tract. This binding is thought to “fool” pathogens by mimicking the receptors used by them to attach to the gut epithelia in the lower gastrointestinal tract. The global enteric diseases is substantial and cases may number in the hundreds of millions annually. HMOs act as soluble decoys for receptors and block the binding of enteric pathogens. Rotavirus infection is prevented most effectively by the HMO 2’FL, although several other HMOs also have similar inhibitory effects (Laucirica *et al.*, 2017). *C. jejuni* infects the mammalian gut and causes diarrhoea and sometimes also motor neuron paralysis. The infection is initiated by the bacterium binding to the fucosylated intestinal H(O) antigen (Fuc alpha 1, 2Gal beta 1, 4GlcNAc). However, FOS in human milk can act as decoys by binding to the pathogen, thereby preventing infection (Ruiz-Palacios *et al.*, 2003).

Uropathogenic *E. coli* (UPEC) uses the extracellular appendages called Type 1 pili to colonise the intestine by binding a mannosylated host receptor; the Type 1 pili are also essential for colonisation and infection in the bladder. Mannosides (**4** in Figure 1B) are small-molecule drugs bearing mannose group(s) that act as decoys by mimicking the mannosylated receptor and can clear both bladder and intestinal UPEC upon oral administration in mouse models, leaving the GM largely intact (Spaulding *et al.*, 2018). The decoy approach has been further extended to combat cholera, and in this case, nanotechnology is also employed. The *V. cholerae* toxin binds to the host receptor monosialotetrahexosylganglioside (GM1), and coating GM1 on the surface of polymeric nanoparticles was enough to reduce cyclic-AMP production in epithelia and fluid responses to live *V. cholerae* in both cell cultures and a mouse infection model (Das *et al.*, 2018). The modulation of disease via molecular mimicry extends to non-sugar molecules, such as metalloenzymes allows for the manipulation of the gut chemical environment using synthetic catalysts. A metalloporphyrin mimic of the enzyme superoxide dismutase could reduce lipid peroxidation levels and thereby shielded epithelial cells from damage in rats injected with the common antigen bacterial lipopolysaccharide (LPS) (Das *et al.*, 2018).

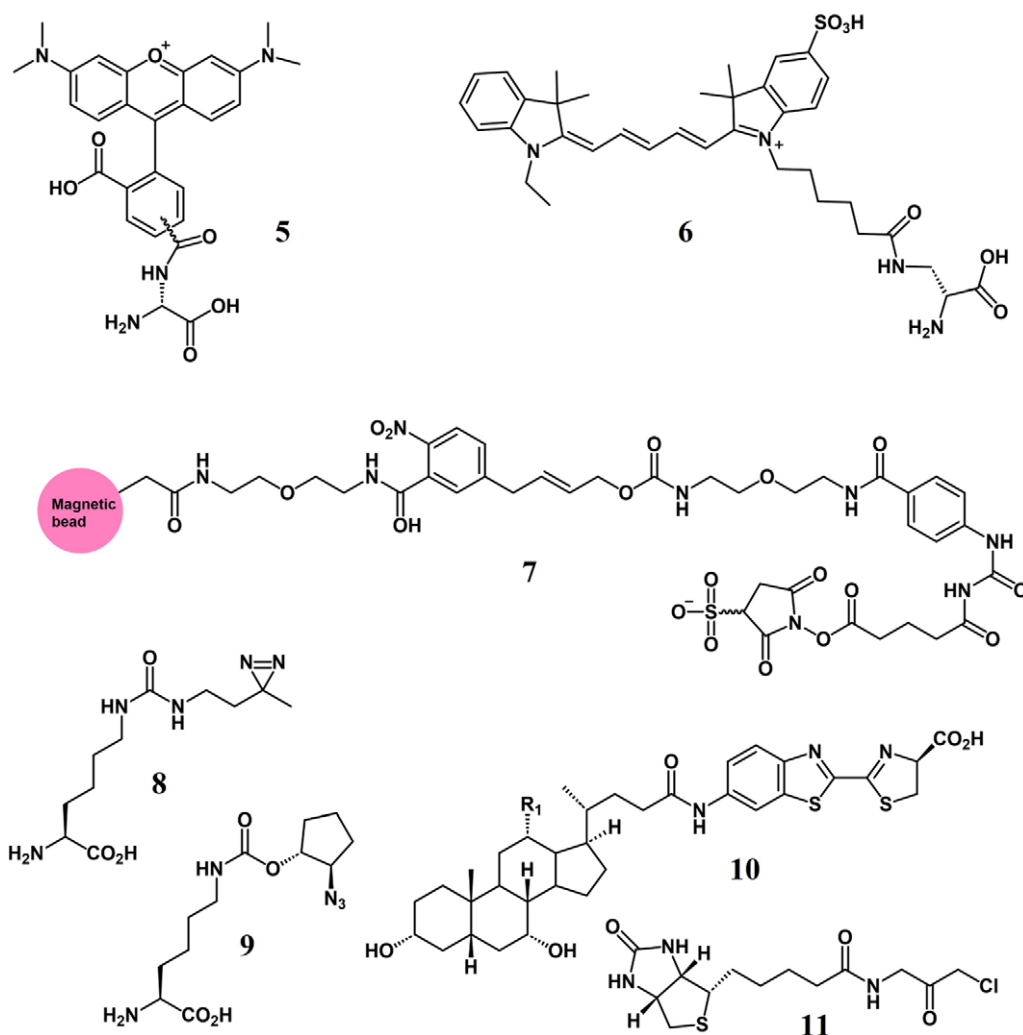
### **Chemical probes of the GM**

The majority of recent chemistry-oriented studies did not deal with direct chemical manipulation of the GM but focussed on probing the GM using bio-orthogonal strategies such as alkyne-cycloazide addition, Staudinger ligation, and tetrazine ligation to create “chemical reporters” (Zhang *et al.*, 2020). Bacterial surface glycans, peptidoglycans (PGNs), lipopolysaccharides, capsular polysaccharides (CPSs), glycoproteins, lipids, and other molecules, such as BAs have been labelled (Zhang *et al.*, 2020). In addition to such surface targeting, protein function may be probed by activity-based protein profiling (ABPP), which involves small molecules reacting with mechanistically related enzymes (Berger *et al.*, 2004). In ABPP, the probe usually contains a reactive group and a tag. Microbiota-metabolite interactions as well as microbiome composition and dynamics can be interrogated via ABPP, while chemoproteomics advances have made the detection of covalent probe-tagged proteins following the ABPP routine (Zhang *et al.*, 2020).



## Fluorophores

The most common tools for probing the GM are fluorophores, which may be attached to different types of other chemical entities. Commensal anaerobic bacteria, including *Bacteroides fragilis* when fed azide-labelled sugars, which subsequently conjugated with alkyne-fluorophores via click chemistry, facilitate the imaging of bacteria in live mice (Geva-Zatorsky et al., 2015). Three different bacterial surface molecules from the GM, which interact with the host immune system, namely LPS, CPS, and PGN, can be tracked (Hatzenpichler et al., 2014), helping to dissect host–microbe interactions. Azide-bearing amino acids when fed to complex gut microbial communities showed that newly synthesised proteins could be visualised *in situ* (Hatzenpichler et al., 2014). Two D-amino acid-based fluorescent probes, TADA and Cy5ADA (5,6 in Figure 2), which get incorporated into bacterial PGN, have been instrumental in enabling live monitoring of



**Figure 2.** Examples of chemical probes used to interrogate the GM – D-amino acid-based fluorescent probes = TADA (5) and Cy5ADA (6); a multifunctional probe showing different parts shaded in distinct colours = amine directed probe based on sulfo-N-hydroxysuccinimide (7); photoactive unnatural amino acid probes = DiZPK (8) and ACPK (9); a cysteine-targeted probe = Biotin-Gly-CMK (10); bioluminescent bile acid-luciferin conjugates for bile salt hydrolase (BSH) activity = series of compounds with H or OH at the positions R1 and R2 (11).

GM growth and division patterns in mice (Lin et al., 2020). Probes based on D-amino acids are also being used to track the viabilities of bacteria in faecal transplants by using sequential tagging (Wang et al., 2019). In this approach, the bacteria are treated with a probe before the transplantation and then the recipient mice are fed a second probe following the transplantation. Therefore, the bacteria surviving the process show the emission for both probes, enabling the identification of viable bacteria in the transplant (Wang et al., 2019).

### **Multifunctional selective probes**

Direct extraction from human faecal samples and release under mild conditions is possible using multifunctional chemoselective probes (Garg et al., 2018), allowing for the analysis of femtomole levels of metabolites with enhanced sensitivity. Probe 7 in Figure 2 is anchored at one end to magnetic beads, linked by a spacer to a novel *p*-nitrocinnamyloxycarbonyl biorthogonal cleavage site, while the reactive site features an amine-selective sulphydroxy-N-hydroxysuccinimide “warhead,” which reacts with metabolic amines (Garg et al., 2018). Since 2011, it has been possible to monitor enteric pathogens via the incorporation of the photoactive unnatural amino acids DiZPK and ACPK (8, 9 in Figure 2) into specific pathogen proteins, which react to form cross links revealing the interactions between the modified protein and its client proteins (Lin et al., 2011). This approach is enabling the direct identification of proteins involved in pathogenesis and acid-stress defence mechanisms, which is quite challenging to perform with conventional methods.

### **Simple reactive probes**

Sphinganine is a bioactive component of foods, but the GM also modifies them. The use of alkyne-tagged sphinganine allows for the identification of sphinganine-utilising GM strains based on labelling followed by a cell sorting workflow (Lee et al., 2021). The subsequent sequencing of the sorted bacteria revealed that this metabolism is nearly exclusively performed by members of the *Bacteroides* (Lee et al., 2021). An activity-based probe, Biotin-Gly-CMK (10 in Figure 2), has been used to differentiate between mice models harbouring “normal” human GM and “inflammatory bowel disease” (IBD) affected human GM, whereby a novel cysteine-reactive probe tagged several proteases and hydrolases in the IBD model, but not in the healthy controls (Mayers et al., 2017).

