

## Research Article

**Cite this article:** Kuzniar ZJ, Harms NE, Ward SM, and Thum RA (2025). Introduction of three cryptic lineages of invasive crested floating heart (*Nymphoides cristata*) in the southeastern United States. *Invasive Plant Sci. Manag.* doi: [10.1017/inp.2024.28](https://doi.org/10.1017/inp.2024.28)

Received: 27 November 2023

Revised: 30 April 2024

Accepted: 12 July 2024

### Associate Editor:

Marie Jasieniuk, University of California, Davis

### Keywords:


Aquatic plant; biological invasions; clonal reproduction; genotyping-by-sequencing

### Corresponding author:

Zachary J. Kuzniar;

Email: [zachary.kuzniar@student.montana.edu](mailto:zachary.kuzniar@student.montana.edu)

# Introduction of three cryptic lineages of invasive crested floating heart (*Nymphoides cristata*) in the southeastern United States

Zachary J. Kuzniar<sup>1</sup> , Nathan E. Harms<sup>2</sup>, Sarah M. Ward<sup>3</sup> and Ryan A. Thum<sup>4</sup>

<sup>1</sup>Graduate Student, Department of Plant Science and Plant Pathology, Montana State University, Bozeman, MT, USA; <sup>2</sup>Research Biologist, U.S. Army Engineer Research and Development Center, Lewisville, TX, USA; <sup>3</sup>Affiliate Professor, Department of Land Resources and Environmental Sciences, Montana State University, Bozeman, MT, USA and <sup>4</sup>Associate Professor, Department of Plant Science and Plant Pathology, Montana State University, Bozeman, MT, USA

## Abstract

Crested floating heart [*Nymphoides cristata* (Roxb.) Kuntze] is an invasive aquatic plant in the southeastern United States. For clonal plants like *N. cristata*, clonal diversity may influence response to control tactics and/or evolutionary potential. However, little is known about the diversity of introduced *N. cristata*. In this study, we used genotyping by sequencing to quantify *N. cristata* diversity in the southeastern United States and determine how that diversity is distributed across the invaded range. Our results show that at least three distinct genetic lineages of *N. cristata* are present in the southeastern United States. Geographic distribution of the lineages varied, with one widespread lineage identified across several states and others only found in a single water body. There is also evidence of extensive asexual reproduction, with invaded water bodies often host to a single genetic lineage. The genetic diversity reported in this study likely results from multiple introductions of *N. cristata* to the southeastern United States and should be considered by managers when assessing control tactics, such as screening for biocontrol agents or herbicide testing. The extent and distribution of genetic diversity should also be considered by researchers studying the potential for invasive spread of *N. cristata* within the United States or hybridization with native *Nymphoides* species.

## Introduction

Genetic variation can influence plant invasions and management (i.e., Barrett 1992; Bossdorf et al. 2005; Sakai et al. 2001; Ward et al. 2008). Additionally, genetic variation can facilitate adaptation to new environments encountered after introductions and during range expansion (e.g., Lee 2002; Prentis et al. 2008). For example, distinct genetic populations or lineages of invasive plants may vary in their response to herbicides (e.g., Chorak and Thum 2020; Kay 1992; Kurniadie et al. 2021; Netherland and Willey 2017; Williams et al. 2020). Similarly, genetic lineages can differ in the effect of biological control agents (e.g., Blossey et al. 2018; Bruckart et al. 2004; Sobhian et al. 2003; Williams et al. 2014).

The amount and distribution of genetic variation in invasive plants will be primarily influenced by the number of introductions; the genetic structure and diversity of source populations; and the extent of admixture, sexual reproduction, and asexual reproduction in the introduced range. At one extreme, introduction of a single genet followed by exclusively asexual reproduction will result in essentially no genetic diversity in the introduced range (e.g., Hollingsworth and Bailey 2000; Le Roux et al. 2007; Loomis and Fishman 2009; Zhang et al. 2010). At the other extreme, multiple introductions from genetically distinct source populations, followed by admixture and sexual reproduction, can generate greater and novel genetic diversity in introduced populations (Facon et al. 2008; Kolbe et al. 2004; Lavergne and Molofsky 2007). Population genetic and genomic descriptions of invasive species can help determine the number of distinct lineages present in the introduced range and provide insight into the extent or potential for admixture of distinct lineages, which in turn can inform studies on herbicidal and biological control development.

