

Research Article

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Early weaning-induced intestinal barrier dysfunction associates with Th17/Treg imbalance in squabs

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Abstract

Research on stress damage induced by weaning and its underlying mechanisms in squabs is notably scarce. The study was designed to uncover the potential mechanisms behind the intestinal epithelial barrier impairment due to early weaning (EW) in squabs by evaluating the function of intestinal epithelial barrier, the balance of T helper cell (Th) subsets, and the link between them. A total of 160 hatched squabs were randomly assigned to two groups: one received artificial pigeon milk starting from day 7 post-hatching, while the other group continued to be nourished by their parent pigeons. Ileal tissue and serum samples from eight replicates were gathered for analyses at intervals of 1, 4, 7, 10, 14, and 21 days after weaning. Results showed that body weight of squabs in the EW group decreased significantly from 1 day after weaning and continued throughout the experiment period. The serum endotoxin, diamine oxidase of weaned squabs increased significantly. The mRNA expression of ileal tight junction proteins of weaned squabs was significantly downregulated at multiple time points from 1 to 21 days after weaning. Compared to squabs in the control group, the weaned squabs exhibited immune imbalances of Th1/Th2 and Th17/Treg in ileum, characterized by abnormal expression of specific transcription factors of Th1, Th2, Th17, and Treg, as well as abnormal concentrations of differentiation-inducing cytokines and effector cytokines. Mantel tests showed that the changes of factors related to the differentiation of Th17/Treg cell subsets were significantly correlated with the diamine oxidase, endotoxin level, and the *CDLN1* mRNA expression. Summarily, EW could lead to impaired growth, compromised intestinal epithelial barrier function and an imbalance in the differentiation of Th cell subsets in squabs, among which the dysbalance between Th17 and Treg cells appeared to be more closely associated with the damage of the intestinal epithelial barrier function in early weaned squabs.

Introduction

As a significant emblem of premium livestock offerings, the consumer interest in pigeon squab is experiencing a surge (Ji. et al. 2020). However, in their natural state, the reliance of pigeon squabs on their parents for nourishment post-hatching poses a considerable limitation on the efficiency of production (Jin et al. 2023; Mahdy 2021). Early weaning (EW) and artificial rearing appear to be the optimal strategies for mitigating the pressure of parental feeding and overcoming this production challenge (Xu. et al. 2024). Similar to mammals, EW means that young pigeons leave their dependent living environment, which can cause severe stress both psychologically and physiologically. The stress of EW in mammals has been extensively studied. Smith (Smith and Clark 2010) and Hu (Hu et al. 2013), among others, have discovered that weaning stress results in damage of the intestinal mucosal barrier, diarrhea, and dysbiosis in piglets, which are also attributed to the direct inhibition of growth in these young animals. Research on weaning stress in young pigeons is quite limited. Preliminary studies by our group indicate that the damage to the intestinal epithelial barrier function accompanying EW stress might be a primary factor contributing to the slow growth rate observed in pigeon squabs post-weaning (Xu et al. 2022).

Naive CD4⁺ T-cells can be differentiated into T helper cell (Th) subsets (Th1/Th2/Th17) or regulatory T cell (Treg) on the basis of cytokines in their milieu (Pagani et al. 2013). It is well characterized that balances of Th1/Th2 and/or Th17/Treg are critical for the homeostatic maintenance of immune responses (Gyun et al. 2020). Moreover, studies in humans and mice have shown that the integrity of intestinal epithelial barrier is essential for homeostatic regulation (Cohen and Denkers 2014; Kayama and Takeda 2012). For instance, macromolecular antigens

in the intestinal cavity can be leaked out and captured by T lymphocytes when the integrity of intestinal epithelial barrier is broken, leading to Th subsets imbalance (Chen 2021). Therefore, a novel hypothesis has been proposed that the intestinal epithelial barrier damage induced by EW might be associated with an imbalance in Th cell differentiation in squabs. However, there are currently no data available on the Th subsets imbalance in pigeon squabs with damaged intestinal epithelial barrier function.

The purpose of this study was to elucidate the intrinsic connection between the damage to the intestinal epithelial barrier function induced by EW and the immune imbalance of Th1/Th2 and Th17/Treg by analyzing the changes in growth performance, intestinal permeability, gene expression of tight junction proteins, ileal cytokine concentrations, and mRNA expression of ileal Th1, Th2, Th17, and Treg-specific transcription factors during the development process of squabs undergoing EW. Understanding the changes in gut barrier function and immunity due to weaning stress could provide potential possibilities for mitigating the damage induced by weaning stress in squabs.

Materials and methods

All animal-related experimental procedures were greenlit by the Animal Care and Welfare Committee of Animal Science College and the Scientific Ethical Committee of Zhejiang University (Hangzhou, China).

Animals and experiment layout

On the day of emergence, 160 newly born White King pigeon squabs (*Columba livia*) with comparable body masses were chosen. They were matched up and distributed to the nests of the parent pigeons, replacing the artificial eggs that were previously used to accommodate the nesting instincts of the parent birds. Parent pigeons were fed a grain-based diet. The dietary composition is shown in Supplementary Table 1. Each pair of parent pigeons nurtured two squabs, which were nourished with pigeon milk directly from their parents' beaks to their own. Seven days post-hatching, the squabs with similar body weight (89.39 ± 1.91 g) were randomly allocated into two groups: a control (CON) group and an EW group. Each group consisted of 8 replicates, with 10 squabs in each replicate. The squabs in the CON group continued receiving nourishment from their parent pigeons, whereas the squabs in the EW group were separated from their parents and provided with artificial pigeon milk. The formula of the artificial pigeon milk, which includes 17.76% protein and 13.07 MJ/kg of energy, is shown in Supplementary Table 2. The artificial pigeon milk powder was blended with lukewarm water at a temperature of 37°C and administered into the crop via a sterile syringe and tube for simulated feeding. The environmental settings for both groups were kept consistent. Throughout the duration of the study, the surrounding temperature was maintained between 18 and 26°C, the relative humidity was held steady at 60–70%, and the lighting cycle was set to 12 h of light followed by 12 h of darkness.

