


REVIEW ARTICLE

Caves on Earth as proxies for Martian subsurface environments

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Abstract

On planet Earth, the inner part of cave systems represents an extreme and isolated environment, characterized by relatively stable microclimatic conditions, nutrient limitation and protection from external stressors such as radiation, wind and surface weathering. These features make them suitable analogues to Martian cave systems, which are identified as high-priority targets for astrobiological investigations. Microbial communities inhabiting Earth's caves exhibit extremotolerant or extremophilic traits, offering valuable insights into potential microbial survival strategies on Mars. This mini-review explores the role of cave microbiomes as models for studying habitability, biosignature preservation and microbial adaptations relevant to Martian subsurface environments. We summarize recent findings on cave microbial diversity and their metabolic strategies, highlighting their implications for astrobiology. Additionally, we discuss how caves can function as biological time capsules, preserving biosignatures and microbial life relevant to future planetary exploration. We conclude that terrestrial caves offer key ecological and mineralogical analogs to Martian subsurface environments, and that microbial strategies observed in cave ecosystems such as chemolithotrophy, endolithic colonization and biofilm formation should be prioritized in designing future Mars life-detection missions.

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Introduction

The search for life beyond Earth has long focused on Mars, a planet that once had conditions more hospitable to life (Wordsworth, 2016; Adams, 2025). While the planet's surface today is cold, dry and bombarded by cosmic radiation and small meteorites, its subsurface offers a more protected and potentially habitable environment for microorganisms. Martian caves are among the most promising locations for astrobiological exploration, which may serve as refuges for microbial life or as sheltered archives preserving biosignatures, i.e., chemical, biological or physical evidence of past or present life, including the existence of ancient ecosystems. The existence of caves on Mars, particularly lava tubes, have been confirmed through high-resolution orbital imaging and application of cutting-edge artificial intelligence-powered image scan approaching, marking them as high-priority targets for future robotic and human exploration (Léveillé and Datta, 2010; Blank *et al.*, 2018; Watson and Baldini, 2024; Domínguez *et al.*, 2025). Compared to terrestrial geomorphological context of caves, the most likely type of underground systems on Mars is lava tubes and basaltic caves (Sauro *et al.*, 2020; Léveillé & Datta, 2010).

On Earth, caves represent unique and extreme ecosystems, providing valuable analogs for studying life in extraterrestrial subsurface environments. Caves are commonly characterized by challenging abiotic factors (i.e., oligotrophy, total darkness, high mineral concentrations, high concentration of CO₂ and variable pH levels) but relatively stable in terms of microclimatic conditions, thus providing a buffered habitat from surface environmental fluctuations. (Léveillé *et al.*, 2010; Colak and Güngör, 2022). These characteristics make terrestrial caves highly relevant for understanding habitability, biosignatures preservation and microbial survival strategies in extreme environments (Blank *et al.*, 2018). Microbial communities thriving in Earth's caves have evolved specialized metabolic strategies, including chemolithoautotrophy. These microbes harness energy from reduced chemical compounds such as nitrogen, sulfur, iron, manganese and atmospheric trace gases, supporting survival and growth under nutrient limitation. These strategies may resemble the survival strategies of potential Martian microbes (Bay *et al.*, 2024; Jurado *et al.*, 2024).

The discovery of over 1000 potential cave entrances on Mars, detected through data from the Mars Reconnaissance Orbiter's CTX and HiRISE cameras, underscores their significance for astrobiological investigations. Unlike the exposed Martian surface, which is subjected to high levels of ionizing radiation (~250 mGy/year), extreme temperature fluctuations, micro-meteorites rain and desiccation, caves provide natural shielding that could allow microbial communities to survive in more stable conditions. Additionally, the presence of subsurface water ice in multiple Martian regions suggests that transient liquid brines may persist in cave environments, further supporting life offering a potential medium for microbial metabolism (Martínez and Renno, 2013; Ojha *et al.*, 2015). Should Mars have ever hosted microbial life, caves would constitute some of the most promising environments in which to investigate extant or fossilized biosignatures. Consequently, robotic mission concepts are being tested in terrestrial lava tubes that serve as analogues for Martian subsurface systems (Domínguez *et al.*, 2025).

Earth's caves provide a natural laboratory for studying how life can persist in environments deprived of sunlight and with limited nutrients (Figure 1). Many of the extremophiles found in terrestrial caves, including psychrophiles, halophiles, acidophiles and chemolithotrophs, demonstrate metabolic flexibility and resilience that could be applicable to potential Martian organisms (Sarbu *et al.*, 1996; Chen *et al.*, 2009; Northup *et al.*, 2011; Tetu *et al.*, 2013; Ortiz *et al.*, 2014; Nguyễn-Thùy *et al.*, 2017; Waring *et al.*, 2017; Cheng *et al.*, 2021; Nicolosi *et al.*, 2022; Bay *et al.*, 2024). Furthermore, cave environments preserve biosignatures exceptionally well due to their stable microclimatic conditions. On Earth, microbial activity leaves behind distinct biochemical, isotopic and morphological traces, such as silica deposits, iron- and manganese-stained biofilms and fossilized microbial filaments, all of which could provide templates for biosignature detection on Mars (Preston *et al.*, 2014; Hays *et al.*, 2017). Terrestrial analog studies, such as those conducted in Lechuguilla Cave (USA), Cueva de Villa Luz (Mexico) and Rio Tinto (Spain), offer direct insights into how microbial communities interact with