An elegant recent study by Nie et al. using a click chemistry strategy isolated and identified a previously unknown BA 3-succinylated cholic acid correlated with reduced progression of metabolic dysfunction-associated steatohepatitis in humans (Nie et al., 2024). Using this discovery, the authors were able to characterise an annotated  $\beta$ -lactamase in the GM member *Bacteroides uniformis* as the enzyme catalysing the 3-succinylation of CA (Mayers et al., 2017).

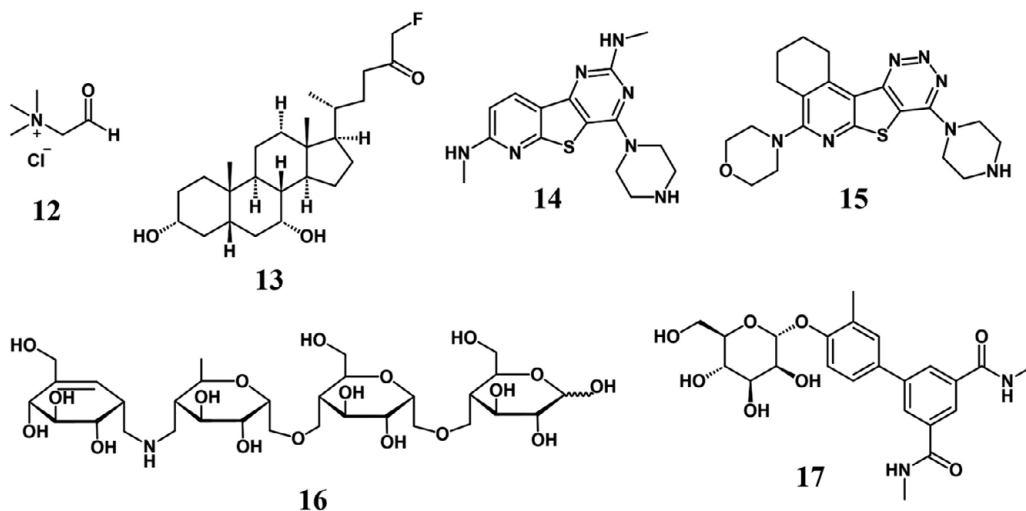
### **Bioluminescent probes**

Luciferin-based bioluminescent probes (11 in Figure 2) have been employed to detect bile salt hydrolase (BSH) activity in a wide variety of sample environments including purified enzymes, bacterial cells, faecal slurries as well as non-invasive imaging in mice and humans (Khodakivskyi et al., 2021). BSH activity releases luciferin from the conjugated BA, which can be further assayed using luciferase. These BA-luciferin probes were useful in demonstrating the stimulatory effect of prebiotics on BSH activity and as diagnostic tests that non-invasively detect the clinical IBD status in human patients (Khodakivskyi et al., 2021).

### **Modulating specific enzyme functions in the GM**

Targeting specific enzymes among the thousands of proteins actively produced by the gut microbes is a viable strategy for microbiome modulation.





**Figure 3.** Specific enzyme inhibition can be a strategy to selectively manipulate the gut microbiome, and some inhibitors of gut bacterial enzymes are shown. **12** = betaine aldehyde, inhibits choline TMA-lyase (CutC); **13** = fluoromethyl ketone suicide inhibitor of bile salt hydrolase (BSH); **14**, **15** = piperazine-containing  $\beta$ -glucuronidase inhibitors; **16** = acarbose, inhibits starch and pullulan utilisation; and **17** = M4284 mannoside, inhibits FimH in uropathogenic *E. coli*.

### Choline metabolism

A “chemically guided functional profiling” could be a strategy to uncover the presence of novel enzymes in the GM and subsequently, to modulate their function to achieve therapeutic effects. The conversion of choline into trimethylamine (TMA) by anaerobic gut bacteria is correlated with disease conditions in humans, and more specifically, the production of TMA in both isolated bacteria and complex communities can be inhibited by betaine aldehyde (**12** in Figure 3) (Orman et al., 2019). The identified target is GM choline TMA-lyase (CutC) and this opens up the scope for the development of other inhibitors.

### Bile salt metabolism

Bile salts have major effects on the physiology and virulence of *C. difficile*. When patients are restored to a *C. difficile*-resistant state, it is observed that the production of deoxycholate from cholate by 7 $\alpha$ -dehydroxylating gut bacteria occurs (Savidge and Sorg, 2019). Broad-spectrum antibiotics block the production of secondary BAs and kill the 7 $\alpha$ -dehydroxylating bacteria, thereby enabling *C. difficile* to colonise the gut (Savidge and Sorg, 2019). BSH enzymes expressed by the GM and bile salt metabolism affect the immune and metabolic processes via engaging host receptors. Therefore, inhibiting BSH enzymes would enable the dissection of the role of bile salts in host–microbe interactions. Screening a library of compounds, Adhikari et al. zeroed in on a covalent suicide inhibitor containing an  $\alpha$ -fluoromethyl ketone moiety (**13** in Figure 3), which reacts with the active site cysteine of BSH enzymes, as way to globally modulate BSH and understand their physiological roles (Adhikari et al., 2020).

### Glucuronidase inhibitors

$\beta$ -Glucuronidase (GUS) enzymes harboured by gut microbes can cause severe toxicity reactions to certain pharmaceuticals including cancer drugs; therefore, GUS inhibitors have been developed (**14**, **15** in Figure 3) to ameliorate these toxic side effects. Pellock et al. reported the discovery of piperazine-based GUS inhibitors by combining chemical biology, protein structural data and mass spectrometry with cell-based assays (Pellock et al., 2018). Their GUS inhibitors interrupt the catalytic cycle of the enzyme and

are substrate-dependent, binding to the catalytic intermediate by means of a piperazine-linked glucuronide. The inhibitor-glucuronide conjugates were detected by LC-MS (Santilli *et al.*, 2018).

### **Carbohydrate metabolism**

The prospects for chemical precision editing of the GM are improving due to an expansion in the knowledge of its metabolism. GM diversity is promoted by the metabolism of complex plant polysaccharides. Selective manipulation of polysaccharide metabolism without microbicidal effects has been achieved using a small molecule inhibitor, acarbose (**16** in Figure 3), which abolished the ability of *Bacteroides thetaiotaomicron* and *B. fragilis* to utilise potato starch and pullulan by interfering with the starch utilisation system (). Shifting the GM metabolic activity selectively in this non-lethal fashion alleviated colitis. Until recently, it was not known if single bacterial species or a small community is needed to drive the degradation of any highly complex polysaccharide. The most complex polysaccharide characterised in the gut environment is rhamnogalacturonan-II, which is depolymerised by *B. thetaiotaomicron* with the cleavage of 20 out of its 21 distinct glycosidic bonds (Ndeh *et al.*, 2017). Further analysis revealed that several previously unknown bacterial enzymes were responsible for the degradation of rhamnogalacturonan-II.

### **Miscellaneous inhibitors**

Zhu *et al.* showed that dysbiosis-linked gut inflammation caused by the expansion of facultative anaerobic Proteobacteria could be blocked via tungstate administration, which inhibits molybdenum-cofactor respiratory chain enzymes (Zhu *et al.*, 2018). The GM composition was undisturbed when tungstate was administered under homeostatic conditions. Recurrent infections of the urinary tract caused by UPEC occur in 30–50% of patients even after antibiotic treatment. This persistence is linked to the type 1 pilus adhesin, FimH, which binds mannose and aids the colonisation of the bladder surface. Type 1 pili were also shown to aid UPEC colonisation in the gut, and the administration of the high affinity FimH inhibitor mannoside M4284 (**17** in Figure 3) reduced gut colonisation and urinary tract infection caused by genetically distinct UPEC isolates, without disrupting the GM composition (Spaulding *et al.*, 2017).

### **Chemoproteomics tools for GM studies**

Over 1900 uncultured gut microbes were discovered in 2019 (Almeida *et al.*, 2019), showing enormous potential for finding metabolic diversity in the GM. Metagenomics projects, including the Human Microbiome Project show that identification of the biochemical functions of genes encoding metabolic enzymes in the human GM accurately is fraught with difficulty. In a survey of 139 stool metagenomes, only around 30% of them could be assigned a gene ontology or enzyme commission annotation; of these annotations, 50% have previously unknown functions (Joice *et al.*, 2014). Even in the case of enzymes/pathways that could be annotated, the gut microbiota contains many uncharacterised gene products detected in genomics/metagenomics analysis. Therefore, chemical information-based analyses (including analysis of chemical structure, chemical reactivity, and potential biological interaction partners) that predict potential GM metabolism and chemoproteomics methods are better placed to elucidate those “unknown” metabolic functions rather than purely metagenomics. Examples of the chemical information-based analysis include the design of gut-targeted drugs (Gil-Pichardo *et al.*, 2023) and predictions of potential drug/xenobiotic metabolism in the GM (Malwe *et al.*, 2023). Herein, however, we focus on some chemoproteomics/metabolomic tools developed for specific metabolite groups.

### **Enzyme-based sulphated metabolome analysis**

Sulphated compounds are derived from gut microbial transformation of dietary material and are related to disease states. Using an arylsulphatase enzyme to hydrolyse sulphated compounds and mass spectrometry-based metabolite analysis, Correia et al. have characterised and validated 235 sulphated metabolites in a single study, which were the products of gut microbiota and subsequent host transformations and discovered 11 previously unknown sulphated metabolites (Correia et al., 2020). The metabolites reported in this study could form the basis of classification of human subjects as harbouring high or low sulphate metabolising microbiota for future cohort studies. Further, the arylsulphatase-based method may be useful for discovering novel sulphated metabolites.