Sample gathering

On days 1 (D1), 4 (D4), 7 (D7), 10 (D10), 14 (D14) and 21 (D21) following weaning, eight squabs (one from each replicate) were randomly chosen from both the CON and EW groups for collection. All the squabs were measured for weight and then humanely euthanized through cervical displacement (those exceeding 250 g

Table 1. Targeted gene primers employed in quantitative real-time PCR analysis

Gene	Accession	Primer sequences (5'–3')	Size (bp)
CLDN1	XM_005513213.2	TCTTTGGTGGGGTGATGT	109
		TTGACAGGGGTGTAAGGG	
CLDN3	XM_005515008.2	ACCTCATCCCCGTCTCCT	109
		CAGCCACGTAGAGCGA	
FOXP	XM_065075368.1	CCACCACCGCGCAACT	144
		TCACTCAAAGCAAAAACACGCA	
GATA3	XM_021284192.1	AAGCAGCGTCAAGCACAGAAGG	114
		GGTAGAGTCCACAGGCGTTACAC	
OCLN	XM_005509325.2	CAGGACGTGGCAGAGGA	105
		GTGGAAGAGCTTGTTCGCT	
RORC	XM_021281623.1	TCCTGCTCAAGCGGGTATGG	125
		TGGGGATGGGGATGTGGTGATG	
TBX21	XM_021295578.1	AAACCGCCTCTACCTGCAC	99
		CCCTTGTGTGGTGAGCTTC	
TJP1	XM_021299314.1	GAACCAAAGCCAGTGTATG	159
		GGTCCCCCTCCTCTAATC	
GAPDH	NM_001282835.1	CCGGGCTGCTATTCTCTCTG	116
		TTCCCATCTCGGCCTTGAC	

Note: CLDN, claudin; FOXP, forkhead box protein P; GATA3, GATA binding protein 3; GAPDH, glyceraldehyde-3-phosphate dehydrogenase; OCLN, occludin; RORC, RAR-related orphan receptor C; TBX21, T-box 21; TJP1, tight junction protein 1.

were sedated prior to cervical displacement). The squabs were then sacrificed to obtain serum and small intestine samples. Blood samples were taken for the analyses of serum endotoxin, diamine oxidase, and D-lactic acid levels. The ileum mucosa was gathered to determine the gene expression of Th1, Th2, Th17, and Treg cell-specific transcription factors and tight junction proteins, along with the cytokine levels.

Intestinal permeability

The levels of endotoxin, diamine oxidase, and D-lactate in serum were assessed using a SpectraMax M5 microplate reader (Molecular Devices, Sunnyvale, CA, USA), in conjunction with commercial ELISA kits (Shanghai Enzyme-linked Biotechnology Co., Ltd., Shanghai, China), adhering to the protocol provided by the manufacturer.

Cytokine concentration assessment

Ileal mucosa homogenates were prepared using phosphate-buffered saline for the assessment of cytokine concentrations. The concentrations of interleukin-1 β (IL-1 β), IL-4, IL-5, IL-6, IL-10, IL-12, IL-13, IL-17, IL-23, tumor necrosis factor- α (TNF- α), transforming growth factor (TGF- β), and interferon- γ (IFN- γ) were quantified using commercial ELISA kits (Shanghai Enzyme-linked Biotechnology Co., Ltd., Shanghai, China). Standard samples and diluent were dispensed at a volume of 100 μ L per well in duplicate. The plate was then incubated for 2 h at 37°C and

subsequently washed thrice with a washing solution. A biotin-labeled antibody was introduced at a volume of 100 μ L per well, followed by incubation at 37°C for 1 h and washed five times. A horseradish peroxidase conjugate was then applied at 100 μ L per well, incubated at 37°C for 1 h, and washed five times. Following this, 100 μ L of the coloring reagent was added to each well to develop the color for 15 min at 37°C. To terminate the reaction, 50 μ L of stop solution was introduced into each well. The changes in absorbance at a wavelength of 450 nm were promptly measured using a SpectraMax M5 microplate reader (Molecular Devices, Sunnyvale, CA, USA). The cytokine concentrations were ultimately expressed as picograms per milligram of protein.

RNA extraction and real-time quantitative PCR analysis

Total RNA was extracted from ileal mucosa utilizing the MolPure® TRleasy™ Plus Total RNA Kit (with spin column) (Yeasten Biotechnology Co., Ltd., Shanghai, China) following the protocol of the manufacturer. The quality of the total RNA was evaluated through 1% denaturing agarose gel electrophoresis, while its concentration and purity were determined with the NanoDrop 2000 (Thermo Fisher, Waltham, MA, USA). The OD260/OD280 ratio ranged from 1.8 to 2.0, indicating that the RNA isolate was of high purity and suitable for further tests. Reverse transcription to obtain cDNA was performed utilizing the Hifair® III 1st Strand cDNA Synthesis SuperMix for qPCR (gDNA digester plus) (Yeasten Biotechnology Co., Ltd., Shanghai, China). The abundance of mRNA was detected using the Bio-Rad CFX Manager 3.0 system (CFX96 Touch, Bio-Rad Laboratories, Inc., Hercules, CA, USA). The primers for both the reference and target genes are detailed in Table 1. Real-time quantitative PCR was performed using the Hieff® qPCR SYBR Green Master Mix (No Rox; Yeasen Biotech Co., Ltd., Shanghai, China), with the thermal cycling program set as follows: initial DNA denaturation at 95°C for 5 min, succeeded by 40 cycles of denaturation at 95°C for 10 s and annealing/extension at 60°C for 30 s. The relative expression levels of each gene were computed employing the $2^{-\Delta\Delta Ct}$ approach (Schmittgen 2001), with GAPDH as endogenous reference and the mean ΔCt value of the CON group as calibrator.

Statistical examination

The experimental data were analyzed using an independent-samples *t*-test in SPSS 26.0 (SPSS Inc., Chicago, IL, USA) for Windows. Statistical significance was set at $P < 0.05$, while a higher threshold for significance was designated at $P < 0.01$. Data visualization was achieved using GraphPad Prism 9.0 (GraphPad Software Inc., San Diego, CA, USA). The results are expressed as means accompanied by their standard errors for a group of eight squabs. Mantel tests were carried out utilizing the R software suite (Hy4m/linkET). The asterisks were used to denote statistical significance as follows: an asterisk (*) denotes a significant difference ($P < 0.05$), while double asterisks (**) signify a highly significant difference ($P < 0.01$).

Results

Body weight

The effects of EW on body weight of squabs are depicted in Figure 1. The body weight of early weaned squabs at different time points

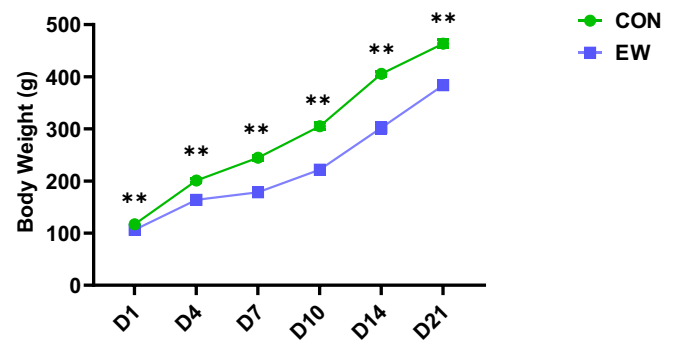


Figure 1. Effects of early weaning on body weight of squabs. The 1D, 4D, 7D, 10D, 14D, and 21D represent the 1st, 4th, 7th, 10th, 14th, and 21st day after weaning, respectively. Values are means with their standard errors of eight squabs. ** $P < 0.01$, respectively, within each time points post-weaning. CON, control group; EW, early weaning group.

(D1, D4, D7, D10, D14, and D21 post-weaning) were markedly decreased ($P < 0.01$) in comparison to that of CON group.