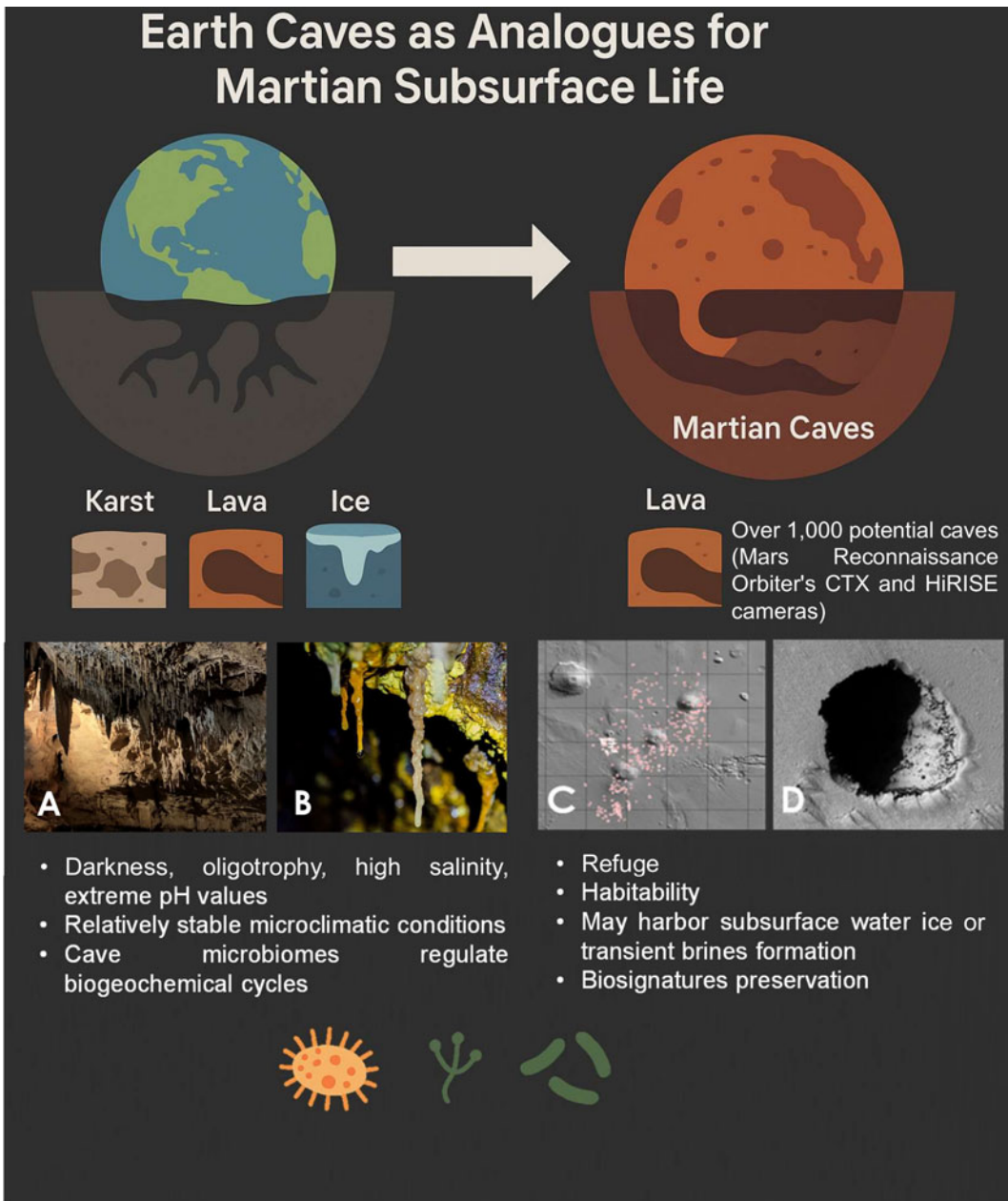


Figure 1. Schematic summary of key environmental features and astrobiological potential of Martian cave systems. Panels A and B show images from Grotta del Caudano (Credit: Dr. Federico Biagioli). Panels C and D are from the U.S. Geological Survey <https://www.usgs.gov>.

geochemical environments in ways that could inform future planetary exploration missions (Tomczyk-Żak *et al.*, 2016; D'Auria *et al.*, 2018; Zhu *et al.*, 2024).

This mini-Review synthesizes current knowledge on terrestrial cave microbiology and its relevance for Martian astrobiology, focusing on three key aspects: *i*) the habitability of cave environments, comparing geochemical and environmental conditions on Earth and Mars; *ii*) microbial diversity and metabolic adaptations in terrestrial caves, with emphasis on extremophiles that serve as potential