Crested floating heart [*Nymphoides cristata* (Roxb.) Kuntze] is a floating-leaved aquatic plant native to Asia that has spread across the southeastern United States. The first introduction of *N. cristata* appears to have occurred in south Florida circa 1996, when plants escaped from cultivation for the aquatic garden trade (Burks 2002). Since then, *N. cristata* has been observed in Louisiana (2012), Mississippi (2016), North Carolina (2017), South Carolina (2006), and Texas (2014) and is considered an invasive/noxious species in several of the inhabited states (Thayer and Pfingsten 2024). The scattered distribution of *N. cristata* populations throughout the southeastern United States may reflect multiple introductions (Burks 2002), but no genetic

© The Author(s), 2024. Published by Cambridge University Press on behalf of Weed Science Society of America. This is an Open Access article, distributed under the terms of the Creative Commons Attribution licence (<https://creativecommons.org/licenses/by/4.0/>), which permits unrestricted re-use, distribution and reproduction, provided the original article is properly cited.



### Management Implications

The identification of three distinct *Nymphoides cristata* (crested floating heart) lineages here—and a fourth interspecific hybrid lineage (*Nymphoides cristata* × *Nymphoides aquatica*) reported elsewhere—has implications for the management of this invasive species. Studies of control techniques, spread, and impacts of *N. cristata* should explicitly consider the genetic diversity identified in this study. For example, herbicides are currently the most common option for controlling *N. cristata*, including submersed applications of diquat, endothall, and florypyrauxifen-benzyl; foliar applications of endothall, imazamox, and imazapyr; and foliar combinations of flumioxazin and glyphosate. However, herbicide testing has not explicitly considered whether *N. cristata* lineages differ in their responses. As differences in herbicide response have been found among distinct clonal genotypes of other aquatic plants (e.g., 2,4-D and fluridone in *Myriophyllum* spp.; fluridone in *Hydrilla verticillata*), we posit that herbicide trials for *N. cristata* should include representatives of the four distinct lineages identified thus far. Similarly, *N. cristata* has been identified as a candidate for biological control. However, there are no biocontrol agents currently in operation. The identification of distinct lineages in the United States suggests at least three independent introductions. A genetic survey of *N. cristata* in its native range could help inform the search for biological control agents, especially if introductions can be traced to distinct geographic origins. Further, any biological control agents identified should be tested on the distinct genetic lineages identified in the United States, as biocontrol efficacy can vary at the subspecific level. Finally, it is possible that the distinct lineages could have distinct environmental preferences or tolerances that could be important for predicting their spread across the landscape.

analysis has been conducted to date to test whether introduced *N. cristata* consists of one or more distinct lineages.

*Nymphoides cristata* is capable of both asexual and sexual reproduction, although the prevailing mode of reproduction remains largely unknown. In its native range, *N. cristata* reproduces sexually through a gynodioecious breeding system derived from heterostyly, wherein female plants with reduced, sterile stamens, rely on the bisexual-morph plants for pollen (Nair 1973). In addition, bisexual plants from the native range have shown self-compatibility in an experimental setting (Nair 1973). However, asexual reproduction via vegetative propagation is likely responsible for the majority of biomass (Nair 1973; Sculthorpe 1967), a strategy common in the *Nymphoides* genus (Gettys et al. 2017; Sivarajan and Joseph 1993). Similarly, *N. cristata* is thought to reproduce primarily through vegetative means in the introduced range (fragmentation, daughter plants, tubers, and rhizomes) (Burks 2002; Willey and Langeland 2011). Spread and range expansion of the species is likely facilitated via fragmentation caused by contact with boat motors, wave action, and mechanical harvesting (Burks 2002; Willey et al. 2014). *Nymphoides cristata* is also capable of producing seeds (Burks 2002; Gettys et al. 2017), and has hybridized with a native species, big floating heart [*Nymphoides aquatica* (J.F. Gmel.) Kuntze] (Harms et al. 2021), but the extent of sexual reproduction in U.S. populations remains unknown.

In this study, we conducted a population genomic survey of introduced *N. cristata* in the southern United States to determine how much genetic diversity is in the introduced range. For

example, if the *N. cristata* invasion results from asexual spread of a single genetic clone, then we would expect to find no population genomic variation in our survey (barring somatic mutation and genotyping error). In contrast, identification of multiple distinct genetic lineages would suggest multiple introductions from different sources and the potential for intermediate genotypes if distinct lineages have hybridized.