### **BSH and BA-based chemoproteomics**

As mentioned, BAs are secreted by the liver and are further converted into secondary BAs by the action of the GM. The latter participate in several processes, including the metabolism of glucose and lipids, and immune homeostasis. The key reaction of secondary BA biosynthesis is catalysed by BSH. BSH are bacterial cysteine hydrolases whose activity precedes other kinds of BA transformations (Devlin and Fischbach, 2015). Parasar et al. developed a strategy based on the covalent labelling of the active site cysteine using a substrate analogue (Parasar et al., 2019). When the substrate analogue is covalently bound, biorthogonal click chemistry could be applied to attach either a fluorescent contrast agent or a biotin affinity tag to the enzyme-bound analogue. In the first case, in situ imaging could be performed following gel electrophoresis, and in the second case, affinity purification was performed using streptavidin (the samples were subsequently analysed using proteomics).

While the expression of metagenomic fragments in well-studied model microbes showed that at least three distinct phyla possess BSH activities in the GM (Jones et al., 2008), genome-based strategies suffer from the issues of potential toxicity, incomplete coverage, incomplete BGC expression, unintended changes in enzyme levels and tissue localisation, all of which led to deviations from the physiologically relevant states of the BSH enzymes. By comparison, the covalent modification of the active sites of BSH enzymes coupled with proteomics has avoided many of the pitfalls associated with genome-based methods and enabled the direct identification of these enzymes.

While BAs promote CR, little was known about the target proteins affected in the gut pathogens inhibited by BA action. Photoaffinity probes based on chenodeoxycholic acid (CDCA) were able to crosslink many host and pathogen proteins in *Salmonella enterica* serovar Typhimurium infection models, of which direct protein inhibition by CDCA probes was reported for HilD, a key regulator of *Salmonella* pathogenesis and virulence (Yang et al., 2023). Chemical proteomics and photoaffinity labelling based on lithocholic acid were also used to identify a previously unknown BA-binding transcription factor called BapR in *C. difficile* (Forster et al., 2022).

### **Direct lysine-acylation chemoproteomics**

In a 2022 report, abundant post-translational lysine-acylation by reactive acyl-CoA species was discovered, whereby the acyl motifs found on several differentially expressed proteins corresponded to the metabolism of specific carboxylic acids in syntrophic bacteria (Muroski et al., 2022). The importance of cross-feeding in the gut environment, the abundance of SCFA and the ability to analyse the proteome for post-translational modifications without highly biased pre-enrichment, direct analysis of lysine acylation in the GM has good potential to shed light on metabolomic aspects.

### **Vitamin affinity probe chemoproteomics**

*Bacteroidetes* are one of the four major GM phylas; their genomes usually encode several B<sub>12</sub>-dependent enzymes, although they lack the ability of *de novo* cobamide synthesis (Shelton et al., 2019). It is therefore

likely that they could harbour B<sub>12</sub> transport proteins different at the sequence level from canonical *E. coli* counterparts. The use of B<sub>12</sub>-based affinity probes and subsequent application of chemoproteomics in *B. thetaiotaomicron* samples revealed the presence of proteins without previously unknown functions; one of these proteins, BtuH2, was shown to capture and transport B<sub>12</sub> directly *in vitro* and responsible for gut fitness of these bacteria in gnotobiotic mice (Putnam *et al.*, 2022).

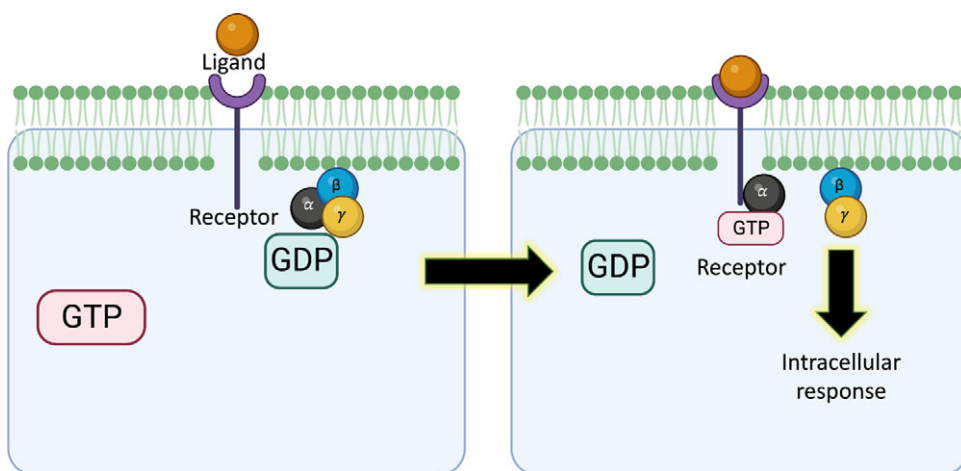
### Modulating host receptors

The intestinal surface senses bacterial surface molecules and GM metabolites through several types of cell-surface receptors, and further effects are exerted by receptor protein complexes inside various types of gut cells. Here, we briefly consider only selected agonists/antagonists linked to the GM activity of a few cell-surface, nuclear, and peroxisome-linked receptors.

### Cell-surface receptors

G protein-coupled receptors (GPCRs) are the largest membrane protein family in humans and sense their ligands through a mechanism outlined in Figure 4. GPCR complexes contain a transmembrane subunit (green in Figure 4) that binds a small molecule (ligand) at the cell surface, whereas a linked trimeric G-protein bound to GDP is located inside the cell. Once the ligand has been captured by the receptor subunit, then a conformational change occurs in the complex, allowing GTP to bind the trimeric G-protein, which usually dissociates, triggering an intracellular response via further downstream events. There are a variety of GPCRs in the gut for various microbial metabolites, such as SCFA (Husted *et al.*, 2017), BAs (Lefebvre *et al.*, 2009), and several other types of effectors (Cohen *et al.*, 2015). Gut bacteria synthesise molecules such as commendamide, which mimic the human (endogenous) ligands of GPCRs (Kroeze *et al.*, 2015).

A forward genetic screen (*i.e.*, trying to identify genes leading to a phenotype) based on the Tango  $\beta$ -arrestin recruitment assay (PRESTO-Tango) was able to measure the activation processes of almost all the non-olfactory human GPCRs (Chen *et al.*, 2019) and revealed several novel GPCR ligands such as L-phenylalanine secreted in the GM (Colosimo *et al.*, 2019). Several other ligands that bind GPCRs (including in immune and nerve cells) such as phenylpropanoic acid, cadaverine, 9-10-methylenehexadecanoic acid, and 12-methyltetradecanoic acid were identified in a high-



**Figure 4.** Molecular mechanism of G protein-coupled receptors on the cell surface. The ligand binds to the receptor protein, causing the G-protein subunits to disassemble and exchange bound GDP with GTP. The G-protein  $\alpha$ -subunit is bound to the receptor, whereas the other subunits signal to other proteins involved in intracellular responses. GTP hydrolysis drives the dissociation of the  $\alpha$ -subunit from the receptor and a return to the GDP-bound multi-subunit G-protein complex.

throughput screening of 241 GPCRs (Kovatcheva-Datchary et al., 2019), using seven gut microbes to represent a simplified human microbiome consortium (Subramanian et al., 2014; Kovatcheva-Datchary et al., 2019).

#### *Free fatty acid receptors*

SCFAs are sensed by specialised GPCRs called the free fatty acid receptor (FFAR), a family of cell surface receptors (Brown et al., 2003; Nilsson et al., 2003; Subramanian et al., 2014). FFAR2 and FFAR3 signalling links the GM and the  $\beta$ -cells in the pancreas and therefore are important targets in type-1 and type-2 diabetes (Nilsson et al., 2003; Priyadarshini et al., 2018). In pigs, the use of trans-glycosylated starches (TGS) led to downregulated FFAR2 via GM modulation, which decreased obesity (Priyadarshini et al., 2018). GM-derived SCFA and LPS also participate in the gut-lung immune axis because these molecules can travel to the lungs and modulate FFAR2/3 activity there (Husted et al., 2017).

#### *Hydroxy carboxylic acid receptor*

This is yet another class of GPCRs, which regulate immunity and energy homeostasis and sense hydroxycarboxylic acids. Most mammals have HCA1 that senses lactic acid, and HCA2 that senses 3-hydroxybutanoate and butyrate (Priyadarshini et al., 2021). Recently, a third hydroxy carboxylic acid receptor (HCAR) called HCA3 was detected in hominin genomes and described in humans; it senses and is potently activated by D-phenyllactic acid (Newman et al., 2018), which is produced as an antimicrobial by GM *Lactobacilli*. HCA2 is expressed in not only the intestinal epithelial cells, but also adipocytes, immune cells, hepatocytes, retinal epithelium, and Langerhans cells (Liu et al., 2021), suggesting involvement in communication between the gut and the fatty tissues, liver, eyes, and skin. It is implicated in pathological states such as intestinal inflammation and cancers, making it a possible therapeutic target in several diseases (Liu et al., 2021).

#### *Nuclear receptors*

The major nuclear receptors in the gut are the AHR, the farnesoid X receptor (FXR), and the pregnane X receptor (PXR).