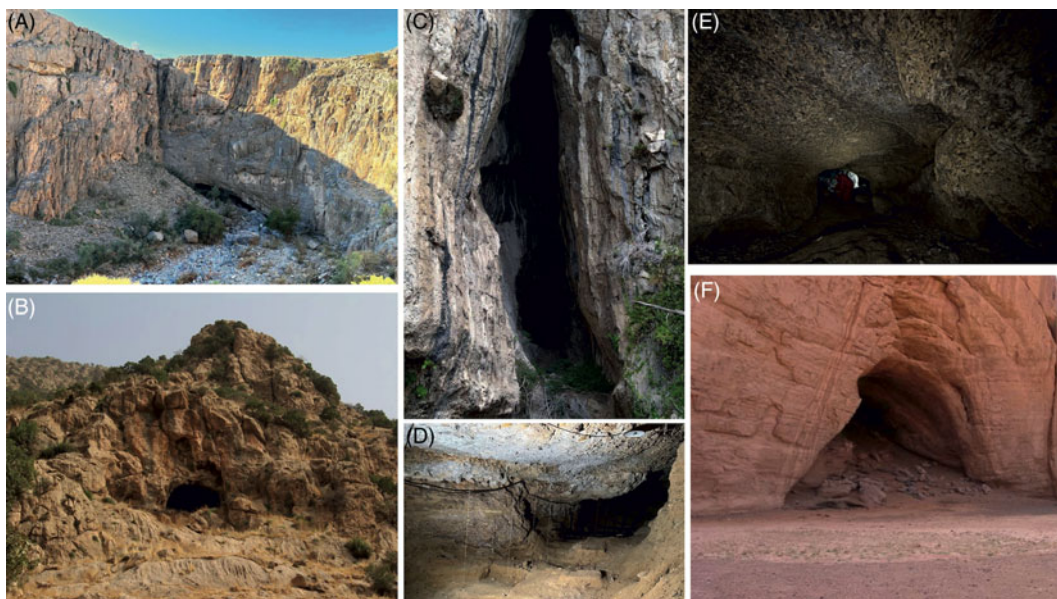


Figure 2. Examples of terrestrial caves carved into different geological bedrocks by karst dissolution or silicates solution. **A)** Northern entrance of the Al Hoota Cave (limestone of the Al Hajar Mt., Sultanate of Oman). **B)** A cave along the limestone of the Zagros Mt. (Kurdistan Region of Iraq). **C)** A cave along the Balzi Rossi cliff (calcarenite, Ventimiglia, Northern Italy). **D)** Madonna dell'Arma Cave carved into conglomerates (Sanremo, Northern Italy). **E)** The Tana della Mussina Cave in the gypsum bedrock of the northern Apennines of Italy (picture: M. Cremaschi). **F)** The Uan Afuda Cave carved into quartz-bearing sandstone (SW Libya).

analogs for Martian life; and *iii*) the preservation of biosignatures in caves, exploring how microbial activity can leave long-lasting traces that may inform life-detection strategies for Mars missions.

Understanding the microbial ecology of Earth's caves provides critical insights into potential Martian subsurface life, refining the strategies for biosignature detection and guiding the design of future robotic exploration missions.

Cave environments as Martian analogs

Caves on Earth are generally defined as openings large enough for human exploration and exploitation (White and Culver, 2019). From the geological point of view, caves are voids below the ground surface, formed by a variety of geological and geomorphological processes. Most cave systems are related to karst processes (Figure 2) and formed in limestone, dolomite, evaporites and conglomerates due chemical dissolution (Jones and White, 2019). Occasionally, also silicates and sandstone bedrocks host cave systems in their subsurface (Figure 2), formed due to solutional processes (mainly driven by hydrolysgais; Wray, 1997; Young *et al.*, 2009; Zerboni *et al.*, 2015). In general, caves represent a gradient of environments, as environmental conditions vary with increasing distance from the entrance and according to the complexity of their development (Lauritzen, 2018). In cave-entrance areas, parameters such as temperature, humidity, ventilation and light exposure do not differ substantially from external conditions. However, deeper within the conduits, environmental conditions change markedly, with implications for the nature of biological colonization (Tuttle and Stevenson, 2011; Lauritzen, 2018). A specific variety of caves – lava tubes (Figure 3) – have been described also in volcanic contexts throughout the world (Sauro *et al.*, 2020). They form as the result of surface cooling of lava flows or, alternatively, the result of sequential lava flows. In the latter case, later flows go underneath the older

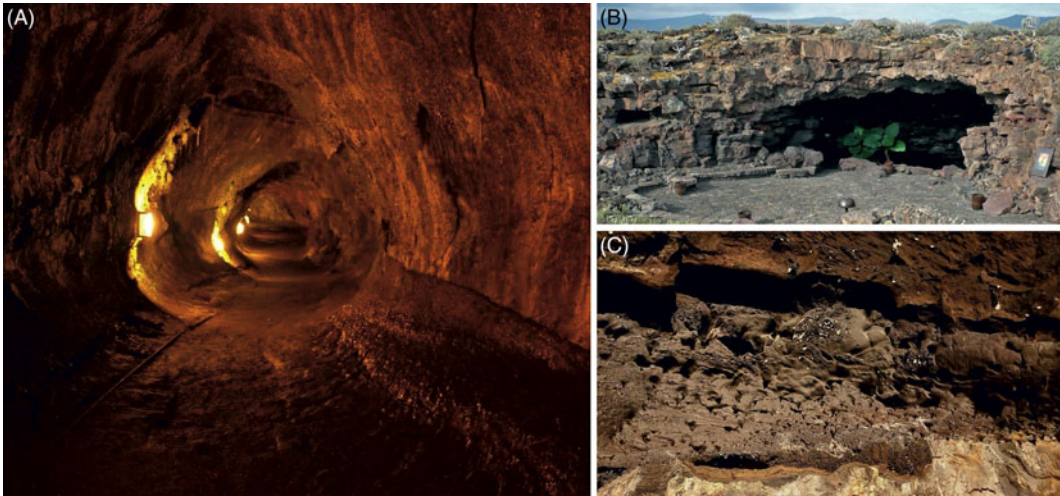


Figure 3. Examples of lava tubes from (A) the Hawai'i Island and (B) Lanzarote; (C) is the external part of a lava tube in Lanzarote (pictures: R.S. Azzoni).

ones and form cavities due to inflation (Pipan and Culver, 2019). Subsurface karst systems (including those developed in sandstone bedrock) are related to underground drainage system and their long-term evolution led to the development of deep (e.g., hundreds to thousands of meters) cave systems with a variety of features (galleries, wells, halls), whereas lava tubes generally form very close to the surface (Figure 3), displaying elongated features of regular diameter.

If we consider the main topic of this contribution, we shall start from the consideration of White and Culver (2019), suggesting that from the biologist's point of view, caves are openings that can accommodate cave-adapted organisms. Cave environments on Earth provide a valuable analog for Martian subsurface habitats, as they share key geological, geochemical and microclimatic characteristics that could support microbial life. Martian caves, particularly lava tubes, have been identified as high-priority targets for astrobiological exploration due to their natural protection from surface radiation, stable thermal conditions and potential for water ice preservation (Léveillé and Datta, 2010; Blank *et al.*, 2018; Sauro *et al.*, 2020).