## Materials and Methods

### Sample Collection

Samples were obtained from a previous study on *N. cristata* (see Harms et al. 2021), using preserved DNA samples and dried plant tissues. Briefly, Harms et al. (2021) collected 1 to 13 plants per water body, depending on the size of the infestation, with small populations minimally sampled (i.e., only a few plants). Within water bodies, plants were sampled 3 to 5 m apart to avoid repeatedly collecting the same plant. The final dataset consisted of 62 samples (i.e., leaves from *N. cristata* plants) from 12 different water bodies throughout the southeast United States, including lakes, ponds, roadside ditches, and canals (Table 1). We acknowledge that our sampling strategy limits our inference regarding diversity within water bodies. However, the main focus of this study was to evaluate evidence for one versus multiple genetic lineages in the United States, and given the likelihood of local clonal reproduction, we prioritized the number of water bodies examined over the number of individuals per water body.

### DNA Sequencing

We used next-generation, genotyping by sequencing (GBS) to generate a single-nucleotide polymorphism (SNP) dataset. Before sequencing, DNA was extracted from plant tissues using the Qiagen DNeasyPlant Kit (Qiagen, Valencia, CA) following the standard plant protocol. A Qubit fluorometer (ThermoFisher Scientific, Waltham, MA) was used to confirm genomic DNA content was high enough for sequencing ( $\geq 60$  ng total gDNA), then all extracts were sent to the University of Minnesota Genomic Center for library assembly and sequencing. The sequencing library was prepared for double digest restriction-site associated DNA sequencing (ddRAD; Peterson et al. 2012) using the restriction enzyme pair *Pst*I and *Msp*I and size-selected for 101-bp fragments using the PippinHT system. The library was sequenced on an Illumina NovaSeq 6000 (Illumina, San Diego, CA) system targeting approximately 2.5M single-end reads per sample.

### Bioinformatics and Filtering

Following sequencing, we processed the raw sequence data using a bioinformatics pipeline to produce an SNP dataset for downstream diversity analyses. First, the reads were demultiplexed by barcode, and adapters were trimmed using *gbstrim*, a custom script designed to preprocess GBS data generated by UMGC (Garbe 2022). The demultiplexed reads were then passed to Stacks v. 2.55 (Catchen et al. 2013), where the *process\_radtags* module removed low-quality reads (i.e., “Phred” score < 25). Next, the core de novo pipeline was executed in Stacks with the minimum stack depth (*m*), mismatch distance between loci within an individual (*M*), and number of mismatches between loci in the catalog (*n*) parameters set at 5, 4, and 4, respectively. These were selected following a parameter optimization procedure similar to that outlined in Paris

**Table 1.** Water bodies where *Nymphoides cristata* were collected for use in this study, including genetic lineage assignments (Lineage ID) and number of individuals (N) collected from each water body.

Water body	Latitude	Longitude	County	State	Lineage ID	N
Lake Fairview	28.6005 °N	81.4128 °W	Orange	FL	CFH-2	1
JW Corbett WMA	26.8579	80.4165	Palm Beach	FL	CFH-2	2
Roadside canal	26.6548	80.1747	Palm Beach	FL	CFH-1, CFH-2	2, 6
Flying Cow ditch	26.6348	80.3001	Palm Beach	FL	CFH-1	6
Business pond	26.0057	80.3018	Broward	FL	CFH-2	2
Flat Lake	30.2817	90.8182	Ascension	LA	CFH-2	13
Private pond	30.4043	91.1576	Avoyelles	LA	CFH-2	1
Lake Marion	33.5372	80.428	Berkeley	SC	CFH-3	7
Caddo Lake	32.7195	94.1198	Harrison	TX	CFH-2	10
Lake Conroe	30.5643	95.6358	Montgomery	TX	CFH-2	3
Houston Arboretum	29.7618	95.4498	Harris	TX	CFH-2	7
Roadside ditch	30.4399	94.7201	Hardin	TX	CFH-2	2

et al. (2017). After catalog creation, SNP identification and genotyping were also performed in Stacks, retaining only one biallelic SNP per RAD locus. The resulting variant-calling files were exported for further filtering and analysis. All bioinformatic work in this study was performed on the Montana State University Tempest computing cluster.