Intestinal permeability

The consequences of EW on the intestinal permeability in squabs are illustrated in Figure 2. The intestinal permeability in squabs was indicated by serum levels of diamine oxidase, endotoxin, and D-lactic acid. EW led to a notable elevation in serum levels of endotoxin and diamine oxidase in squabs at D4, D7, D10, D14, and D21 following weaning ($P < 0.01$, $P < 0.01$, $P < 0.05$, $P < 0.05$, and $P < 0.05$, respectively). Additionally, the serum D-lactic acid level was significantly elevated ($P < 0.05$) in early weaned squabs at D4 post-weaning relative to the CON group, whereas no significant difference ($P > 0.05$) in serum D-lactic acid level was observed between the two groups at the other time points.

Relative gene expression of tight junction proteins

The impacts of EW on the relative gene expression of tight junction proteins in ileum of squabs are presented in Fig. 3. In comparison to the CON group, the mRNA expression of *OCN* and *CLDN1* in the EW group was significantly downregulated from 4 to 21 days after weaning ($P < 0.05$ or $P < 0.01$). The mRNA expression of *TJP1* was markedly downregulated in early weaned squabs at D10 and D14 after weaning ($P < 0.05$, $P < 0.01$). The mRNA expression of *CLDN3* was markedly downregulated from D1 to D21 post-weaning ($P < 0.05$ or $P < 0.01$).

Concentrations of Th cell subset differentiation-inducing factors

Inexperienced $CD4^+$ T cells, referred to as Th0 cells, have the capacity to mature and proliferate into diverse subsets including Th1, Th2, Th17, and Treg under the influence of distinct cytokines. Upon exposure to IL-12 and IL-4, Th0 cells can differentiate into Th1 and Th2 cells, respectively. In the presence of TNF- α , IL-6, IL-1 β , and IL-23, they tend to develop into Th17 cells. When acted upon by TGF- β alone, Th0 cells differentiate into Treg cells. However, when exposed to both TGF- β and IL-6, Th0 cells are steered toward differentiation into Th17 cells instead.

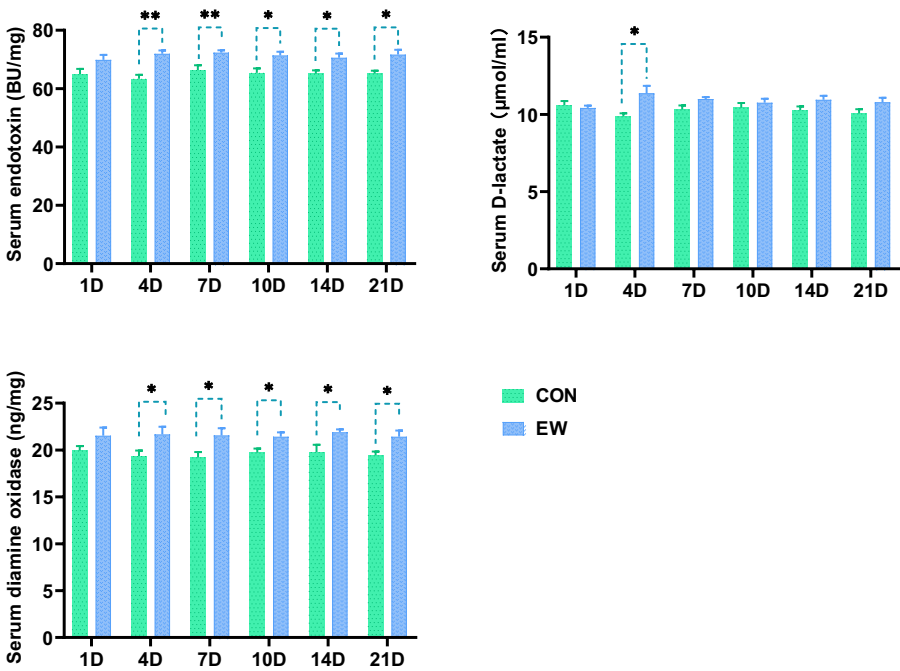


Figure 2. Effects of early weaning on the levels of serum endotoxin, D-lactate and diamine oxidase of squabs. The 1D, 4D, 7D, 10D, 14D, and 21D represent the 1st, 4th, 7th, 10th, 14th, and 21st day after weaning, respectively. Values are means with their standard errors of 8 squabs. * $p < 0.05$ and ** $p < 0.01$, respectively, within each time points post-weaning. CON, control group; EW, early weaning group.