Terrestrial and Martian caves exhibit similar formation processes and mineralogical compositions, making them excellent comparative systems for astrobiological investigations (Figure 1). While Earth hosts a diverse range of caves, formed by the dissolution of limestone, solutional weathering on silicates and volcanic lava tubes, Mars is predominantly characterized by basaltic lava tubes, which have been detected in multiple regions using orbital imaging (Martínez and Renno, 2013; Ojha *et al.*, 2015; Sauro *et al.*, 2020). However, some evidence suggests that geological substrate that can suffer dissolution or solutional weathering could also exist on Mars, thus confirming the potential presence of caves. This hypothetical occurrence of underground karst-related landforms on Mars is confirmed by the identification of soluble evaporitic rocks such as gypsum and halite, and signs of past flowing water on the Martian surface (Hajna *et al.*, 2017; Baioni, 2018). Furthermore, the Mars Global Cave Candidate Catalog (MGC3) has identified potential cave entrances associated with deep fractures and atypical pit craters (APCs) in regions exhibiting karst-similar characteristics (Cushing *et al.*, 2015).

On Earth, the activity of microbial communities is often a co-factor promoting the occurrence of (bio-)chemical weathering of rocks, likely enhancing the effects of dissolution and hydrolysis (Viles, 1984; Berthelin, 1988; Barker *et al.*, 1997; Bin *et al.*, 2008; Zhao *et al.*, 2024). Microbial communities are thus involved in karst corrosion and silicates solutional contributing to the formation of caves. At the same time, microbial communities occupy all parts of cave systems from their atrial parts, where photosynthetic organisms can survive although the limited penetration of light, up to the inner parts of

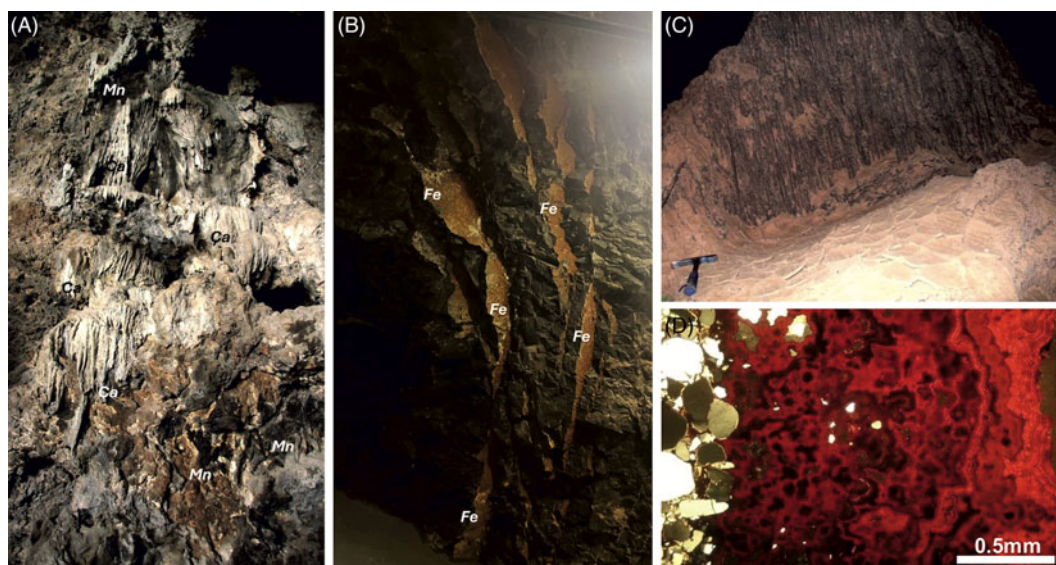


Figure 4. Examples of bio-mineralization found along caves on Earth. **A)** Calcitic (Ca) and Mn-bearing (Mn) mineralization along a wall of a cave in the Kurdistan Region of Iraq. **B)** Fe-bearing (Fe) mineralization along the wall of the Al Hoota Cave. **C)** Fe-bearing speleothem, likely formed after biomineralization in the underground network of the Tadrart Acacus Massif (SW Libya). **D)** Microphotograph of C) illustrating the stromatolitic structure of Fe-bearing cave concretions (Zerboni *et al.*, 2015).

galleries and deepest abysses. Therein, subsurface microbial communities primarily rely on organic carbon inputs from surface-derived sources, with the majority predicted to be organoheterotrophs. However, many can also derive energy from chemolithotrophic processes, including nitrification (ammonia and nitrite oxidation), sulfur oxidation and, in some cases, manganese and iron oxidation (Sarbu *et al.*, 1996; Chen *et al.*, 2009; Tetu *et al.*, 2013; Ortiz *et al.*, 2014; Carmichael and Bräuer, 2015).

Biomineralizations resulting from the activities of chemolithotrophic communities are commonly found along the walls of caves on Earth (Forti, 2001; Northup *et al.*, 2001; Boston *et al.*, 2004; Miller *et al.*, 2013) and includes the deposition of Fe- and/or Mn-bearing coatings and nodules, consisting of mixtures of iron and manganese oxides and hydroxides (Figs. 4, 5). Microbial communities also contribute to the growth of calcitic biocrusts and occasionally speleothems (Figure 4). Biomineralization of iron and manganese have been described with a plethora of shapes, form micrometric staining up to more complex features including thin laminate layers of oxides and clays resembling stromatolites. Figs. 4, 6 illustrate an example of laminate biomineralization alternating clay rich layers to layers of iron oxides (hematite) from the solutional underground network of the central Sahara. Greenish photosynthetic biofilms and other kind of microbial mat can also appear in the twilight zone (Figure 5) on the walls and roof of the atrial parts of caves (Villa *et al.*, 2022) or in the deeper and aphotic parts of underground systems as consequence of human contamination or positioning of artificial lights (Northup *et al.*, 2011; Saiz-Jimenez, 2012; Zerboni *et al.*, 2022). The existence of inactive disappearances of microbial communities is occasionally suggested by the occurrence of other mineralization, such as Ca-oxalates efflorescence (Figure 5), that can represent the byproduct of metabolic activities (Gallinaro and Zerboni, 2021). Additionally, numerous cave microbes have been shown to utilize atmospheric trace gases such as H₂, CO and CH₄ as alternative energy sources (Waring *et al.*, 2017; Nguyễn-Thuỳ *et al.*, 2017; Cheng *et al.*, 2021; Bay *et al.*, 2024). Some chemoautotrophs are hypothesized to fix CO₂ using H₂ as an energy source, while aerobic methanotrophs metabolize CH₄ as both an energy and carbon source (Waring *et al.*, 2017; Cheng *et al.*, 2021; Bay *et al.*, 2024).