To ensure only high-quality variants were included in downstream analysis, the genetic data were filtered with the R package vCFR v. 1.13.0 (Knaus and Grunwald 2017). To reduce the amount of missing data, we excluded loci that were absent in >25% of the individuals and removed any individuals with >75% missing genotype calls. We also removed loci with unusual read depth (under 10th percentile or over 90th percentile) and set the minimum minor allele frequency to 0.05. After filtering, the final genetic dataset contained 62 *N. cristata* samples genotyped at 2,242 SNPs.

**Data Analysis**

To avoid violating any model assumptions associated with the clonality of *N. cristata*, we used a model-free approach to explore the amount and distribution of genetic diversity. A Euclidean genetic distance matrix was generated in adegenet v. 2.1.7 (Jombart and Ahmed 2011) for use in principal components analysis (PCA). The PCA was completed in the ADE4 package (Dray and Dufour 2007) to visualize genetic variation among samples and determine whether distinct genetic groups of *N. cristata* exist. We also generated a genetic dissimilarity matrix using POPPR v. 2.9.3 (Kamvar et al. 2014) to summarize the actual number of SNP differences between individuals. The distribution of *N. cristata* across the invaded range was mapped with GGPlot2 (Wickham 2016). All data analyses were conducted in R v. 4.2.1 (R Core Team 2021).

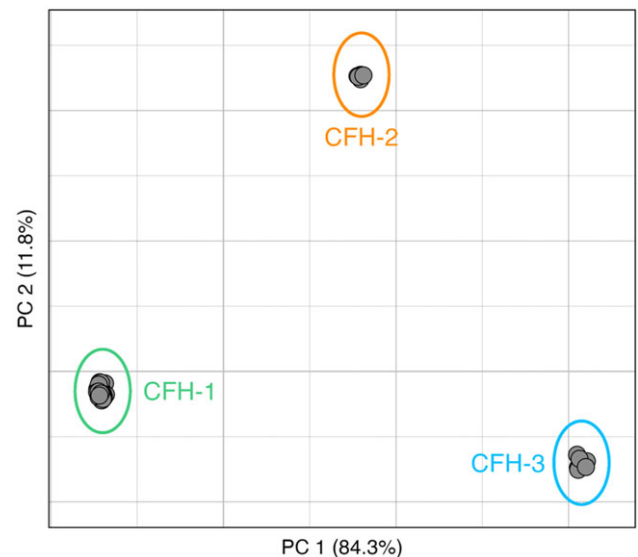
**Results and Discussion**

Our survey of *N. cristata* diversity identified three distinct genetic groups (Figure 1). We refer to those groups as genetic lineages, collections of closely related individuals distinguished by the genomic variants (i.e., SNPs) inherited from a common ancestor. Genetic variation was found both within and between lineages, although the amount of between-lineage variation was far greater than within-lineage variation. Within lineages, individuals averaged approximately 35 SNP differences, while individuals

**Table 2.** Summary of individual-based genetic distance within and between lineages.<sup>a</sup>

	CFH-1	CFH-2	CFH-3
CFH-1	30	—	—
CFH-2	1,514	31	—
CFH-3	1,099	1,028	41

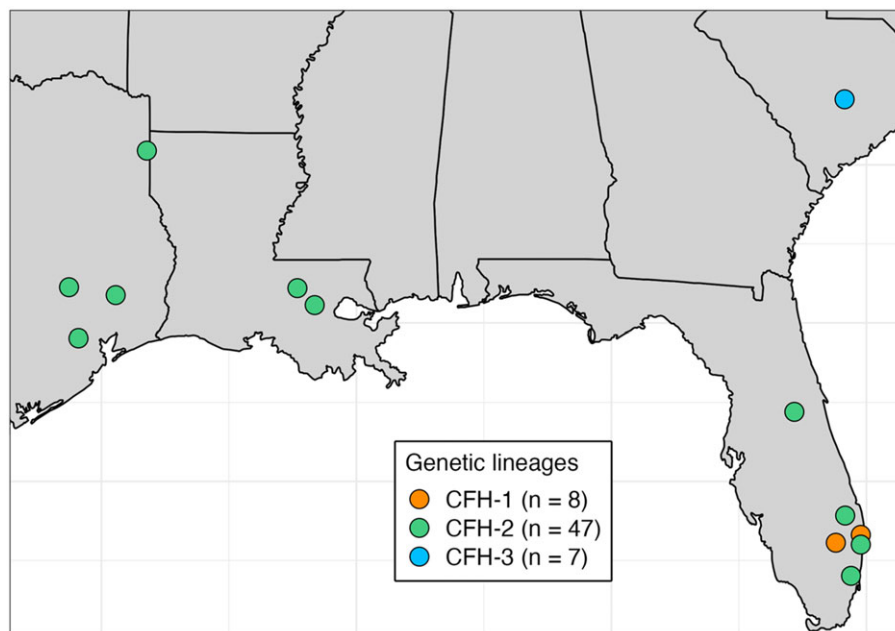
<sup>a</sup>Reported values along the diagonal are the absolute number of single-nucleotide polymorphism (SNP) differences between individuals within a lineage. Between-lineage values represent the average number of SNP differences observed across lineages.