#### *Aryl hydrocarbon receptor*

This receptor is a transcription factor with a helix-loop-helix motif and senses compounds bearing an aromatic ring, such as indole/tryptophan compounds, polyphenols, flavonoids, and synthetic pollutants like dioxins and polycyclic aromatic hydrocarbons. It controls immunity at the gut barrier via the differentiation and inflammatory responses of innate and adaptive immune cells (Offermanns, 2017; Peters et al., 2019). GM tryptophan catabolism produces AHR ligands, such as indole-3-aldehyde, which stimulate intestinal immunity against *Candida albicans* colonisation via IL-22 (Li et al., 2021). Tryptophan metabolites also communicate bi-directionally between the GM and the brain (gut-brain axis) via the AHR (Shinde and McGaha, 2018). The natural dye indigo binds the AHR and induces the production of interleukins IL-10 and IL-22, which confers protection against high-fat diet (HFD)-induced insulin resistance and fatty liver disease in mice (Tripathi and Lee, 2020). This was linked to specific increases in *Lactobacillus* cell counts and the elicitation of IL-22 secretion in the gut (Zelante et al., 2013). Intestinal inflammation can be modulated by AHR ligands such as oxazoles (Ma et al., 2020) and 6-formylindolo (3,2-b) carbazole (Ficz) (Lin et al., 2019).

#### *Farnesoid X receptor*

FXR is activated by BAs and is involved in lipid and glucose metabolism as well as energy homeostasis through the enterohepatic route (Iyer et al., 2018; Lamas et al., 2020). The antioxidant compound tempol leads to the accumulation of tauro- $\beta$ -muricholic acid (T- $\beta$ -MCA) in mice by blocking BSH enzymes in

the Lactobacilli; T- $\beta$ -MCA inhibits FXR signalling, consequently reducing obesity (Li and Chiang, 2014). Glycine- $\beta$ -MCA prevents obesity, insulin resistance, and fatty liver disease in mice by decreasing the Firmicutes to Bacteroidetes ratio, leading to reduced SCFA levels (Li et al., 2013). BAs conjugated to the amino acids phenylalanine, tyrosine, and leucine are FXR agonists and are elevated in cystic fibrosis and IBD (Zhang et al., n.d.).

The BA derivative obeticholic acid (OCA) can reshape the small intestine microbiome in humans and mice via the FXR receptor (Quinn et al., 2020). These studies demonstrated the links between the GM, FXR and metabolic diseases and showed that FXR agonists could be promising anti-obesity leads via microbiome remodelling. In addition, OCA could also reduce the severity of *C. difficile* infection in mice fed an HFD by an FXR-mediated drop in primary BA levels, which decreases *C. difficile* spore germination (Friedman et al., 2018). Owing to the communication between the GM and the brain (the gut-brain axis), OCA can influence the GM-triggered microglial accumulation in the brain and ameliorate the anxiety associated with metabolic disease of treated mice (Jose et al., 2021). Small-molecule manipulation of the GM therefore enables the modulation of distant organs via the gut-brain, the gut-liver, the gut-heart, and the gut-lung axes.

#### *Pregnane X receptor*

PXR is implicated in the metabolism of xenobiotic compounds and is expressed in the vascular endothelium lining the blood vessels and is in direct contact with the serum (Wu et al., 2021). It is involved in innate immunity via the inflammasome and protection of the endothelia from oxidative damage (Wang et al., 2014). The natural product tanshinone IIA protects the endothelial cells from ROS damage via PXR activation (Zhu et al., 2017), while the GM metabolite indole-3-propionate (IPA) regulates PXR-dependent vasodilation (Pulakazhi Venu et al., 2019). Using IPA as a scaffold, Dvořák et al. synthesised a series of indole derivatives that were the first ever non-cytotoxic PXR agonists that reduced inflammation in mice (Dvořák et al., 2020), suggesting that GM metabolite mimicry might be a viable strategy to discover novel drugs with good efficacy and low toxicity.

#### ***Peroxisome proliferator-activated receptors***

Peroxisome proliferator-activated receptors (PPARs) are found throughout the gut tissue and have roles in fatty acid sensing, metabolism, and modulation of immunity; PPAR $\alpha$  is crucial for fatty acid and branched chain amino acid catabolism in the mitochondria and peroxisomes (Grabacka et al., 2021), while PPAR $\gamma$  is important in innate immunity (Croasdell et al., 2015). Double agonists of both these receptors have been successful in animal models of *Citrobacter rodentium* and DSS-induced colitis of reducing tissue damage and bacterial loads leading to infection clearance and resolved inflammation compared with agonists of each receptor separately (Katkari et al., 2022). PPAR $\alpha$  and  $\gamma$  activation has been reported for keto- and hydroxy-octadecanoic acid species, which were produced by *Lactiplantibacillus plantarum* (Goto et al., 2015). Oleoylethanolamide, an endogenous PPAR ligand, can be administered exogenously in mice to shift the microbiota in the colon to a higher Bacteroidetes/Firmicutes ratio, with corresponding increases in *Bacteroides*, *Prevotella*, and *Parabacteroides* and decreases in *Bacillus* and *Lactobacillus* strains (Di Paola et al., 2018). The GM has also been modulated by synthetic agonists, such as fenofibrate, which led to increased SCFA levels in serum and tissues in mice fed HFDs (Wang et al., 2022). Dysbiosis induced by either high-fructose diets or HFD in mice could be remediated by the PPAR agonist Wy-16434, whereby the Bacteroidetes/Firmicutes ratio increased (reduced Proteobacteria and increased Actinobacteria) (Silva-Veiga et al., 2020; Silva-Veiga et al., 2022).

#### **Future directions and conclusions**

As outlined in this article, approaches such as the inhibition of specific GM metabolism, the use of COINS, the prophylactic use of small-molecule determinants of CR, and GM metabolite mimicry could



emerge as therapeutic avenues in GM modulation and precision medicine. Outside the coverage of this article, developments in canonical amino acid modification, biorthogonal chemistry, non-canonical amino acids, ribosome engineering, mass spectrometry, natural product databases, and machine learning have increased the scope of chemical and chemical information-based tools to interrogate GM-related metabolism and discover GM-related natural products. The emergence of chemical and informatics technologies alongside advances in deep sequencing (Liu et al., 2022a), improvement in technologies to cultivate “uncultivable microbes” (Liu et al., 2022b), and isolate GM-specific microbes via “culturomics” (Diakite et al., 2020) make it an exciting time to be a chemical biologist interested in GM research, with expanding opportunities for chemistry-based discovery and interventions to benefit human health.

**Author contribution.** Conceptualisation, A.P.; Writing (original draft), A.P., P.K.M., K.K.; Writing (review and editing – lead), A.P., P.K.M.; Writing (review and editing – supporting), B.S.K., S.R.S., N.G.P.; Visualisation – B.S.K.

**Disclosure statement.** The author Dr. Pavan Kumar Mantravadi is employee of Cytokinetics, South San Francisco, CA, USA. However, his company did not contribute to this written material. The remaining authors declare that the research/writing was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest. The paper was initiated by Dr. Anutthaman Parthasarathy and Dr. Ganesh Pandian Namasivayam, while the former was on an academic visit to Japan.

## References

- Abt MC, Buffie CG, Sušac B, Becattini S, Carter RA, Leiner I, Keith JW, Artis D, Osborne LC and Pamer EG (2016) TLR-7 activation enhances IL-22-mediated colonization resistance against vancomycin-resistant enterococcus. *Science Translational Medicine* 8(327), 327ra325. <https://doi.org/10.1126/scitranslmed.aad6663>.
- Adhikari AA, Seegar TCM, Ficarro SB, McCurry MD, Ramachandran D, Yao L, Chaudhari SN, Ndousse-Fetter S, Banks AS, Marto JA, et al. (2020) Development of a covalent inhibitor of gut bacterial bile salt hydrolases. *Nature Chemical Biology* 16(3), 318–326. <https://doi.org/10.1038/s41589-020-0467-3>.
- Almeida A, Mitchell AL, Boland M, Forster SC, Gloor GB, Tarkowska A, Lawley TD and Finn RD (2019) A new genomic blueprint of the human gut microbiota. *Nature* 568(7753), 499–504. <https://doi.org/10.1038/s41586-019-0965-1>.
- Barrea L, Di Somma C, Muscogiuri G, Tarantino G, Tenore GC, Orio F, ... Savastano S (2017) Nutrition, inflammation and liver-spleen axis. *Critical Reviews in Food Science and Nutrition* 58(18), 3141–3158. <https://doi.org/10.1080/10408398.2017.1353479>.
- Baumgärtner F, Seitz L, Sprenger GA and Albermann C (2013) Construction of *Escherichia coli* strains with chromosomally integrated expression cassettes for the synthesis of 2'-fucosyllactose. *Microbial Cell Factories* 12, 40. <https://doi.org/10.1186/1475-2859-12-40>.
- Becattini S, Taur Y and Pamer EG (2016) Antibiotic-induced changes in the intestinal microbiota and disease. *Trends in Molecular Medicine* 22(6), 458–478. <https://doi.org/10.1016/j.molmed.2016.04.003>.
- Bédard F and Biron E (2018) Recent progress in the chemical synthesis of class II and S-glycosylated bacteriocins. *Frontiers in Microbiology* 9, 1048. <https://doi.org/10.3389/fmicb.2018.01048>.
- Berger AB, Vitorino PM and Bogyo M (2004) Activity-based protein profiling: applications to biomarker discovery, in vivo imaging and drug discovery. *American Journal of Pharmacogenomics* 4(6), 371–381. <https://doi.org/10.2165/00129785-200404060-00004>.
- Brown AJ, Goldsworthy SM, Barnes AA, Eilert MM, Tcheang L, Daniels D, Muir AI, Wigglesworth MJ, Kinghorn I, Fraser NJ, et al. (2003) The Orphan G protein-coupled receptors GPR41 and GPR43 are activated by propionate and other short chain carboxylic acids. *Journal of Biological Chemistry* 278(13), 11312–11319. <https://doi.org/10.1074/jbc.M211609200>.
- Cabezón E, de la Cruz F and Arechaga I (2017) Conjugation inhibitors and their potential use to prevent dissemination of antibiotic resistance genes in bacteria. *Frontiers in Microbiology* 8, 2329. <https://doi.org/10.3389/fmicb.2017.02329>.
- Cabezón E, Ripoll-Rozada J, Peña A, de la Cruz F and Arechaga I (2015) Towards an integrated model of bacterial conjugation. *FEMS Microbiology Reviews* 39(1), 81–95. <https://doi.org/10.1111/1574-6976.12085>.
- Chen C, Zhang Y, Xue M, Liu X-w, Li Y, Chen X, Wang PG, Wang F and Cao H (2015) Sequential one-pot multienzyme (OPME) synthesis of lacto-N-neotetraose and its sialyl and fucosyl derivatives. *Chemical Communications* 51(36), 7689–7692. <https://doi.org/10.1039/C5CC01330E>.
- Chen H, Nwe PK, Yang Y, Rosen CE, Bielecka AA, Kuchroo M, Cline GW, Kruse AC, Ring AM, Crawford JM, et al. (2019) A forward chemical genetic screen reveals gut microbiota metabolites that modulate host physiology. *Cell* 177(5), 1217–1231. e1218. <https://doi.org/10.1016/j.cell.2019.03.036>.
- Chen PB, Black AS, Sobel AL, Zhao Y, Mukherjee P, Molparia B, Moore NE, Aleman Muench GR, Wu J, Chen W, et al. (2020) Directed remodeling of the mouse gut microbiome inhibits the development of atherosclerosis. *Nature Biotechnology* 38(11), 1288–1297. <https://doi.org/10.1038/s41587-020-0549-5>.