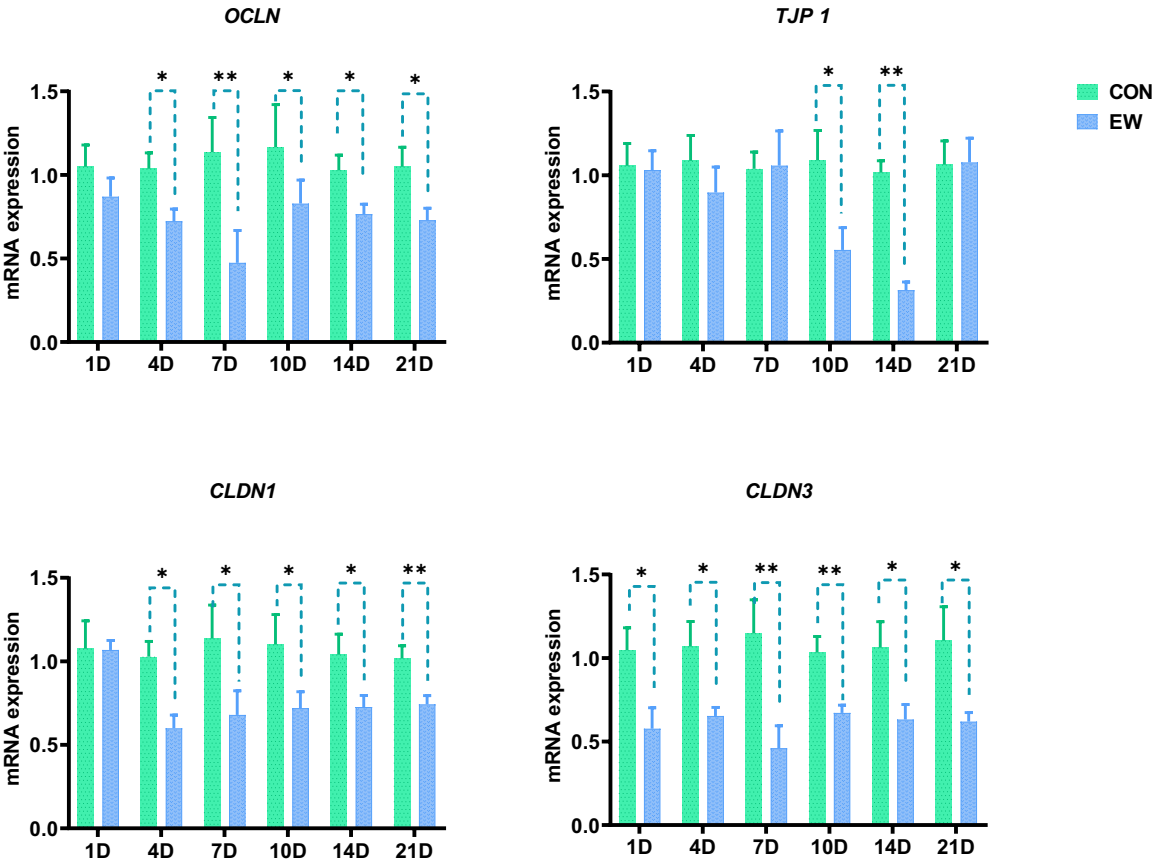


Figure 3. Effects of early weaning on the mRNA expression of tight junction proteins in ileum of squabs. CLDN, claudin; OCLN, occludin; TJP, tight junction protein. The 1D, 4D, 7D, 10D, 14D, and 21D represent the 1st, 4th, 7th, 10th, 14th, and 21st day after weaning, respectively. * $p < 0.05$ and ** $p < 0.01$, respectively, within each time points post-weaning. CON, control group; EW, early weaning group.

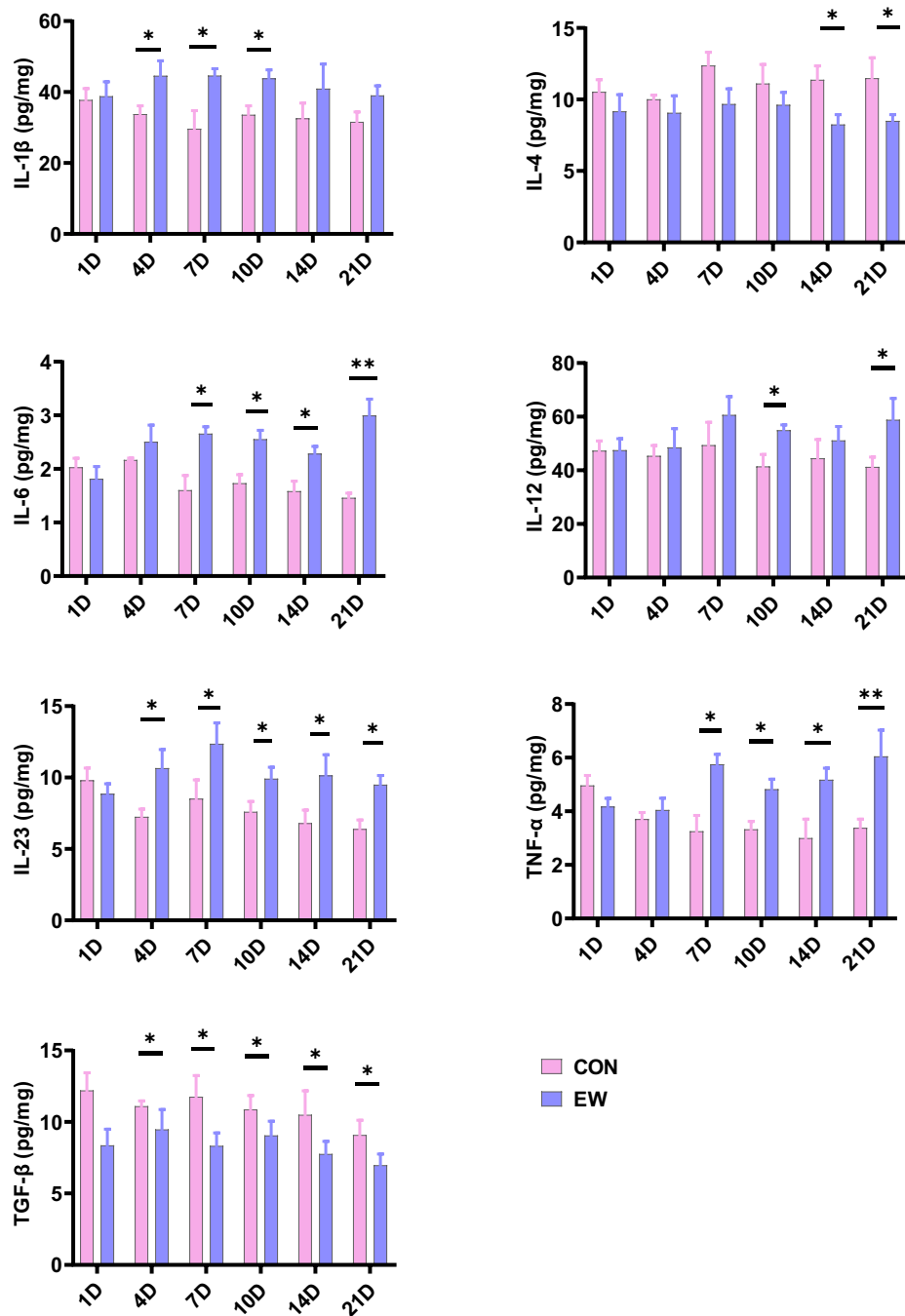


Figure 4. Effects of early weaning on the levels of differentiation-inducing factors (IL-1 β , IL-4, IL-6, IL-12, IL-23, TNF- α , TGF- β) of Th1, Th2, Th17, and Treg in ileum of squabs. The 1D, 4D, 7D, 10D, 14D, and 21D represent the 1st, 4th, 7th, 10th, 14th, and 21st day after weaning, respectively. * $p < 0.05$ and ** $p < 0.01$, respectively, within each time points post-weaning. CON, control group; EW, early weaning group.

The concentrations of Th1, Th2, Th17, and Treg differentiation-inducing factors in ileum of the squabs in the CON and EW groups are displayed in Figure 4. In comparison to the CON group, the EW group presented a significant rise in IL-12 level at D10 and D21 after weaning ($P < 0.05$). The IL-4 level markedly reduced in the early weaned squabs from 14 to 21 days post-weaning ($P < 0.05$), while the levels of IL-1 β and IL-23 significantly increased in the early weaned squabs from 4 to 21 days post-weaning ($P < 0.05$). Additionally, the levels of IL-6 and TNF- α significantly increased in the early weaned squabs from 7 to 21 days post-weaning

($P < 0.05$ or $P < 0.01$). Besides, the TGF- β level significantly decreased in the EW group from 1 to 21 days post-weaning ($P < 0.05$).