**Figure 1.** Principal component analysis (PCA) of 62 *Nymphoides cristata* samples. PC1 and PC2 account for 96.1% of cumulative variation. Colored ellipses (non-statistical) were added a posteriori to denote putative genetic lineages identified by ordination.

compared across lineages differed by an average of 1,028 to 1,514 differences (Table 2).

The CFH-2 lineage was the most common and widespread; it was found in Florida, Louisiana, and Texas and occurred in 10 of the 12 water bodies sampled overall (Figure 2). In contrast, CFH-3 was only found in one water body (Lake Marion, SC), and CFH-1 was only found in two water bodies, both in southeast Florida (Figure 2).



**Figure 2.** Distribution of *Nymphoides cristata* lineages across the introduced range. Points represent water bodies where samples were collected, and colors denote the genetic lineage(s) present, assigned according to the principal component analysis (PCA) analysis in Figure 1. Note there are two overlapping points representing the roadside canal in south Florida where CFH-1 and CFH-2 co-occurred.

The differences in relative abundance among lineages may be due to introduction dynamics and/or ecological differences among lineages. *Nymphoides cristata* was likely brought to the United States for trade as an aquatic garden ornamental and remains available for purchase from vendors in the industry. Although it is now illegal to possess, import, or distribute *N. cristata* in several southern states (FL, SC, NC, TX), the ornamental industry is a likely candidate for initial introduction(s) and subsequent range expansion. The relative abundance of the CFH-2 lineage may indicate that it was a preferential/popular lineage in the industry or that it was the first to escape cultivation. Indeed, *N. cristata* is clonally propagated for sale in the industry, although it was not possible to include commercially available ornamental samples in this study due to time and funding constraints. In addition, CFH-2 may be more vigorous and widely adapted in the introduced range than others, facilitating its spread across the southern states, while CFH-1 and CFH-3 remained isolated. Further investigation would be necessary to determine whether CFH-2 presents a greater management challenge. We recognize that the number of water bodies sampled and number of individuals sampled within a water body were limited and there may be more diversity in the introduced range than we detected.

Burks (2002) suggested multiple introductions, possibly escaped from ornamental water gardens, based on the scattered distribution of *N. cristata* across southern Florida. Indeed, we identified two distinct lineages in Florida: one that was restricted to southeastern Florida, and a second lineage that is widespread across Florida and the U.S. Gulf Coast (Figure 2). These may represent two independent introductions in Florida followed by range expansion of CFH-2. It is also possible that CFH-2 has been repeatedly introduced across the Gulf Coast states, with its widespread distribution representing numerous independent introductions. Further, we identified a third unique lineage found only in South Carolina (CFH-3), which could represent another introduction.

We cannot rule out with certainty the alternative hypotheses of a single introduction from a genetically variable source or accumulation of new mutations following introduction. However, we would have expected more within-water body lineage diversity if *N. cristata* was introduced from a genetically variable source (although we recognize that within-water body sample sizes were low). Similarly, we find clonal evolution postintroduction unlikely because of the relatively large number of allelic differences (1,028 to 1,514) separating the three distinct lineages combined with the relatively recent introduction (~1996). Plant invasions resulting from multiple introductions are common (Dlugosch and Parker 2008), particularly for ornamental species like *N. cristata*, for which the plant trade increases the likelihood of repeated introductions (Dehnen-Schmutz et al. 2007). Sampling efforts in the native range (i.e., southeastern Asia), along with additional sampling in the United States, could help clarify the number and location(s) of sources of *N. cristata* introduction. In addition, *N. cristata* has been identified as a good candidate for biological control, although natural enemies/potential agents have yet to be described (Harms and Nachtrieb 2019). Identification of source populations in the native range might also yield natural enemies of this plant that could be tested as potential biocontrol agents (Bossdorf et al. 2005; Gaskin et al. 2011).