- Cohen LJ, Kang HS, Chu J, Huang YH, Gordon EA, Reddy BV, Ternei MA, Craig JW and Brady SF (2015) Functional metagenomic discovery of bacterial effectors in the human microbiome and isolation of commendamide, a GPCR G2A/132 agonist. *Proceedings of the National Academy of Sciences of the United States of America* **112**(35), E4825–E4834. <https://doi.org/10.1073/pnas.1508737112>.
- Colosimo DA, Kohn JA, Luo PM, Piscotta FJ, Han SM, Pickard AJ, Rao A, Cross JR, Cohen LJ and Brady SF (2019) Mapping interactions of microbial metabolites with human g-protein-coupled receptors. *Cell Host Microbe* **26**(2), 273–282.e277. <https://doi.org/10.1016/j.chom.2019.07.002>.
- Correia MSP, Jain A, Alotaibi W, Young Tie Yang P, Rodriguez-Mateos A and Globisch D (2020) Comparative dietary sulfated metabolome analysis reveals unknown metabolic interactions of the gut microbiome and the human host. *Free Radical Biology and Medicine* **160**, 745–754. <https://doi.org/10.1016/j.freeradbiomed.2020.09.006>.
- Coyne MJ, Zitomersky NL, McGuire AM, Earl AM and Comstock LE (2014) Evidence of extensive DNA transfer between bacteroidales species within the human gut. *mBio* **5**(3), e01305–01314. <https://doi.org/10.1128/mBio.01305-14>.
- Croasdell A, Duffney PF, Kim N, Lacy SH, Sime PJ and Phipps RP (2015) PPAR $\gamma$  and the innate immune system mediate the resolution of inflammation. *PPAR Res* **2015**, 549691. <https://doi.org/10.1155/2015/549691>.
- Cully M (2019) Microbiome therapeutics go small molecule. *Nature Reviews Drug Discovery* **18**(8), 569–572. <https://doi.org/10.1038/d41573-019-00122-8>.
- Das S, Angsantikul P, Le C, Bao D., Miyamoto Y, Gao W, Zhang L and Eckmann L (2018) Neutralization of cholera toxin with nanoparticle decoys for treatment of cholera. *PLOS Neglected Tropical Diseases* **12**(2), e0006266. <https://doi.org/10.1371/journal.pntd.0006266>.
- Devlin AS and Fischbach MA (2015) A biosynthetic pathway for a prominent class of microbiota-derived bile acids. *Nature Chemical Biology* **11**(9), 685–690. <https://doi.org/10.1038/nchembio.1864>.
- Di Paola M, Bonechi E, Provensi G, Costa A, Clarke G, Ballerini C, De Filippo C and Passani MB (2018) Oleoylethanolamide treatment affects gut microbiota composition and the expression of intestinal cytokines in Peyer's patches of mice. *Scientific Reports* **8**(1), 14881. <https://doi.org/10.1038/s41598-018-32925-x>.
- Diakité A, Dubourg G, Dione N, Afouda P, Bellali S, Ngom II, Valles C, Tall MI, Lagier J-C and Raoult D (2020) Optimization and standardization of the culturomics technique for human microbiome exploration. *Scientific Reports* **10**(1), 9674. <https://doi.org/10.1038/s41598-020-66738-8>.
- Ducarmon QR, Zwitterink RD, Hornung BVH, van Schaik W, Young VB and Kuijper EJ (2019) Gut microbiota and colonization resistance against bacterial enteric infection. *Microbiology and Molecular Biology Reviews* **83**(3), e0007–19. <https://doi.org/10.1128/MMBR.00007-19>.
- Dvořák Z, Kopp F, Costello CM, Kemp JS, Li H, Vrzalová A, Štěpánková M, Bartoňková I, Jiskrová E, Pouliková K, *et al.* (2020) Targeting the pregnane X receptor using microbial metabolite mimicry. *EMBO Molecular Medicine* **12**(4), e11621. <https://doi.org/10.15252/emmm.201911621>.
- Enam F and Mansell TJ (2019) Prebiotics: tools to manipulate the gut microbiome and metabolome. *Journal of Industrial Microbiology and Biotechnology* **46**(9–10), 1445–1459. <https://doi.org/10.1007/s10295-019-02203-4>.
- Fair RJ, Hahm HS and Seeberger PH (2015) Combination of automated solid-phase and enzymatic oligosaccharide synthesis provides access to  $\alpha$ (2,3)-sialylated glycans. *Chemical Communications* **51**(28), 6183–6185. <https://doi.org/10.1039/C5CC01368B>.
- Fernandez-Lopez R, Machón C, Longshaw CM, Martin S, Molin S, Zechner EL, Espinosa M, Lanka E and de la Cruz F (2005) Unsaturated fatty acids are inhibitors of bacterial conjugation. *Microbiology (Reading)* **151**(Pt 11), 3517–3526. <https://doi.org/10.1099/mic.0.28216-0>.
- Forster ER, Yang X, Tai AK, Hang HC and Shen A (2022) Identification of a bile acid-binding transcription factor in *Clostridioides difficile* using chemical proteomics. *ACS Chemical Biology* **17**(11), 3086–3099. <https://doi.org/10.1021/acscchembio.2c00463>.
- Friedman ES, Li Y, Shen TD, Jiang J, Chau L, Adorini L, Babakhani F, Edwards J, Shapiro D, Zhao C, *et al.* (2018) FXR-dependent modulation of the human small intestinal microbiome by the bile acid derivative obeticholic acid. *Gastroenterology* **155**(6), 1741–1752.e1745. <https://doi.org/10.1053/j.gastro.2018.08.022>.
- Garg N, Conway LP, Ballet C, Correia MSP, Olsson FKS, Vujasinovic M, Löhr JM and Globisch D (2018) Chemoselective probe containing a unique bioorthogonal cleavage site for investigation of gut microbiota metabolism. *Angewandte Chemie International Edition* **57**(42), 138005–13809. <https://doi.org/10.1002/anie.201804828>.
- Getino M, Fernández-López R, Palencia-Gándara C, Campos-Gómez J, Sánchez-López JM, Martínez M, Fernández A and de la Cruz F (2016) Tanzawaic acids, a chemically novel set of bacterial conjugation inhibitors. *PLoS One* **11**(1), e0148098. <https://doi.org/10.1371/journal.pone.0148098>.
- Getino M, Sanabria-Ríos DJ, Fernández-López R, Campos-Gómez J, Sánchez-López JM, Fernández A, Carballeira NM and de la Cruz F (2015) Synthetic fatty acids prevent plasmid-mediated horizontal gene transfer. *mBio* **6**(5), e01032–01015. <https://doi.org/10.1128/mBio.01032-15>.
- Geva-Zatorsky N, Alvarez D, Hudak JE, Reading NC, Erturk-Hasdemir D, Dasgupta S, von Andrian UH and Kasper DL (2015) In vivo imaging and tracking of host-microbiota interactions via metabolic labeling of gut anaerobic bacteria. *Nature Medicine* **21**(9), 1091–1100. <https://doi.org/10.1038/nm.3929>.