Gene expression of Th cell subset-specific transcription factors

T-bet, GATA3, ROR γ t, and FOXP serve as specific transcription factors which control the differentiation of Th0 cells into Th1, Th2, Th17, and Treg subtypes, respectively. The genes *TBX21*, *GATA3*, *RORC*, and *FOXP* are responsible for encoding these four

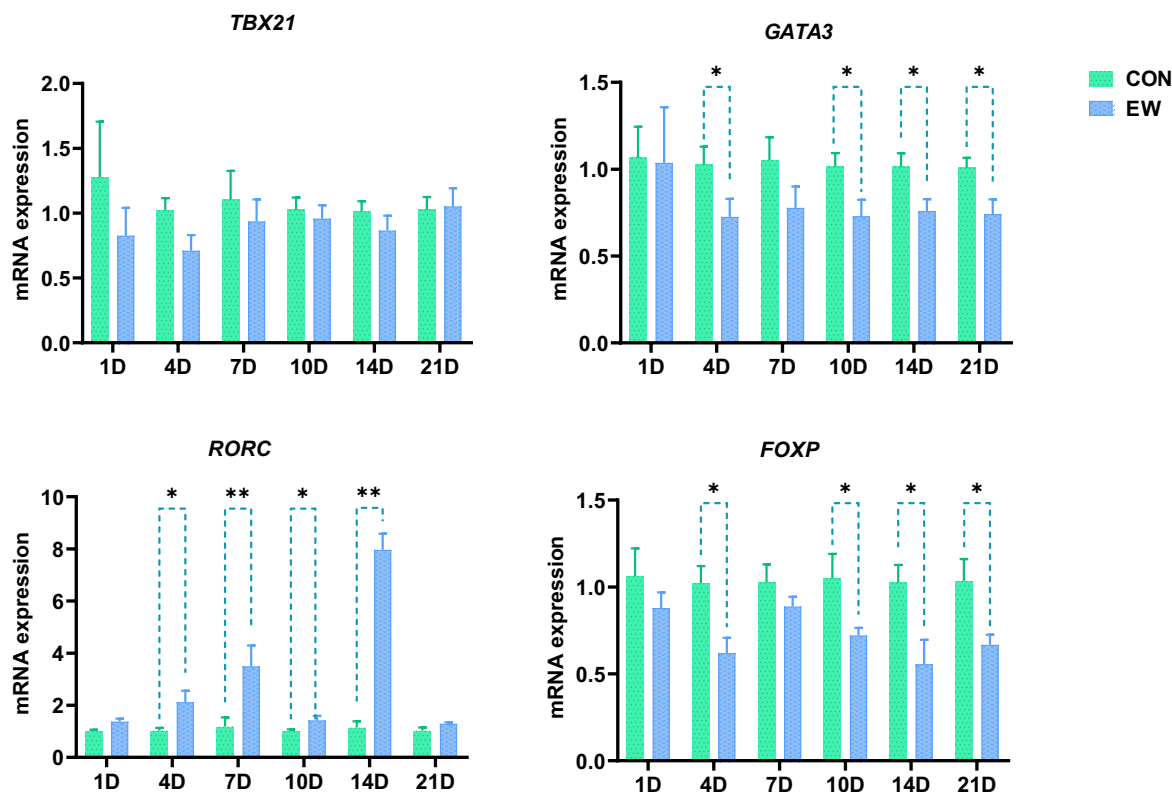


Figure 5. Effects of early weaning on the mRNA expression of specific transcription factors of Th1, Th2, Th17, and Treg in ileum of squabs. The 1D, 4D, 7D, 10D, 14D, and 21D represent the 1st, 4th, 7th, 10th, 14th, and 21st day after weaning, respectively. * $p < 0.05$ and ** $p < 0.01$, respectively, within each time points post-weaning. *TBX21*, T-box transcription factor 21; *GATA3*, GATA binding protein 3; *RORC*, RAR-related orphan receptor C; *Foxp*, forkhead box protein P; CON, control group; EW, early weaning group.

transcription factors. The changes in the mRNA expression of the transcription factors for Th1, Th2, Th17, and Treg cells within 21 days post-weaning in pigeon squabs are presented in Figure 5. In the EW group, the mRNA expression of the transcription factor *RORC* was significantly elevated compared to the CON group from D4 to D14 post-weaning ($P < 0.05$ or $P < 0.01$), while the mRNA expression of *GATA3* and *FOXP* was significantly reduced compared to the CON group at D4 and from D10 to D21 post-weaning ($P < 0.05$). There was no notable variation in the *TBX21* mRNA expression between these two groups ($P > 0.05$).

Concentrations of effector cytokines of Th cell subsets

Th1 cells primarily secrete cytokines such as INF- γ and TNF- α ; Th2 cells chiefly produce cytokines like IL-5, IL-6, and IL-13; Th17 cells predominantly secrete cytokines including IL-17 and IL-22; while Treg cells mainly release cytokines such as IL-10 and TGF- β . The cytokines secreted by Th1, Th2, Th17, and Treg in ileum of squabs in the CON and EW groups were mainly depicted in Figure 6. In comparison to the CON group, the levels of INF- γ and TNF- α (Figure 4) in the EW group were markedly increased from 7 days after weaning ($P < 0.05$). In comparison to the CON group, the IL-6 (Fig. 4) level in the EW group was markedly increased from 7 days after weaning ($P < 0.05$), while the IL-5 level was markedly reduced in the EW group from 7 days after weaning ($P < 0.05$). Besides, the IL-13 level had no significant change at each time point after weaning ($P > 0.05$). The IL-17 in the EW group was markedly increased from 7 days after weaning ($P < 0.05$), and the IL-22 level

was markedly increased in the EW group from 10 days after weaning ($P < 0.05$). Besides, the IL-10 level was markedly reduced in the EW group from 10 days after weaning ($P < 0.05$). The TGF- β (Figure 4) level was markedly decreased in the EW group from 1 day after weaning ($P < 0.05$).

Correlation analysis

Potential associations between factors related to Th cell subset differentiation balance and indicators related to intestinal epithelial barrier function were evaluated based on Mantel tests (Figure 7). The results showed that during EW, the changes of differentiation-related factors of Th17/Treg cell subsets were significantly correlated with the levels of serum diamine oxidase, serum endotoxin, and the mRNA expression of *CDLN1* in ileum ($P < 0.01$). The Th1/Th2 cell subsets differentiation-related factors were significantly correlated with serum endotoxin level ($P < 0.05$).

Discussion

In studies on mammals, it has been found that the sudden dietary switch from breastfeeding to solid nourishment after weaning could cause insufficient intake in young animals, which may subsequently result in inadequate utilization of feed nutrients, localized inflammatory reactions and oxidative stress, excessive proliferation of harmful bacteria, diarrhea, suboptimal growth, and potentially mortality (Campbell et al. 2013; Yang et al. 2021). The phenomenon of growth inhibition also occurred in the weaned