The introduction of three distinct lineages provides opportunities for genetic admixture and the generation of novel genetic variants in the introduced range (e.g., Facon et al. 2008; Kolbe et al. 2004; Lavergne and Molofsky 2007). Although interspecific hybrids between *N. cristata* and native *N. aquatica* have been identified in the Santee Cooper Reservoir system in South Carolina (Harms et al. 2021), we did not find any evidence for sexual reproduction among the three lineages, as evidenced by the lack of genetically intermediate individuals (Figure 1). In addition, the average pairwise genetic distances between individuals were approximately 97% greater when comparing between lineages versus within lineages. It is possible that these lineages have



sexually reproduced with one another, but that we did not sample any of their offspring. Alternatively, it is possible that the distinct lineages are capable of sexual reproduction but have not yet had sufficient opportunity. The three lineages were largely allopatric, but there was some overlap between two of them in south Florida. Finally, it is possible that the different lineages have some reproductive barriers (e.g., pre- or post-mating, pre- or post-zygotic) that limit sexual reproduction between them. Additional study of reproductive potential among the introduced lineages is warranted.

While we cannot rule out some degree of sexual reproduction within lineages, we hypothesize that the low genetic variation observed within each of the lineages primarily reflects sequencing/genotyping errors and that the *N. cristata* lineages identified here have primarily reproduced asexually throughout the southeastern United States. Although individuals within each lineage were not genetically identical per se, clonal genotypes are not expected to have identical genotypes across thousands of SNPs generated by ddRAD due to sequencing/genotyping error and somatic mutations (da Cunha et al. 2021; Reynes et al. 2021). The interpretation of the low within-lineage variation as clonal reproduction is consistent with previous field observations and reports of reproductive biology of *N. cristata* that hypothesize prolific vegetative propagation and spread in the invaded range (Burks 2002; Harms and Nachtrieb 2019; Willey and Langeland 2011). Inbreeding could also account for the within-lineage variation we observed. *Nymphoides cristata* has been proven to be self-compatible through artificial pollination of bisexual plants in an experimental setting (Nair 1973). However, heterostyly provides a morphological-based incompatibility system in *N. cristata*. Although incomplete, this system is thought to promote reproduction between the female and bisexual plant morphs (i.e., dioecism) in the native range (Nair 1973). Further sampling and detailed description of flower morphology in the introduced range could help decipher the reproductive capabilities of the introduced populations. Finally, we acknowledge that the low sample sizes within water bodies preclude an understanding of the relative extent of sexual versus asexual reproduction in any single population of *N. cristata*.

**Acknowledgments.** The authors would like to thank Greg Chorak, Del Hannay, and Ashley Wolfe for providing thoughtful comments to improve the article. We thank Cassidy Kempf, Michael Coulon, Tim Bister, Jacob Green, Casey Moorer, John Riser, and Carl Bussells for assistance with sample collection.

**Funding statement.** This work was supported by the U.S. Army Engineer Research and Development Center Aquatic Plant Control Research Program Cooperative Agreement W912HZ-18-2-0010, and the Montana Agricultural Experiment Station (Project MONB00249).