- Gibson GR, Probert HM, Loo JV, Rastall RA and Roberfroid MB (2004) Dietary modulation of the human colonic microbiota: updating the concept of prebiotics. *Nutrition Research Reviews* 17(2), 259–275. <https://doi.org/10.1079/NRR200479>.
- Gil-Pichardo A, Sánchez-Ruiz A and Colmenarejo G (2023) Analysis of metabolites in human gut: illuminating the design of gut-targeted drugs. *Journal of Cheminformatics* 15, 96. <https://doi.org/10.1186/s13321-023-00768-y>.
- Goto T, Kim YI, Furuzono T, Takahashi N, Yamakuni K, Yang HE, Li Y, Ohue R, Nomura W, and Sugawara T, *et al.* (2015) 10-oxo-12(Z)-octadecenoic acid, a linoleic acid metabolite produced by gut lactic acid bacteria, potentially activates PPAR $\gamma$  and stimulates adipogenesis. *Biochemical and Biophysical Research Communications* 459(4), 597–603. <https://doi.org/10.1016/j.bbrc.2015.02.154>.
- Grabacka M, Pierzchalska M, Płonka PM and Pierzchalski P (2021) The role of PPAR alpha in the modulation of innate immunity. *International Journal of Molecular Sciences* 22(19), 10545. <https://doi.org/10.3390/ijms221910545>.
- Hatzenpichler R, Scheller S, Tavormina PL, Babin BM, Tirrell DA and Orphan VJ (2014) In situ visualization of newly synthesized proteins in environmental microbes using amino acid tagging and click chemistry. *Environmental Microbiology* 16(8), 2568–2590. <https://doi.org/10.1111/1462-2920.12436>.
- Hua S (2020) Advances in oral drug delivery for regional targeting in the gastrointestinal tract – influence of physiological, pathophysiological and pharmaceutical factors. *Frontiers in Pharmacology* 11, 524. <https://doi.org/10.3389/fphar.2020.00524>.
- Husted AS, Trauelsen M, Rudenko O, Hjorth SA and Schwartz TW (2017) GPCR-mediated signaling of metabolites. *Cell Metabolism* 25(4), 777–796. <https://doi.org/10.1016/j.cmet.2017.03.008>.
- Iyer SS, Gensollen T, Gandhi A, Oh SF, Neves JF, Collin F, Lavin R, Serra C, Glickman J, de Silva PSA, *et al.* (2018) Dietary and microbial oxazoles induce intestinal inflammation by modulating aryl hydrocarbon receptor responses. *Cell* 173(5), 1123–1134.e1111. <https://doi.org/10.1016/j.cell.2018.04.037>.
- Joice R, Yasuda K, Shafquat A, Morgan XC and Huttenhower C (2014) Determining microbial products and identifying molecular targets in the human microbiome. *Cell Metabolism* 20(5), 731–741. <https://doi.org/10.1016/j.cmet.2014.10.003>.
- Jones BV, Begley M, Hill C, Gahan CG and Marchesi JR (2008) Functional and comparative metagenomic analysis of bile salt hydrolase activity in the human gut microbiome. *Proceedings of the National Academy of Sciences of the United States of America* 105(36), 13580–13585. <https://doi.org/10.1073/pnas.0804437105>.
- Jose S, Mukherjee A, Horrigan O, Setchell KDR, Zhang W, Moreno-Fernandez ME, Andersen H, Sharma D, Haslam DB, and Divanovic S, *et al.* (2021) Obeticholic acid ameliorates severity of *Clostridioides difficile* infection in high fat diet-induced obese mice. *Mucosal Immunology* 14(2), 500–510. <https://doi.org/10.1038/s41385-020-00338-7>.
- Joseph AA, Pardo-Vargas A and Seeberger PH (2020) Total synthesis of polysaccharides by automated glycan assembly. *Journal of the American Chemical Society* 142(19), 8561–8564. <https://doi.org/10.1021/jacs.0c00751>.
- Kang JD, Myers CJ, Harris SC, Kakiyama G, Lee IK, Yun BS, Matsuzaki K, Furukawa M, Min HK, Bajaj JS, *et al.* (2019) Bile acid 7 $\alpha$ -dehydroxylating gut bacteria secrete antibiotics that inhibit *Clostridium difficile*: role of secondary bile acids. *Cell Chemical Biology* 26(1), 27–34.e24. <https://doi.org/10.1016/j.chembiol.2018.10.003>.
- Katkar GD, Sayed IM, Anandachar MS, Castillo V, Vidales E, Toobian D, Usmani F, Sawires JR, Leriche G, Yang J, *et al.* (2022) Artificial intelligence-rationalized balanced PPAR $\alpha/\gamma$  dual agonism resets dysregulated macrophage processes in inflammatory bowel disease. *Communications Biology* 5(1), 231. <https://doi.org/10.1038/s42003-022-03168-4>.
- Khodakivskiy PV, Lauber CL, Yevtodiyenko A, Bazhin AA, Bruce S, Ringel-Kulka T, Ringel Y, Bétrisey B, Torres J, Hu J, *et al.* (2021) Noninvasive imaging and quantification of bile salt hydrolase activity: From bacteria to humans. *Science Advances* 7(6), eaaz9857. <https://doi.org/10.1126/sciadv.aaz9857>.
- Kovatcheva-Datchary P, Shoae S, Lee S, Wahlström A, Nookaew I, Hallen A, Perkins R, Nielsen J and Bäckhed F (2019) Simplified intestinal microbiota to study microbe-diet-host interactions in a mouse model. *Cell Reports* 26(13), 3772–3783. <https://doi.org/10.1016/j.celrep.2019.02.090>.
- Kraehenbuhl JP and Neutra MR (1992) Molecular and cellular basis of immune protection of mucosal surfaces. *Physiological Reviews* 72(4), 853–879. <https://doi.org/10.1152/physrev.1992.72.4.853>.
- Kroeze WK, Sassano MF, Huang XP, Lansu K, McCorvy JD, Giguère PM, Sciaky N and Roth BL (2015) PRESTO-Tango as an open-source resource for interrogation of the druggable human GPCRome. *Nature Structural & Molecular Biology* 22(5), 362–369. <https://doi.org/10.1038/nsmb.3014>.
- Lamas B, Hernandez-Galan L, Galipeau H, J, Constante M, Clarizio A, Jury J, Breyner NM, Caminero A, Rueda G, Hayes CL, *et al.* (2020) Aryl hydrocarbon receptor ligand production by the gut microbiota is decreased in celiac disease leading to intestinal inflammation. *Science Translational Medicine* 12(566), eaba0624. <https://doi.org/10.1126/scitranslmed.aba0624>.
- Lauricica DR, Triantis V, Schoemaker R, Estes MK and Ramani S (2017) Milk oligosaccharides inhibit human rotavirus infectivity in MA104 cells. *Journal of Nutrition* 147(9), 1709–1714. <https://doi.org/10.3945/jn.116.246090>.
- Lee MT, Le HH and Johnson EL (2021) Dietary sphinganine is selectively assimilated by members of the mammalian gut microbiome. *Journal of Lipid Research* 62, 100034. <https://doi.org/10.1194/jlr.RA120000950>.
- Lefebvre P, Cariou B, Lien F, Kuipers F and Staels B (2009) Role of bile acids and bile acid receptors in metabolic regulation. *Physiological Reviews* 89(1), 147–191. <https://doi.org/10.1152/physrev.00010.2008>.

- Li F, Jiang C, Krausz KW, Li Y, Albert I, Hao H, Fabre KM, Mitchell JB, Patterson AD and Gonzalez FJ (2013) Microbiome remodelling leads to inhibition of intestinal farnesoid X receptor signalling and decreased obesity. *Nature Communications* **4**, 2384. <https://doi.org/10.1038/ncomms3384>.
- Li T and Chiang JY (2014) Bile acid signaling in metabolic disease and drug therapy. *Pharmacological Reviews* **66**(4), 948–983. <https://doi.org/10.1124/pr.113.008201>.
- Li W, McArthur JB and Chen X (2019) Strategies for chemoenzymatic synthesis of carbohydrates. *Carbohydrate Research* **472**, 86–97. <https://doi.org/10.1016/j.carres.2018.11.014>.
- Li Z, McCafferty KJ and Judd RL (2021) Role of HCA<sub>2</sub> in regulating intestinal homeostasis and suppressing colon carcinogenesis. *Frontiers in Immunology* **12**, 606384. <https://doi.org/10.3389/fimmu.2021.606384>.
- Lin L, Wu Q, Song J, Du Y, Gao J, Song Y, Wang W and Yang C (2020) Revealing the in vivo growth and division patterns of mouse gut bacteria. *Science Advances* **6**(36). <https://doi.org/10.1126/sciadv.abb2531>.
- Lin S, Zhang Z, Xu H, Li L, Chen S, Li J, Hao Z and Chen PR (2011) Site-specific incorporation of photo-cross-linker and bioorthogonal amino acids into enteric bacterial pathogens. *Journal of the American Chemical Society* **133**(50), 20581–20587. <https://doi.org/10.1021/ja209008w>.
- Lin YH, Luck H, Khan S, Schneeberger PHH, Tsai S, Clemente-Casares X, Lei H, Leu YL, Chan YT, Chen HY, *et al.* (2019) Aryl hydrocarbon receptor agonist indigo protects against obesity-related insulin resistance through modulation of intestinal and metabolic tissue immunity. *International Journal of Obesity* **43**(12), 2407–2421. <https://doi.org/10.1038/s41366-019-0340-1>.
- Liu J, Xu Q, Zhang J, Zhou X, Lyu F, Zhao P and Ding Y (2015) Preparation, composition analysis and antioxidant activities of konjac oligo-glucomannan. *Carbohydrate Polymers* **130**, 398–404. <https://doi.org/10.1016/j.carbpol.2015.05.025>.
- Liu P, Hu S, He Z, Feng C, Dong G, An S, Liu R, Xu F, Chen Y and Ying X (2022a) Towards strain-level complexity: sequencing depth required for comprehensive single-nucleotide polymorphism analysis of the human gut microbiome. *Frontiers in Microbiology* **13**, 828254. <https://doi.org/10.3389/fmicb.2022.828254>.
- Liu Q, Tian X, Maruyama D, Arjomandi M and Prakash A (2021) Lung immune tone via gut-lung axis: gut-derived LPS and short-chain fatty acids' immunometabolic regulation of lung IL-1 $\beta$ , FFAR2, and FFAR3 expression. *American Journal of Physiology Lung Cellular and Molecular Physiology* **321**(1), L65–L78. <https://doi.org/10.1152/ajplung.00421.2020>.
- Liu S, Moon CD, Zheng N, Huws S, Zhao S and Wang J (2022b) Opportunities and challenges of using metagenomic data to bring uncultured microbes into cultivation. *Microbiome* **10**(1), 76. <https://doi.org/10.1186/s40168-022-01272-5>.
- Ltd., I. U. (2018) *Microbiome Modulator Drugs – The New Generation of Therapeutic*. <https://www.scribd.com/document/513306656/Pharmaprojects-Microbiome-Whitepaper>.
- Ma N, He T, Johnston LJ and Ma X (2020) Host-microbiome interactions: the aryl hydrocarbon receptor as a critical node in tryptophan metabolites to brain signaling. *Gut Microbes* **11**(5), 1203–1219. <https://doi.org/10.1080/19490976.2020.1758008>.
- Malwe AS, Srivastava GN and Sharma VK (2023) GutBug: A tool for prediction of human gut bacteria mediated biotransformation of biotic and xenobiotic molecules using machine learning. *Journal of Molecular Biology* **435**(14), 168056. <https://doi.org/10.1016/j.jmb.2023.168056>.
- Mayers MD, Moon C, Stupp GS, Su AI and Wolan DW (2017) Quantitative metaproteomics and activity-based probe enrichment reveals significant alterations in protein expression from a mouse model of inflammatory bowel disease. *Journal of Proteome Research* **16**(2), 1014–1026. <https://doi.org/10.1021/acs.jproteome.6b00938>.
- Munck C, Sheth RU, Freedberg DE and Wang HH (2020) Recording mobile DNA in the gut microbiota using an Escherichia coli CRISPR-Cas spacer acquisition platform. *Nature Communications* **11**(1), 95. <https://doi.org/10.1038/s41467-019-14012-5>.
- Muroski JM, Fu JY, Nguyen HH, Wofford NQ, Mouttaki H, James KL, McInerney MJ, Gunsalus RP, Loo JA and Ogorzalek Loo RR (2022) The acyl-proteome of syntrophus aciditrophicus reveals metabolic relationships in benzoate degradation. *Molecular & Cellular Proteomics* **21**(4), 100215. <https://doi.org/10.1016/j.mcpro.2022.100215>.
- Ndeh D, Rogowski A, Cartmell A, Luis AS, Baslé A, Gray J, Venditto I, Briggs J, Zhang X, Labourel A, *et al.* (2017) Complex pectin metabolism by gut bacteria reveals novel catalytic functions. *Nature* **544**(7648), 65–70. <https://doi.org/10.1038/nature21725>.
- Negatu DA, Gengenbacher M, Dartois V and Dick T (2020) Indole propionic acid, an unusual antibiotic produced by the gut microbiota, with anti-inflammatory and antioxidant properties. *Frontiers in Microbiology* **11**, 575586. <https://doi.org/10.3389/fmicb.2020.575586>.
- Negatu DA, Liu JJJ, Zimmerman M, Kaya F, Dartois V, Aldrich CC, Gengenbacher M and Dick T (2018) Whole-cell screen of fragment library identifies gut microbiota metabolite indole propionic acid as antitubercular. *Antimicrobial Agents and Chemotherapy* **62**(3), e01571–17. <https://doi.org/10.1128/aac.01571-17>.
- Newman MA, Petri RM, Grüll D, Zebeli Q and Metzler-Zebeli BU (2018) Transglycosylated starch modulates the gut microbiome and expression of genes related to lipid synthesis in liver and adipose tissue of pigs. *Frontiers In Microbiology* **9**, 224. <https://doi.org/10.3389/fmicb.2018.00224>.
- Nie Q, Luo X, Wang K, Ding Y, Jia S, Zhao Q, Li M *et al.* (2024) *Cell* **187**(11), 2717–2734.e33. <https://doi.org/10.1016/j.cell.2024.03.034>.