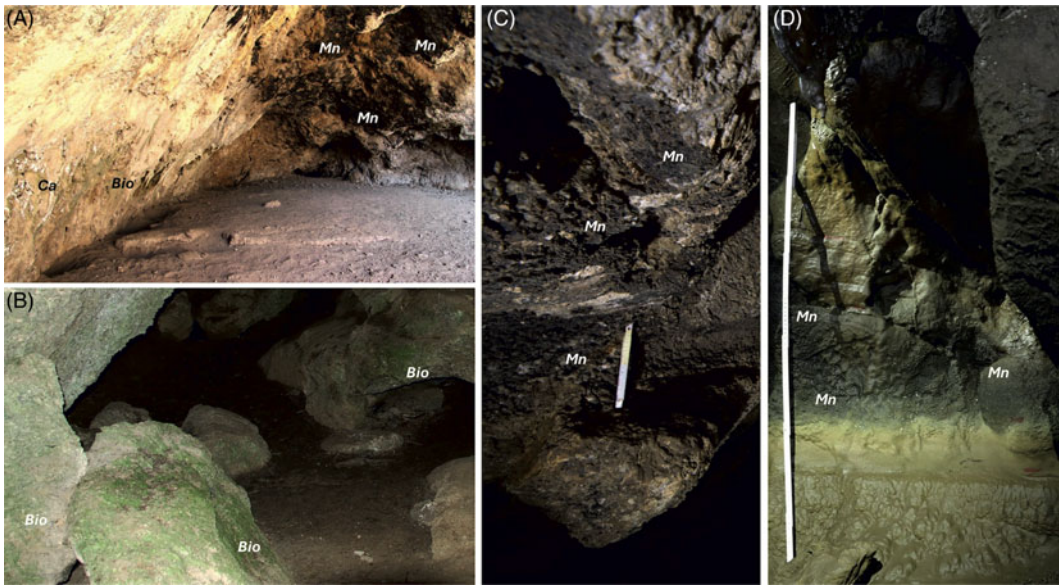


Figure 5. Examples of biofilms found along caves on Earth. A) Cave entrance in the Zagros Mt. showing Mn-bearing (Mn) and photosynthetic (Bio) biofilms; (Ca) indicates the presence of oxalates effloresces, likely related to epiliths. B) Photosynthetic (Bio) biofilms in the Tana della Mussina Cave (picture: M. Cremaschi). C) Mn-bearing (Mn) biofilms in the Tana della Mussina Cave (picture: M. Cremaschi). D) Mn-bearing (Mn) biofilms in a cave from the central Apennines of Italy (picture: L. Forti).

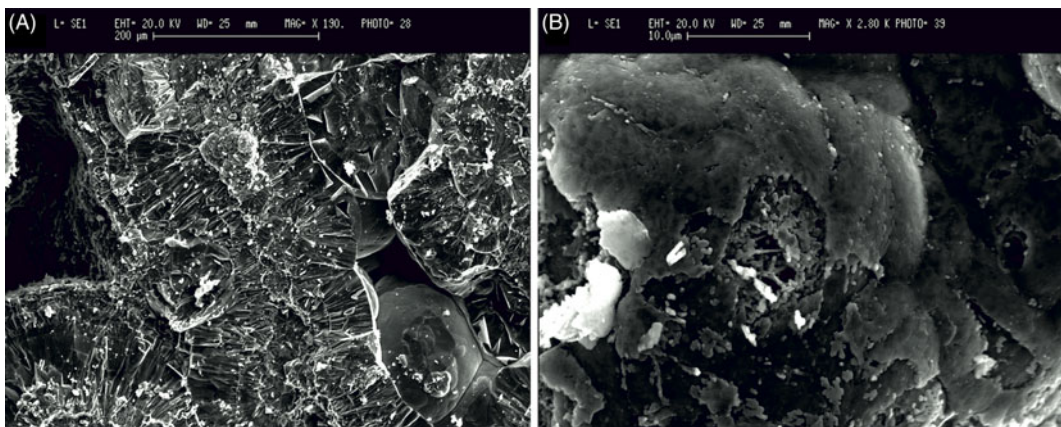


Figure 6. Scanning electron micrographs of Fe-bearing speleotem of Figure 4. A) Stromatolitic structure of the mineralization. B) Potential fossil extracellular iron oxide biomineral structures.

These metabolic pathways are particularly relevant for understanding potential Martian ecosystems, given that Mars exhibits substantial iron oxide and sulfate deposits, as well as potential trace gases in its atmosphere that could sustain analogous microbial processes (Boston *et al.*, 2004; Northup *et al.*, 2011). Additionally, studies of hydrothermal and sulfur-rich caves suggest that analogous environments may have existed on ancient Mars, where volcanic and aqueous interactions likely shaped habitable niches (D'Auria *et al.*, 2018; Tomczyk-Żak *et al.*, 2016).