**Competing interests.** The authors declare no conflicts of interest.

## References

- Barrett SCH (1992) Genetics of weed invasions. Pages 91–119 in Jain SK, Botsford LW, eds. Applied Population Biology. Dordrecht, Netherlands: Kluwer Academic
- Blossey B, Häfliger P, Tewksbury L, Dávalos A, Casagrande R (2018) Host specificity and risk assessment of *Archanara geminipuncta* and *Archanara neurica*, two potential biocontrol agents for invasive *Phragmites australis* in North America. *Biol Control* 125:98–112
- Bossdorf O, Auge H, Lafuma L, Rogers WE, Siemann E, Prati D (2005) Phenotypic and genetic differentiation between native and introduced plant populations. *Oecologia* 144:1–11
- Bruckart W, Cavin C, Vajna L, Schwarczinger I, Ryan FJ (2004) Differential susceptibility of Russian thistle accessions to *Colletotrichum gloeosporoides*. *Biol Control* 30:306–311
- Burks KC (2002) *Nymphoides cristata* (Roxb.) Kuntze, a recent adventive expanding as a pest in Florida. *Castanea* 67:206–211
- Catchen J, Hohenlohe PA, Bassham S, Amores A, Cresko WA (2013) Stacks: an analysis tool set for population genomics. *Mol Ecol* 22:3124–3140
- Chorak GM, Thum RA (2020) Identification of resistant clones of Eurasian (*Myriophyllum spicatum*) and hybrid (*Myriophyllum spicatum* x *Myriophyllum sibiricum*) watermilfoil to an operational rate of fluridone. *Invasive Plant Sci Manag* 13:247–251
- da Cunha NL, Xue H, Wright SI, Barrett SCH (2021) Genetic variation and clonal diversity in floating aquatic plants: comparative genomic analysis of water hyacinth species in their native range. *Mol Ecol* 31:5307–5325
- Dehnen-Schmutz K, Touza J, Perrings C, Williamson M (2007) A century of the ornamental plant trade and its impact on invasion success. *Divers Distrib* 13:527–534
- Dlugosch KM, Parker IM (2008) Founding events in species invasions: genetic variation, adaptive evolution, and the role of multiple introductions. *Mol Ecol* 17:431–449
- Dray S, Dufour A (2007) The ade4 package: implementing the duality diagram for ecologists. *J Stat Soft* 22:1–20
- Facon B, Pointier JP, Jarne P, Sarda W, David P (2008) High genetic variation in life-history strategies within invasive populations by way of multiple introductions. *Curr Biol* 18:363–367
- Garbe J (2022) gbstrim. <https://bitbucket.org/jgarbe/gbstrim/src/master> Accessed: September 12, 2022
- Gaskin JF, Bon MC, Cock MJW, Cristofaro M, Biase A, Clerck-Floate R, Ellison CA, Hinz HL, Hufbauer RA, Julien MH, Sforza R (2011) Applying molecular-based approaches to classical biological control of weeds. *Biol Control* 58:1–21
- Gettys LA, Della Torre CJ, Thayer KM, Markovich IJ (2017) Asexual reproduction and ramet sprouting of crested floating heart (*Nymphoides cristata*). *J Aquat Plant Manag* 55:83–88
- Harms NE, Nachtrieb JG (2019) Suitability of Introduced *Nymphoides* spp. (*Nymphoides cristata*, *N. peltata*) as Targets for Biological Control in the United States. U.S. Army Corps of Engineers Aquatic Plant Control Research Program Technical Note TN APCRP-BC-42. 18 p
- Harms NE, Thum RA, Gettys LA, Markovich IJ, French A, Simantel L, Richardson R (2021) Hybridization between native and invasive *Nymphoides* species in the United States. *Biol Invasions* 23:3003–3011
- Hollingsworth MI, Bailey J (2000) Evidence for massive clonal growth in the invasive weed, *Fallopia japonica* (Japanese knotweed). *Bot J Linn Soc* 133:463–472
- Jombart T, Ahmed I (2011) adegenet 1.3-1: new tools for the analysis of genome-wide SNP data. *Bioinformatics* 27:3070–3071
- Kamvar ZN, Tabima JF, Grunwald NJ (2014) Poppr: an R package for genetic analysis of populations with clonal, partially clonal, and/or sexual reproduction. *PeerJ* 2:e281
- Kay SH (1992) Response of two alligatorweed biotypes to quinlorac. *J Aquat Plant Manag* 30:35–40
- Knaus BJ, Grunwald NJ (2017) vcfR: a package to manipulate and visualize variant call format data in R. *Mol Ecol Res* 17:44–53
- Kolbe JJ, Glor RE, Rodríguez Schettino L, Lara AC, Larson A, Losos JB (2004) Genetic variation increases during biological invasion by a Cuban lizard. *Nature* 431:177–181
- Kurniadie D, Widiyanto R, Widayat D, Umiyati U, Nasahi C, Kato-Noguchi H (2021) Herbicide-resistant invasive plant species *Ludwigia decurrens* Walter. *Plants* 10:1973
- Lavergne S, Molofsky J (2007) Increased genetic variation and evolutionary potential drive the success of an invasive grass. *Proc Natl Acad Sci USA* 104:3883–3888
- Lee CE (2002) Evolutionary genetics of invasive species. *Trends Ecol Evol* 17:386–391