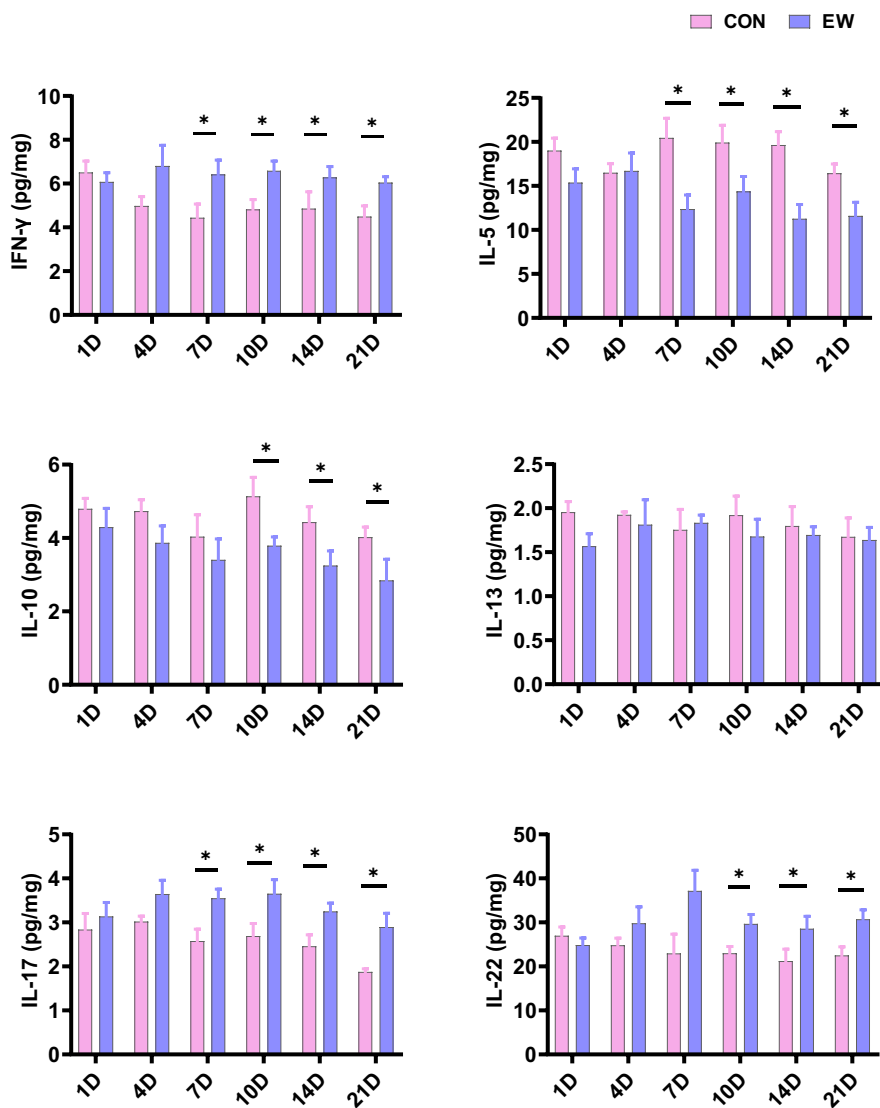


Figure 6. Effects of early weaning on the secretion of cytokines (IFN-γ, IL-5, IL-10, IL-13, IL-17, IL-22) by Th1, Th2, Th17, and Treg cells in ileum of squabs. The 1D, 4D, 7D, 10D, 14D, and 21D represent the 1st, 4th, 7th, 10th, 14th, and 21st day after weaning, respectively. * $p < 0.05$ within each time points post-weaning. CON, control group; EW, early weaning group.

squabs in our study. The outcomes revealed that the growth performance of the early weaned squabs conspicuously lagged behind that of the parent-fed squabs as early as 1 day post-weaning. The nutrient absorption primarily occurs in the intestine, and a well-functioning intestine plays a pivotal role in promoting animal growth and development while serving as a critical physiological foundation for enhancing immune competence (Greenwood-Van et al. 2017; Pluske et al. 2018). Similar to mammals, squabs also experienced a sudden change in environment and diet during weaning. Therefore, the damage to the intestinal structure and immune function caused by weaning stress might be a direct reason for the growth impairment observed in weaned pigeons. An important target in current nutritional studies is to ensure the maintenance of the intestinal structural integrity and its standard functionality (Cui et al. 2021; Guo et al. 2021). Therefore, the study further investigated the implications of EW on the intestinal epithelial barrier and immune function in squabs. Serum diamine oxidase, D-lactic acid, and endotoxin levels are commonly used to reflect intestinal permeability (Xu et al. 2024). The findings of the current research showed that at one or multiple age in days post-weaning, there was a significant boost in the levels of serum diamine oxidase, D-lactic acid, and endotoxin, pointing to a rise

in intestinal permeability in weaned squabs. Notably, the elevated serum endotoxin and diamine oxidase persisted up to 21 days post-weaning, which was in agreement with the findings in the previous study (Xu et al. 2022). A sustained elevation in intestinal permeability indicated that the epithelial barrier was compromised in weaned pigeons. Additionally, tight junctions which form a highly polarized barrier are indispensable elements of the intestinal epithelial barrier and are crucial for upholding the internal environment stable. Impaired synthesis or reduced activity of these proteins may increase the selective permeability of the intestinal epithelial barrier (Chen et al. 2016). The outcomes of the present investigation demonstrated a significant downturn in the mRNA expression of *OCN*, *CLDN1*, *CLDN3*, and *TJP1* in the EW group, with low mRNA expression of *OCN*, *CLDN1*, and *CLDN3* being persistent. These findings further illustrated that the intestinal epithelial barrier in weaned pigeons suffered damage to its integrity, and the injury was long-term, not improving with age.

The development of the intestinal mucosal immune system represents the maturation process of both innate and acquired immunity. The physical defense provided by the intestinal epithelial lining, the protective mucosal coat, complement, antimicrobial