One key factor influencing habitability is water availability. Earth's caves are often fed by percolating water or containing groundwater (karst systems), drip pools or ice deposits, in the case of lava tubes, they are close to the surface and can exchange humidity from the atmosphere or occasionally flooded. On the contrary, Martian caves may harbor subsurface water ice or transient briny solutions, particularly in polar and mid-latitude regions (Martínez and Renno, 2013). If liquid water or high-humidity microenvironments persist within Martian lava tubes, they could sustain microbial life, much like their terrestrial counterparts.

In terms of astrobiological modeling, terrestrial karst caves and lava tubes offer two distinct frameworks for understanding potential Martian habitats. Karst caves are formed by the chemical weathering of rocks such as limestone, dolomite and evaporites through interactions with acidic water (enriched in CO₂ coming from the atmosphere or produced by soil metabolism). These systems typically exhibit high microbial diversity, mineral deposits and redox gradients, which make them excellent environments for biosignature studies. However, since carbonate-based cave systems (limestone caves) have not been identified on Mars, they serve primarily as biological analogs rather than geological ones (Northup *et al.*, 2011). In contrast, lava tubes, which form when the surface of a lava flow cools while the interior remains molten, eventually leaving behind hollow subsurface tunnels, are known to exist on Mars (Léveillé *et al.*, 2010; Sauro *et al.*, 2020; Qiu *et al.*, 2023). These structures provide highly stable microclimates, shielding against cosmic and solar radiation and could serve as potential reservoirs for ice preservation, making them prime targets for astrobiological exploration. This makes them more relevant as Martian analogs (Léveillé and Datta, 2010; Sauro *et al.*, 2020). For instance, microbial mat composed of different microorganisms, including cyanobacteria, and associated biomineralizations (e.g., kerolite and aragonite) cover the Maniniholo Dry Cave on Kauai (Hawai'i; Léveillé *et al.*, 2000; Sauro *et al.*, 2020). Furthermore, the atmospheric conditions inside Martian caves represent an interesting astrobiological aspect to investigate. While the Martian atmosphere is extremely rarefied (with a surface pressure less than 1% of that of Earth) the deepest cave zones or sealed chambers could potentially maintain different atmospheric compositions and pressure, spawning favorable conditions for microbial-like life forms specialized in gas (i.e., CO₂ and CH₄) exploitation and fixation, as reported in terrestrial subsurface habitats (Martin-Pozas *et al.*, 2022).

Several well-studied caves on Earth provide direct insights into potential Martian habitats, serving as astrobiological test sites for understanding microbial survival and biosignature formation. One notable example is Lechuguilla Cave in the United States, one of the world deepest caves in limestone. It is an oligotrophic environment with extremely low organic input from the surface, making it a valuable model for understanding how microbial life can persist in nutrient-poor subsurface environments (Northup *et al.*, 2011). Another significant example is Cueva de Villa Luz in Mexico, a sulfidic cave characterized by high concentrations of hydrogen sulfide and sulfuric acid (D'Auria *et al.*, 2018). The Rio Tinto caves in Spain offer yet another relevant model, as they are highly acidic, iron-rich environments that support extremophiles capable of iron oxidation and sulfate reduction. Given that the Martian surface is rich in iron oxides and apparently also of iron sulfates (Benison, 2016; Ehlmann, 2016), Rio Tinto provides insights into how similar metabolic pathways could sustain life in Martian caves, as much as analogous extreme environments for life on its surface (Papike, 2006; Amils *et al.*, 2011; Fernández-Remolar *et al.*, 2008).

Additionally, ice caves such as those in Antarctica represent another key terrestrial analog for Martian environments. These caves maintain stable subzero temperatures and host microbial life in ice and rock substrates, demonstrating how psychrophilic organisms can persist in extreme cold with minimal nutrients. Notably, the lava tube caves of Mt. Erebus in Antarctica provide an even more relevant comparison, as they combine both volcanic and cryogenic conditions, resembling potential subsurface habitats on Mars. These lava tubes harbor microbial communities adapted to thermal and geochemical gradients, with some microbes utilizing volcanic gases as energy sources, further reinforcing their astrobiological significance as analogs for putative Martian cave ecosystems, where ice deposits have been detected (Martínez *et al.*, 2014; Stibal *et al.*, 2012).

All these terrestrial analogues can be used as training grounds to develop the technologies required for the exploration of Martian caves. As recently demonstrated in a study carried out on lava tubes in Lanzarote (Domínguez *et al.*, 2025), the advancement of robotic technology is crucial. In this pilot experiment, a team of robots succeeded in generating a three-dimensional model of the area surrounding cave entrances and skylight, deploying a scout rover via rappelling and exploring the interior of the cave. The production of three-dimensional maps of Martian caves therefore represents a fundamental starting point, for which equipping rovers and robots with remote sensing instruments such as LiDAR is essential. However, this is only the beginning: it is also necessary to develop hardware and software technologies capable of detecting microorganisms in specific refugia within the caves, as well as identifying biomineralized traces of their past or present existence.

Microbial diversity in terrestrial caves

Microorganisms in terrestrial caves include bacteria, archaea, fungi and micro-eukaryotes, forming highly diverse and functionally complex communities that contribute to ecosystem stability and biogeochemical cycling (e.g. Martin-Sanchez *et al.*, 2014; Tomczyk-Żak *et al.*, 2016; Kosznik-Kwaśnicka *et al.*, 2022; Biagioli *et al.*, 2023, 2024; Bay *et al.*, 2024; Poli *et al.*, 2024). Fungal and bacterial communities in caves exhibit remarkable taxonomic and functional diversity. At the phylum level, cave microbial communities often reflect the diversity found in surface soils, with bacterial groups such as Proteobacteria, Actinobacteria, Acidobacteria and Firmicutes commonly detected (Weng *et al.*, 2022; Biagioli *et al.*, 2024; Hathaway *et al.*, 2024). Fungi represented by Ascomycota, Basidiomycota and Mortierellomycota are abundant in humid cave environments and play key roles in organic matter decomposition and symbiotic interactions with bacteria (Northup and Hathaway, 2012; Gonzalez-Henao and Schrenk, 2025). Additionally, Archaea domain constitutes an important fraction of terrestrial cave microbiomes, with phyla Thaumarchaeota and Euryarchaeota largely reported across different subterranean environments (Bontemps *et al.*, 2022; Biagioli *et al.*, 2023), contributing to various biogeochemical processes (Cheng *et al.*, 2023). Studies on microbial communities from Mount Erebus, an active volcano in Antarctica, have revealed a predominance of bacteria belonging to the phyla Proteobacteria and Firmicutes (Rasmussen *et al.*, 2024).