- Nilsson NE, Kotarsky K, Owman C and Olde B (2003) Identification of a free fatty acid receptor, FFA2R, expressed on leukocytes and activated by short-chain fatty acids. *Biochemical and Biophysical Research Communications* **303**(4), 1047–1052. [https://doi.org/10.1016/S0006-291X\(03\)00488-1](https://doi.org/10.1016/S0006-291X(03)00488-1).
- Offermanns S (2017) Hydroxy-carboxylic acid receptor actions in metabolism. *Trends in Endocrinology & Metabolism* **28**(3), 227–236. <https://doi.org/10.1016/j.tem.2016.11.007>.
- Orman M, Bodea S, Funk MA, Campo AM, Bollenbach M, Drennan CL and Balskus EP (2019) Structure-guided identification of a small molecule that inhibits anaerobic choline metabolism by human gut bacteria. *Journal of the American Chemical Society* **141**(1), 33–37. <https://doi.org/10.1021/jacs.8b04883>.
- Oyedemi BO, Shinde V, Shinde K, Kakalou D, Stapleton PD and Gibbons S (2016) Novel R-plasmid conjugal transfer inhibitory and antibacterial activities of phenolic compounds from *Mallotus philippensis* (Lam.) Mull. Arg. *Journal of Global Antimicrobial Resistance* **5**, 15–21. <https://doi.org/10.1016/j.jgar.2016.01.011>.
- Palencia-Gándara C, Getino M, Moyano G, Redondo S, Fernández-López R, González-Zorn B and de la Cruz F (2021) Conjugation inhibitors effectively prevent plasmid transmission in natural environments. *mBio* **12**(4), e0127721. <https://doi.org/10.1128/mBio.01277-21>.
- Parasar B, Zhou H, Xiao X, Shi Q, Brito IL and Chang PV (2019) Chemoproteomic profiling of gut microbiota-associated bile salt hydrolase activity. *ACS Central Science* **5**(5), 867–873. <https://doi.org/10.1021/acscentsci.9b00147>.
- Patangia DV, Anthony Ryan C, Dempsey E, Paul Ross R and Stanton C (2022) Impact of antibiotics on the human microbiome and consequences for host health. *Microbiologyopen* **11**(1), e1260. <https://doi.org/10.1002/mbo3.1260>.
- Pellock SJ, Creekmore BC, Walton WG, Mehta N, Biernat KA, Cesmat AP, Ariyaratna Y, Dunn ZD, Li B, Jin J, et al. (2018) Gut microbial  $\beta$ -glucuronidase inhibition via catalytic cycle interception. *ACS Central Science* **4**(7), 868–879. <https://doi.org/10.1021/acscentsci.8b00239>.
- Peters A, Krumbholz P, Jäger E, Heintz-Buschart A, Çakir MV, Rothmund S, Gaudl A, Ceglarek U, Schöneberg T and Stäubert C (2019) Metabolites of lactic acid bacteria present in fermented foods are highly potent agonists of human hydroxycarboxylic acid receptor 3. *PLoS Genetics* **15**(5), e1008145. <https://doi.org/10.1371/journal.pgen.1008145>.
- Poletto P, Pereira GN, Monteiro CRM, Pereira MAF, Bordignon SE and de Oliveira D (2020) Xylooligosaccharides: Transforming the lignocellulosic biomasses into valuable 5-carbon sugar prebiotics. *Process Biochemistry* **91**, 352–363. <https://doi.org/10.1016/j.procbio.2020.01.005>.
- Priyadarshini M, Lednovich K, Xu K, Gough S, Wicksteed B and Layden BT (2021) FFAR from the gut microbiome crowd: SCFA receptors in T1D pathology. *Metabolites* **11**(5), 302. <https://doi.org/10.3390/metabo11050302>.
- Priyadarshini M, Navarro G and Layden BT (2018) Gut microbiota: FFAR reaching effects on islets. *Endocrinology* **159**(6), 2495–2505. <https://doi.org/10.1210/en.2018-00296>.
- Pulakazhi Venu VK, Saifeddine M, Mihara K, Tsai YC, Nieves K, Alston L, Mani S, McCoy KD, Hollenberg MD and Hirota SA (2019) The pregnane X receptor and its microbiota-derived ligand indole 3-propionic acid regulate endothelium-dependent vasodilation. *American Journal of Physiology Endocrinology and Metabolism* **317**(2), E350–E361. <https://doi.org/10.1152/ajpendo.00572.2018>.
- Putnam EE, Abellon-Ruiz J, Killingier BJ, Rosnow JJ, Wexler AG, Folta-Stogniew E, Wright AT, van den Berg B and Goodman AL (2022) Gut commensal *Bacteroidetes* encode a novel class of vitamin B<sub>12</sub>-Binding Proteins. *mBio* **13**, e02845–21. <https://doi.org/10.1128/mbio.02845-21>.
- Quinn RA, Melnik AV, Vrbanc A, Fu T, Patras KA, Christy MP, Bodai Z, Belda-Ferre P, Tripathi A and Chung LK, et al. (2020) Global chemical effects of the microbiome include new bile-acid conjugations. *Nature* **579**(7797), 123–129. <https://doi.org/10.1038/s41586-020-2047-9>.
- Rackaityte E and Lynch SV (2020) The human microbiome in the 21st century. *Nature Communications* **11**(1), 5256. <https://doi.org/10.1038/s41467-020-18983-8>.
- Randall TD and Mebius RE (2014) The development and function of mucosal lymphoid tissues: a balancing act with microorganisms. *Mucosal Immunology* **7**(3), 455–466. <https://doi.org/10.1038/mi.2014.11>.
- Rea MC, Sit CS, Clayton E, O'Connor PM, Whittall RM, Zheng J, Vederas JC, Ross RP and Hill C. (2010) Thuricin CD, a posttranslationally modified bacteriocin with a narrow spectrum of activity against *Clostridium difficile*. *Proceedings of the National Academy of Sciences of the United States of America* **107**(20), 9352–9357. <https://doi.org/10.1073/pnas.0913554107>.
- Ripoll-Rozada J, García-Cazorla Y, Getino M., Machón C, Sanabria-Ríos D, de la Cruz F, Cabezón E and Arechaga I (2016) Type IV traffic ATPase TrwD as molecular target to inhibit bacterial conjugation. *Molecular Microbiology* **100**(5), 912–921. <https://doi.org/10.1111/mmi.13359>.
- Ruiz-Palacios GM, Cervantes LE, Ramos P, Chavez-Munguia B and Newburg DS (2003) *Campylobacter jejuni* binds intestinal H(O) antigen (Fuc alpha 1, 2Gal beta 1, 4GlcNAc), and fucosyloligosaccharides of human milk inhibit its binding and infection. *Journal of Biological Chemistry* **278**(16), 14112–14120. <https://doi.org/10.1074/jbc.M207744200>.
- Santilli AD, Dawson EM, Whitehead KJ and Whitehead DC (2018) Nonmicrobicidal small molecule inhibition of polysaccharide metabolism in human gut microbes: a potential therapeutic avenue. *ACS Chemical Biology* **13**(5), 1165–1172. <https://doi.org/10.1021/acscchembio.8b00309>.
- Savidge T and Sorg JA (2019) Role of bile in infectious disease: the gall of 7 $\alpha$ -dehydroxylating gut bacteria. *Cell Chemical Biology* **26**(1), 1–3. <https://doi.org/10.1016/j.chembiol.2018.12.010>.