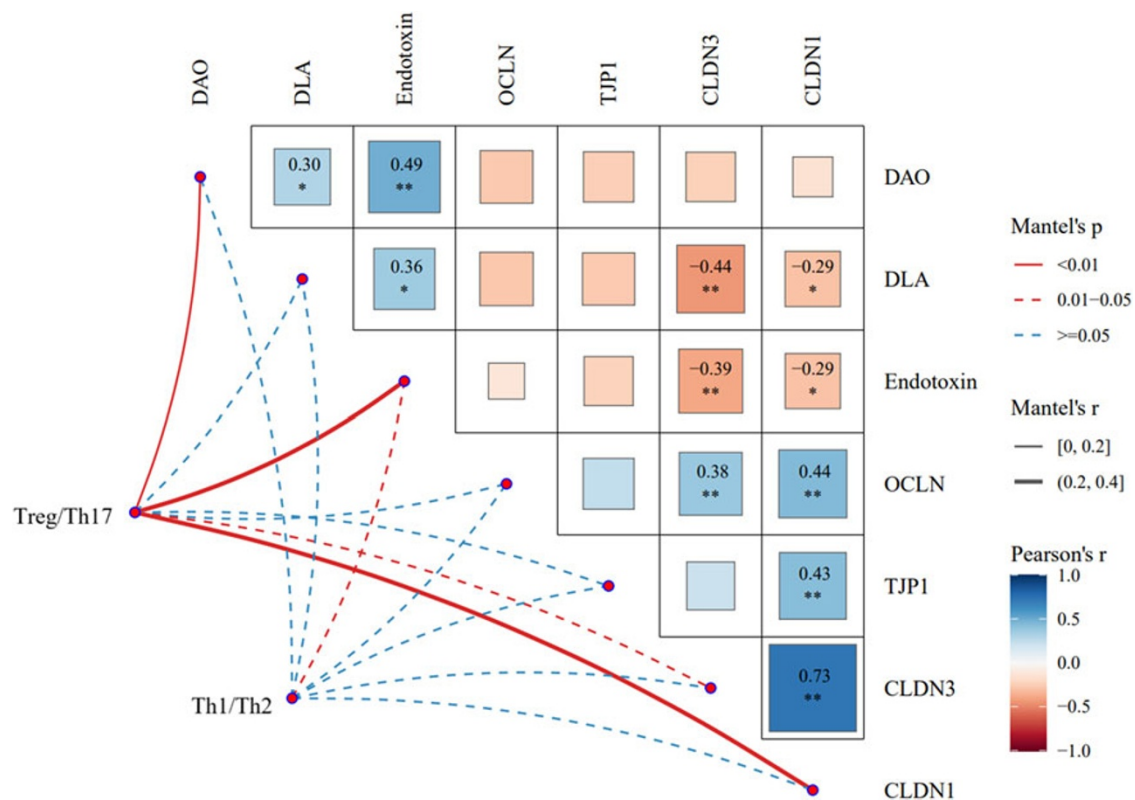


Figure 7. Correlation of Th1, Th2, Th17, and Treg with epithelial barrier function was evaluated based on mantel test. CLDN, claudin; OCLN, occlude; TJP, tight junction protein; DAO, diamine oxidase; DLA, D-lactate.

peptides, and the cytokines released by resident immune cells like macrophages and dendritic cells collectively constitute the innate immune system (Malago 2015). CD4⁺ T cells serve as pivotal intermediaries and crucial regulators of adaptive immunity, with their differentiation pathways being controlled by various factors including antigen-specific stimulation, cytokines, and transcription factors (Gyun et al. 2020). Depending on the cytokine expression profiles and differences in immunoregulatory functions, effector T cells derived from CD4⁺ T cell differentiation are categorized into distinct functional subsets (Shale et al. 2013). These subsets include Th1, Th2, Th17, and Treg, among others, which coordinate with each other to collectively exert immunoregulatory effects. It is commonly believed that Th1 and Th17 subsets, along with their associated cytokines, play pro-inflammatory roles, while Th2 and Treg subsets, along with their respective cytokines, exert anti-inflammatory effects. The dynamic balance between Th1/Th2 and Th17/Treg subsets is considered a crucial mechanism for maintaining immune stability (Aiping et al. 2010; Fang et al. 2019). Once this dynamic balance is disrupted, it can lead to or exacerbate intestinal inflammation. On account of the presence of intestinal epithelial barrier, the vast majority of microbes normally do not interact directly with immune cells. Once the intestinal epithelial barrier is damaged, macromolecular antigens such as microorganisms in the intestinal cavity are more likely to leak out, and thus be captured by intraepithelial lymphocytes, leading to the imbalance of Th cell differentiation (Chen 2021). Thus, we hypothesize that the intestinal epithelial barrier damage induced by EW might be associated with an imbalance in Th cell differentiation in ileum of squabs. Consequently, the differentiation of four Th cell subsets and the changes of related cytokine networks

were measured. The results revealed that in the ileum of weaned squabs, cytokines that induce Th1 and Th17 cell differentiation significantly increased at multiple time points, while cytokines that induce Th2 and Treg differentiation significantly decreased at various points post-weaning. The mRNA expression of *GATA3* and *FOXP3*, nuclear transcription factors associated with Th2 and Treg cells, respectively, significantly decreased. Conversely, the mRNA expression of *RORC*, a nuclear transcription factor associated with Th17 cells, significantly increased. The effector cytokine levels associated with Th1 and Th17 significantly rose, while those associated with Th2 and Treg either dropped or displayed no significant change. Collectively, these findings indicated that post-weaning squabs exhibited a disruption of Th1/Th2 and Th17/Treg immune homeostasis in the ileum. It is noteworthy that the significant changes in the expression of transcription factors were commonly observed at 4 days post-weaning, whereas the significant changes in effector cytokine levels generally became apparent starting at 7 days post-weaning.

Prior research has shown that intestinal inflammation in mice can be mediated by Th1/Th2 cells and is associated with abnormal concentration of IL-1 β , IL-6, IL-12, TNF- α , and IFN- γ (Peng 2012). Specifically, Th17 cells can mediate Th17-type immunity at the intestinal mucosa by secreting the distinct cytokines IL-22 and IL-17, which in turn induces the occurrence of epithelial inflammation in the intestine (Proietti et al. 2014; Ray et al. 2014). Treg cells and the specific cytokine IL-10 secreted by Treg cells negatively regulated the mucosal immune process, and at the same time promoted the renewal and remodeling of small intestinal epithelial cells (Mayne and Williams 2013; Rau et al. 2018). In short, The cells in the intestine participate in the development

of intestinal epithelium and are instrumental in preserving the homeostasis of the intestinal epithelial barrier together with their related cytokines. Presumably, EW is linked to the occurrence of Th1/Th2 and Th17/Treg immune imbalances in small intestine of squabs, which in turn altered the cytokine environment in the intestine, ultimately leading to aggravation of intestinal barrier injury in squabs. To further verify intrinsic connection between the compromise of intestinal epithelial barrier function resulting from EW and the Th1/Th2, Th17/Treg immune imbalance in squabs, Mantel test was employed to assess the correlation between these related factors. The results revealed that during the process of EW, the Th17/Treg imbalance was highly significantly correlated with increased intestinal mucosal permeability and abnormal expression of tight junction proteins. Meanwhile, the Th1/Th2 imbalance was only moderately correlated with a decrease in serum endotoxin level. These suggested that the Th17/Treg imbalance appeared to be more closely associated with the intestinal epithelial barrier impairment in pigeon squabs undergoing EW.

However, two limitations of this study must be taken into consideration. First, it is important to point out that differentiation of naive CD4⁺ T cells into Th (Th1/Th2/Th17) or Treg cells is influenced by many exogenous and endogenous factors. For example, intestinal commensal bacterial metabolites or bacteria themselves can activate naive CD4⁺ T cells to effector T cells, Tregs, or Th17 cells by altering cytokines environment through pattern-recognition receptors pathways (Gopalakrishnan et al. 2018). Because several previous studies have shown that the composition of commensal bacterial and their metabolites is influenced by diet composition (Liu et al. 2017), the effect of diets on Th cell differentiation should not be ignored. As such, it will be necessary to further investigate whether and how diets and microbiota are involved in regulating Th subsets imbalance in pigeon squabs during EW period. Second, we do not elucidate the mechanisms behind the interplay between the Th17/Treg imbalance and the intestinal epithelial barrier function impairment in early weaned squabs. Further studies are required to determine the signal transduction pathways involved. Nevertheless, our results are probably useful for researchers as a first step to understand the potent role of Th subsets in the development and progression of intestinal epithelial barrier function in pigeon squabs under stress.