Microbial diversity also varies across different cave zones. The atrial entrance, which receives some light and organic input from the external environment, typically supports a more diverse microbial community, including phototrophic and heterotrophic organisms. In contrast, the twilight and deep zones, where darkness and oligotrophy prevail, host specialized microbial assemblages. While these deeper zones are characterized by low organic input, they can still support diverse microbial life through inorganic energy sources, including trace gas oxidizers and nitrifiers (Biagioli *et al.*, 2023; Bay *et al.*, 2024).

Comparative studies between different cave systems worldwide have shown that microbial diversity is not only shaped by abiotic factors but also by historical biogeographic patterns. Some caves harbor endemic microbial taxa, while others show similarities with microbial communities from distant but geochemically comparable environments (Gonzalez-Henao and Schrenk, 2025; Northup *et al.*, 2012). This spatial heterogeneity underscores the complexity of cave microbiomes and their potential for harboring novel microbial lineages with unique ecological roles. Despite the broad environmental, geological and geo-morphological heterogeneity of subterranean ecosystems, 52 bacterial and 50 fungal phylotypes were found dominant in caves worldwide. Mainly represented by genera *Nitrospira*, *Pseudomonas*, *Bacillus*, *Flavobacterium* and *Mortierella*, *Cladosporium*, *Pseudogymnoascus* and *Cephalotrichum*, their diversity patterns were largely driven by outdoor growing aridity conditions, selecting specific dominant taxa and constraining the total cave microbial diversity (Biagioli *et al.*, 2024). These findings may be particularly relevant in supporting the hypothesis that if primordial life forms had originated on Mars, they could have evolved specific adaptations to colonize underground

habitats. This adaptation might have occurred regardless of their biogeographic features, leading to the development of a shared microbial core similar to that observed in Earth's caves.

Microbial adaptations and survival strategies of caves microbes

Studies on karst underground systems and lava tube have demonstrated that microbial diversity is strongly influenced by factors such as nutrient availability, humidity, temperature and mineral composition (Biagioli *et al.*, 2023; L  veill   and Datta, 2010). Recent advances in systems biology and community-level ecology suggest that microbial consortia in caves may exhibit emergent properties, such as metabolic complementarity and cooperative stress responses. These interactions could be critical for sustaining life in oligotrophic environments and may resemble the dynamics of early microbial ecosystems in extraterrestrial settings (Bay *et al.*, 2024). Karst caves, primarily composed of calcium carbonate, typically exhibit more alkaline conditions, while volcanic lava tubes tend to be more acidic. These cave systems, shaped by distinct geological and climatic processes, impose different environmental filters such as pH, mineralogy, water availability and redox conditions that result in heterogeneous selection and variable successional trajectories of microbial communities. According to community assembly theory, these environmental pressures lead to differences in microbial composition and function across cave types, where deterministic processes such as niche differentiation and environmental filtering dominate over stochasticity. As such, karst and volcanic caves may host functionally distinct microbial consortia, with succession patterns shaped by the availability of electron donors and the stability of microclimatic niches (Costantin *et al.*, 2019).

Despite these differences, microbial communities in both environments have adapted to nutrient scarcity, darkness and fluctuating humidity, relying on organic carbon from the surface, trace gases, ammonia, nitrite and sulfide for survival (Luis-Vargas *et al.*, 2024; Bay *et al.*, 2024). These adaptations, including chemolithotrophy, biofilm formation and endolithic colonization, highlight the potential for microbial life in Martian caves, where similar constraints exist. Additionally, cave microbes exhibit resistance to desiccation, radiation and temperature variations, making them valuable analogs for extraterrestrial life (Boston *et al.*, 2004; Northup *et al.*, 2011; L  veill   and Datta, 2010). In the following parts, we briefly summarize the different strategies adopted by microbial communities to survive caves' environmental conditions (Figure 7).

Psychrophiles and halophiles in subsurface environments

One of the most significant adaptations observed in cave environments is the ability of microbes to survive in permanently cold and hypersaline conditions, which are directly relevant to Martian analog environments. In ice caves such as those found in Antarctica and Greenland, psychrophilic fungi, bacteria and archaea thrive in permanently frozen conditions, demonstrating metabolic activity at subzero temperatures (Connell and Staudigel, 2013; Tebo *et al.*, 2015; Kelly *et al.*, 2023). Many of these microbes rely on antifreeze proteins and cryoprotectants to prevent cellular damage caused by ice crystal formation. Likewise, halophilic microbes, such as those found in hypersaline cave lakes, can tolerate extreme osmotic stress by accumulating compatible solutes to maintain cellular integrity (Hallsworth *et al.*, 2021).

The relevance of low temperature resistance to the life in underground habitats was emphasized by the high frequency of isolates psychrophilic/psychrotolerant microbial strains, belonging to the bacterial genera *Pseudomonas*, *Arthrobacter* and *Bacillus* and to the fungal genera *Aureobasidium*, *Cladosporium* and *Pseudogymnoascus*, regardless of geographic and climatic location of caves surveyed (Burow *et al.*, 2019; Poli *et al.*, 2024; Rusakova *et al.*, 2024). However, the presence of some genera, particularly ubiquitous laboratories and environmental taxa such as *Pseudomonas* and *Bacillus*, may indicate potential contamination and should therefore be interpreted with caution.