- Schroeder BO and Bäckhed F (2016) Signals from the gut microbiota to distant organs in physiology and disease. *Nature Medicine* 22(10), 1079–1089. <https://doi.org/10.1038/nm.4185>.
- Shelton AN, Seth EC, Mok KC, Han AW, Jackson SN, Haft DR and Taga ME (2019) Uneven distribution of cobamide biosynthesis and dependence in bacteria predicted by comparative genomics. *The ISME Journal* 13(3), 789–804. <https://doi.org/10.1038/s41396-018-0304-9>.
- Shinde R and McGaha TL (2018) The aryl hydrocarbon receptor: connecting immunity to the microenvironment. *Trends in Immunology* 39(12), 1005–1020. <https://doi.org/10.1016/j.it.2018.10.010>.
- Shreiner AB, Kao JY and Young VB (2015) The gut microbiome in health and in disease. *Current Opinion in Gastroenterology* 31(1), 69–75. <https://doi.org/10.1097/MOG.0000000000000139>.
- Silva-Veiga FM, Miranda CS, Martins FF, Daleprane JB, Mandarin-de-Lacerda CA and Souza-Mello V (2020) Gut-liver axis modulation in fructose-fed mice: a role for PPAR- $\alpha$  and linagliptin. *Journal of Endocrinology* 247(1), 11–24. <https://doi.org/10.1530/JOE-20-0139>.
- Silva-Veiga FM, Miranda CS, Vasques-Monteiro IML, Souza-Tavares H, Martins FF, Daleprane JB and Souza-Mello V (2022) Peroxisome proliferator-activated receptor- $\alpha$  activation and dipeptidyl peptidase-4 inhibition target dysbiosis to treat fatty liver in obese mice. *World Journal of Gastroenterology* 28(17), 1814–1829. <https://doi.org/10.3748/wjg.v28.i17.1814>.
- Sorbara MT and Pamer EG (2019) Interbacterial mechanisms of colonization resistance and the strategies pathogens use to overcome them. *Mucosal Immunology* 12(1), 1–9. <https://doi.org/10.1038/s41385-018-0053-0>.
- Spaulding CN, Klein RD, Ruer S, Kau AL, Schreiber HL, Cusumano ZT, Dodson KW, Pinkner JS, Fremont DH, Janetka JW, *et al.* (2017) Selective depletion of uropathogenic *E. coli* from the gut by a FimH antagonist. *Nature* 546(7659), 528–532. <https://doi.org/10.1038/nature22972>.
- Spaulding CN, Klein RD, Schreiber HL, Janetka JW and Hultgren SJ (2018) Precision antimicrobial therapeutics: the path of least resistance? *npj Biofilms and Microbiomes* 4(1), 4. <https://doi.org/10.1038/s41522-018-0048-3>.
- Stecher B, Denzler R, Maier L, Bernet F, Sanders MJ, Pickard DJ, Barthel M, Westendorf AM, Krogfelt KA, Walker AW, *et al.* (2012) Gut inflammation can boost horizontal gene transfer between pathogenic and commensal Enterobacteriaceae. *Proceedings of the National Academy of Sciences of the United States of America* 109(4), 1269–1274. <https://doi.org/10.1073/pnas.1113246109>.
- Subramanian S, Huq S, Yatsunenkov T, Haque R, Mahfuz M, Alam MA, Benezra A, DeStefano J, Meier MF, Muegge BD, *et al.* (2014) Persistent gut microbiota immaturity in malnourished Bangladeshi children. *Nature* 510(7505), 417–421. <https://doi.org/10.1038/nature13421>.
- Trikha P and Lee DA (2020) The role of AhR in transcriptional regulation of immune cell development and function. *Biochimica et Biophysica Acta (BBA) – Reviews on Cancer* 1873(1), 188335. <https://doi.org/10.1016/j.bbcan.2019.188335>.
- Turnbaugh PJ, Ley RE, Hamady M, Fraser-Liggett CM, Knight R and Gordon JI (2007) The human microbiome project. *Nature* 449(7164), 804–810. <https://doi.org/10.1038/nature06244>.
- Vohra Y, Vasan M, Venot A and Boons GJ (2008) One-pot synthesis of oligosaccharides by combining reductive openings of benzylidene acetals and glycosylations. *Organic Letters* 10(15), 3247–3250. <https://doi.org/10.1021/ol801076w>.
- Wang B, Yao M, Lv L, Ling Z and Li L (2017) The human microbiota in health and disease. *Engineering* 3(1), 71–82. <https://doi.org/10.1016/j.ENG.2017.01.008>.
- Wang S, Lei T, Zhang K, Zhao W, Fang L, Lai B, Han J, Xiao L and Wang N (2014) Xenobiotic pregnane X receptor (PXR) regulates innate immunity via activation of NLRP3 inflammasome in vascular endothelial cells. *Journal of Biological Chemistry* 289(43), 30075–30081. <https://doi.org/10.1074/jbc.M114.578781>.
- Wang W, Lin L, Du Y, Song Y, Peng X, Chen X and Yang CJ (2019) Assessing the viability of transplanted gut microbiota by sequential tagging with D-amino acid-based metabolic probes. *Nature Communications* 10(1), 1317. <https://doi.org/10.1038/s41467-019-09267-x>.
- Wang X, Yu C, Liu X, Yang J, Feng Y, Wu Y, Xu Y, Zhu Y and Li W (2022) Fenofibrate ameliorated systemic and retinal inflammation and modulated gut microbiota in high-fat diet-induced mice. *Frontiers in Cellular and Infection Microbiology* 12, 839592. <https://doi.org/10.3389/fcimb.2022.839592>.
- Wu L, Han Y, Zheng Z, Zhu S, Chen J, Yao Y, Yue S, Teufel A, Weng H, Li L, *et al.* (2021) Obeticholic acid inhibits anxiety via alleviating gut microbiota-mediated microglia accumulation in the brain of high-fat high-sugar diet mice. *Nutrients* 13(3), 940. <https://doi.org/10.3390/nu13030940>.
- Yang X, Stein KR and Hang HC (2023) Anti-infective bile acids bind and inactivate a Salmonella virulence regulator. *Nature Chemical Biology* 19, 91–100. <https://doi.org/10.1038/s41589-022-01122-3>.
- Zelante T, Iannitti RG, Cunha C, De Luca A, Giovannini G, Pieraccini G, Zecchi R, D’Angelo C, Massi-Benedetti C, Fallarino F, *et al.* (2013) Tryptophan catabolites from microbiota engage aryl hydrocarbon receptor and balance mucosal reactivity via interleukin-22. *Immunity* 39(2), 372–385. <https://doi.org/10.1016/j.immuni.2013.08.003>.
- Zhang L, Xie C, Nichols RG, Chan S H, Jiang C, Hao R, Smith PB, Cai J, Simons MN, Hatzakis E, *et al.* (2016) Farnesoid X receptor signaling shapes the gut microbiota and controls hepatic lipid metabolism. *mSystems* 1(5), e00070–16. <https://doi.org/10.1128/mSystems.00070-16>.
- Zhang ZJ, Wang YC, Yang X and Hang HC (2020) Chemical reporters for exploring microbiology and microbiota mechanisms. *ChemBiochem* 21(1–2), 19–32. <https://doi.org/10.1002/cbic.201900535>.



- Zheng J, Li H, Zhang X, Jiang M, Luo C, Lu Z, Xu Z and Shi J (2018) Prebiotic mannan-oligosaccharides augment the hypoglycemic effects of metformin in correlation with modulating gut microbiota. *Journal of Agricultural and Food Chemistry* **66**(23), 5821–5831. <https://doi.org/10.1021/acs.jafc.8b00829>.
- Zhu H, Chen Z, Ma Z, Tan H, Xiao C, Tang X, Zhang B, Wang Y and Gao Y (2017) Tanshinone IIA protects endothelial cells from H<sub>2</sub>O<sub>2</sub>-induced injuries via PXR activation. *Biomolecules & Therapeutics (Seoul)* **25**(6), 599–608. <https://doi.org/10.4062/biomolther.2016.179>.
- Zhu W, Winter MG, Byndloss MX, Spiga L, Duerkop BA, Hughes ER, Büttner L, de Lima Romão E, Behrendt CL, Lopez CA, *et al.* (2018) Precision editing of the gut microbiota ameliorates colitis. *Nature* **553**(7687), 208–211. <https://doi.org/10.1038/nature25172>.
- Zimmermann M., Zimmermann-Kogadeeva M, Wegmann R and Goodman AL (2019) Separating host and microbiome contributions to drug pharmacokinetics and toxicity. *Science* **363**(6427), eaat9931. <https://doi.org/10.1126/science.aat9931>.

---

**Cite this article:** Mantravadi PK, Kovi BS, Reddy SR, Namasivayam GP, Kalesh K and Parthasarathy A 2025. Probing and manipulating the gut microbiome with chemistry and chemical tools. *Gut Microbiome*, **6**, e6, 1–21. <https://doi.org/10.1017/gmb.2025.4>