Conclusion

In conclusion, EW stress could bring about growth retardation in squabs. The intestinal epithelial barrier function was compromised in weaned squabs, primarily manifesting as increased intestinal mucosal permeability, loose tight junctions, and an imbalance in the Th subsets and their cytokine network. Moreover, correlation analysis suggested that the Th17/Treg imbalance might be more closely associated with the damage of intestinal epithelial barrier function in young pigeons undergoing EW.

Supplementary material. The supplementary material for this article can be found at <https://doi.org/10.1017/anr.2025.6>.

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Conflict of interest. The contributors state that they have no conflicting interests.

References

- Aiping B, Nonghua L, Hao Z *et al.* (2010) All-trans retinoic acid ameliorates trinitrobenzene sulfonic acid-induced colitis by shifting Th1 to Th2 profile. *Journal of Interferon and Cytokine Research* **30**(6), 399–406.
- Campbell JM, Crenshaw JD and Polo J (2013) The biological stress of early weaned piglets. *Journal of Animal Science and Biotechnology* **4**(1), 19.
- Chen D (2021) Cytokine regulation and function in T cells. *Annual Review of Immunology* **39**, 51–76.
- Chen X, Naehrer K and Applegate T (2016) Interactive effects of dietary protein concentration and Aflatoxin B1 on performance, nutrient digestibility, and gut health in broiler chicks. *Poultry Science* **95**, 1312–1325.
- Cohen SB and Denkers EY (2014) Border maneuvers: Deployment of mucosal immune defenses against *Toxoplasma gondii*. *Mucosal Immunology* **7**(4), 744–752.
- Cui L, Guan X, Ding W *et al.* (2021) *Scutellaria baicalensis* Georgi polysaccharide ameliorates DSS-induced ulcerative colitis by improving intestinal barrier function and modulating gut microbiota. *International Journal of Biological Macromolecules* **166**, 1035–1045.
- Fang YC, Juan JZ, Chao Q *et al.* (2019) Inonotus obliquus polysaccharide ameliorates dextran sulphate sodium induced colitis involving modulation of Th1/Th2 and Th17/Treg balance. *Artificial Cells, Nanomedicine, and Biotechnology* **47**(1), 757–766.
- Gopalakrishnan V, Helmink BA, Spencer CN *et al.* (2018) The influence of the gut microbiome on cancer, immunity, and cancer immunotherapy. *Cancer Cell* **33**(4), 570–580.
- Greenwood-Van MB, Johnson AC and Grundy D (2017) Gastrointestinal physiology and function. *Handbook of Experimental Pharmacology* **239**, 1–16.
- Guo S, Geng W, Chen S *et al.* (2021) Ginger alleviates DSS-induced ulcerative colitis severity by improving the diversity and function of gut microbiota. *Frontiers in Pharmacology* **12**, 632569.
- Gyun HL, Zi MC and Min JC (2020) Bystander CD4⁺T cells: Crossroads between innate and adaptive immunity. *Experimental and Molecular Medicine* **52**(8), 1255–1263.
- Hu CH, Xiao K, Luan SZ *et al.* (2013) Early weaning increases intestinal permeability, alters expression of cytokine and tight junction proteins, and activates mitogen-activated protein kinases in pigs. *Journal of Animal Science* **91**(3), 1094–1101.
- Ji F, Zhang D, Shao Y *et al.* (2020) Changes in the diversity and composition of gut microbiota in pigeon squabs infected with *Trichomonas gallinae*. *Scientific Reports* **10**, 19978.
- Jin C, Y. H, Jiang S *et al.* (2023) Chemical composition of pigeon crop milk and factors affecting its production: A review. *Poultry Science* **102**, 102681.
- Kayama H and Takeda K (2012) Regulation of intestinal homeostasis by innate and adaptive immunity. *International Immunology* **24**(11), 673–680.
- Liu J, Bian G, Sun D *et al.* (2017) Starter feeding supplementation alters colonic mucosal bacterial communities and modulates mucosal immune homeostasis in newborn lambs. *Frontiers in Microbiology* **8**, 429.
- Mahdy MAA (2021) Comparative morphological study of the oropharyngeal floor of squabs and adult domestic pigeons (*Columba livia domestica*). *Microscopy Research and Technique* **84**, 499–511.
- Malago JJ (2015) Contribution of microbiota to the intestinal physicochemical barrier. *Beneficial Microbes* **6**(3), 295–311.
- Mayne CG and Williams CB (2013) Induced and natural regulatory T cells in the development of inflammatory bowel disease. *Inflammatory Bowel Diseases* **19**(8), 1772–1788.
- Pagani M, Rossetti G, Panzeri I *et al.* (2013) Role of microRNAs and long-non-coding RNAs in CD4⁺ T-cell differentiation. *Immunological Reviews* **253**(1), 82–96.
- Peng X (2012) *The Expression Study of Th1/Th2 Inflammatory Cytokines in Rat Treatment Model of Ulcerative Colitis*. Changsha: Central South University, M.D.
- Pluske RJ, Turpin LD and Kim J (2018) Gastrointestinal tract (gut) health in the young pig. *Animal Nutrition* **4**(2), 187–196.

- Proietti M, Cornacchione V, Rezzonico JT *et al.* (2014) ATP-gated ionotropic P2X7 receptor controls follicular T helper cell numbers in Peyer's Patches to promote host-microbiota mutualism. *Immunity* **41**(5), 789–801.
- Rau M, Rehman A, Dittrich M *et al.* (2018) Fecal SCFAs and SCFA-producing bacteria in gut microbiome of human NAFLD as a putative link to systemic T-cell activation and advanced disease. *United European Gastroenterology Journal* **6**(10), 1496–1507.
- Ray S, De Salvo C and Pizarro TT (2014) Central role of IL-17/Th17 immune responses and the gut microbiota in the pathogenesis of intestinal fibrosis. *Current Opinion in Gastroenterology* **30**(6), 531–538.
- Schmittgen TD (2001) Real-time quantitative PCR. *Methods* **25**, 383–385.
- Shale M, Schiering C and Powrie F (2013) CD4⁺ T-cell subsets in intestinal inflammation. *Immunological Reviews* **252**(1), 164–182.
- Smith F and Clark JE (2010) Early weaning stress impairs development of mucosal barrier function in the porcine intestine. *American Journal of Physiology-Gastrointestinal and Liver Physiology* **298**(3), G352–G363.
- Xu Q, M. L., Yuan Y *et al.* (2024) Early weaning damages the intestinal epithelial barrier of squabs through toll-like receptor signaling pathways. *Poultry Science* **103**(12), 104243.
- Xu QQ, Jian HF, Zhao WY *et al.* (2022) Early weaning stress induces intestinal microbiota disturbance, mucosal barrier dysfunction and inflammation response activation in pigeon squabs. *Frontiers in Microbiology* **13**, 877866.
- Yang Z, Wang Y, T. H *et al.* (2021) Effects of dietary yucca schidigera extract and oral *Candida utilis* on growth performance and intestinal health of weaned piglets. *Frontiers in Nutrition* **8**, 685540.