- Le Roux JJ, Wicczorek AM, Wright MG, Tran CT (2007) Super-genotype: global monoclonality defies the odds of nature. *PLoS ONE* 2:e590
- Loomis ES, Fishman L (2009) A continent-wide clone: population genetic variation of the invasive plant *Hieracium aurantiacum* (orange hawkweed; Asteraceae) in North America. *Int J Plant Sci* 170:759–765
- Nair RV (1973) Heterostyly and breeding mechanism of *Nymphoides cristatum* (Roxb.) O. Kuntze. *J Bombay Nat Hist Soc* 72:677–682
- Netherland MD, Willey L (2017) Mesocosm evaluation of three herbicides on Eurasian watermilfoil (*Myriophyllum spicatum*) and hybrid watermilfoil (*Myriophyllum spicatum* x *Myriophyllum sibiricum*): developing a predictive assay. *J Aquat Plant Manag* 55:39–41
- Paris JR, Stevens JR, Catchen JM (2017) Lost in parameter space: a road map for *Stacks*. *Methods Ecol Evol* 8:1360–1373
- Peterson BK, Weber JN, Kay EH, Fisher HS, Hoekstra HE (2012) Double digest RADseq: an inexpensive method for de novo SNP discovery and genotyping in model and non-model species. *PLoS ONE* 7:e37135
- Prentis P, Wilson JRU, Dormontt EE, Richardson DM, Lowe AJ (2008) Adaptive evolution in invasive species. *Trends Plant Sci* 13:288–294
- R Core Team (2021) R: A Language and Environment for Statistical Computing. Vienna, Austria: R Foundation for Statistical Computing
- Reynes L, Thibaut T, Mauger S, Blanfune A, Holon F, Cruaud C, Coulloux A, Valero M, Aurelle D (2021) Genomic signatures of clonality in deep water kelp *Laminaria rodriguezii*. *Mol Ecol* 30:1806–1822
- Sakai AK, Allendorf FW, Holt JS, Lodge DM, Molofsky J, With KA, Baughman S, Cabin RJ, Cohen JE, Ellstrand NC, McCauley DE, O'Neil P, Parker IM, Thompson JN, Weller SG (2001) The population biology of invasive species. *Annu Rev Ecol Syst* 32:305–332
- Sculthorpe CD (1967) *The Biology of Aquatic Vascular Plants*. 1st ed. London: Edward Arnold. 610p
- Sivarajan VV, Joseph KT (1993) The genus *Nymphoides* Seguirer (*Menyanthaceae*) in India. *Aquat Bot* 45:145–170
- Sobhian R, Ryan FJ, Khamraev A, Pitcairn MJ, Bell DE (2003) DNA phenotyping to find a natural enemy in Uzbekistan for California biotypes of *Salsola tragus* L. *Biol Control* 28:222–228
- Thayer DD, Pflingsten IA (2024) *Nymphoides cristata* (Roxb.) Kuntze: U.S. Geological Survey Nonindigenous Aquatic Species Database. <https://nas.er.usgs.gov/queries/FactSheet.aspx?SpeciesID=2216>. Accessed: April 29, 2024
- Ward SM, Gaskin JF, Wilson LM (2008) Ecological genetics of plant invasion: what do we know? *Invasive Plant Sci Manag* 1:98–109
- Wickham H (2016) *ggplot2: Elegant Graphics for Data Analysis*. New York: Springer. 260 p
- Willey LN, Langeland KA (2011) *Aquatic Weeds: Crested Floating Heart (Nymphoides cristata)*. SS-AGR-344. Gainesville: Florida Cooperative Extension Service, Institute of Food and Agricultural Sciences, University of Florida. 4 p
- Willey LN, Netherland MD, Haller WT, Langeland KA (2014) Evaluation of aquatic herbicide activity against crested floating heart. *J Aquat Plant Manag* 52:47–56
- Williams D, Harms N, Knight I, Grewell B, Futrell C, Pratt P (2020) High genetic diversity in the clonal aquatic weed *Alternanthera philoxeroides* in the United States. *Invasive Plant Sci Manag* 13:217–225
- Williams WI, Friedman JM, Gaskin JF, Norton AP (2014) Hybridization of an invasive shrub affects tolerance and resistance to defoliation by biological control agent. *Evol Appl* 7:381–393
- Zhang Y, Zhang D, Barrett SCH (2010) Genetic uniformity characterizes the invasive spread of water hyacinth (*Eichornia crassipes*), a clonal aquatic plant. *Mol Ecol* 19:1774–1786