Caves microbiomes








Adaptation strategies	Fungi	Bacteria	Archaea
Psychrotolerance 	<i>Aureobasidium</i> (Ascomycota) <i>Cladosporium</i> (Ascomycota) <i>Pseudogymnoascus</i> (Ascomycota)	<i>Arthrobacter</i> (Actinomycetota) <i>Flavobacterium</i> (Bacteroidota) <i>Pseudomonas</i> (Pseudomonadota)	<i>Thermoplasmata</i>
Halotolerance 	–	<i>Bacillus</i> (Firmicutes) <i>Nocardiopsis</i> (Actinomycetota) <i>Streptomyces</i> (Actinomycetota)	–
Radiation resistance 	<i>Acremonium</i> (Ascomycota) <i>Cladosporium</i> (Ascomycota) <i>Devriesia</i> (Ascomycota)	<i>Bacillus</i> (Firmicutes) <i>Deinococcus</i> (Deinococcota)	<i>Halobacterium</i> (Euryarchaeota)
Desiccation resistance 	<i>Acremonium</i> (Ascomycota) <i>Cladosporium</i> (Ascomycota) <i>Devriesia</i> (Ascomycota)	<i>Bacillus</i> (Firmicutes) <i>Deinococcus</i> (Deinococcota)	<i>Halobacterium</i> (Euryarchaeota)
Chemosynthesis 	–	<i>Dormibacterota</i> , <i>Eremiobacterota</i> , <i>Actinobacteriota</i> , <i>Chloroflexota</i> , <i>Armatimonadota</i>	<i>Methanobacterium</i> (Euryarchaeota)
Endolithism 	<i>Cladosporium</i> (Ascomycota) <i>Devriesia</i> (Ascomycota) <i>Exophiala</i> (Ascomycota)	<i>Dormibacterota</i> , <i>Eremiobacterota</i> , <i>Chloroflexota</i> , <i>Armatimonadota</i>	–
Limited nutrient resistance 	<i>Cephalotrichum</i> (Ascomycota) <i>Devriesia</i> (Ascomycota) <i>Mortierella</i> (Mucoromycota)	<i>Arthrobacter</i> (Actinomycetota) <i>Pseudomonas</i> (Pseudomonadota) <i>Streptomyces</i> (Actinomycetota)	–

Figure 7. Adaptation strategies of extremophilic microorganisms in subsurface and extreme environments. The table summarizes key adaptive strategies employed by fungi, bacteria and archaea. Microbial taxa are listed according to their respective domain and associated adaptation mechanisms.

These adaptations are particularly relevant for Mars, where subsurface brines have been detected, and cave environments may contain pockets of liquid water that intermittently become available for microbial metabolism (Ojha *et al.*, 2015).

Chemolithotrophic life and subsurface ecosystems

Chemolithotrophy is one of the most significant metabolic strategies employed by cave-dwelling microbes, allowing them to obtain energy from the oxidation of inorganic compounds rather than relying on sunlight or organic matter. In sulfidic and iron-rich caves, microbial communities commonly rely on sulfur oxidation, iron oxidation and manganese reduction to drive energy metabolism. For example, bacteria such as *Beggiatoa* and *Desulfobivrio* thrive in sulfur-rich caves like Cueva de Villa

Luz, where they metabolize hydrogen sulfide, producing biomineralized deposits (Northup *et al.*, 2001; Hedrich *et al.*, 2021). Similarly, Fe/Mn bacteria, including *Acidithiobacillus ferrooxidans* and *Leptospirillum spp.*, dominate acidic cave systems such as the Rio Tinto caves in Spain, where they use iron oxidation to generate energy under extreme pH conditions (Jones *et al.*, 2023; 2014; 2016; Martin-Pozas *et al.*, 2023; Kelly *et al.*, 2023). Carmichael *et al.* (2013) suggest that a unique consortia of Mn(II)-oxidizing bacteria dominates the environment of the predominantly epigenic cave systems of the upper Tennessee River Basin (USA) also promoting biomineralization processes and playing an important role in the formation of cave ferromanganese deposits.

Beyond sulfur, iron and manganese cycling, nitrogen-based chemolithotrophy also plays a significant role in cave ecosystems. Nitrifying bacteria, such as those described in metagenomic studies of microbial slime communities, have been shown to drive inorganic nitrogen metabolism in cave environments (Tetu *et al.*, 2013; Ortiz *et al.*, 2014). In addition, sulfide-oxidizing bacteria form the foundation of chemoautotrophic food webs in sulfidic caves, as demonstrated in the Movile Cave ecosystem (Sarbu *et al.*, 1996; Chen *et al.*, 2009; Nicolosi *et al.*, 2022). Methanotrophy represents yet another key metabolic pathway in subterranean systems. Cave-dwelling methanotrophs, such as those detected in limestone caves and tropical karst systems, contribute to methane depletion and atmospheric methane oxidation (Waring *et al.*, 2017; Nguyễn-Thùy *et al.*, 2017; Cheng *et al.*, 2021).

These diverse metabolic strategies underscore the remarkable adaptability of cave microbial communities to extreme environments. In particular, nitrifiers, sulfide oxidizers and methanotrophs not only sustain subsurface ecosystems in the absence of light but also provide valuable insights into analogous biogeochemical processes that may occur on other planetary bodies. Given that Mars hosts extensive iron oxide and sulfate deposits, along with potential atmospheric methane, these metabolic pathways serve as key models for identifying biosignatures and developing strategies for extraterrestrial life detection.

Endolithic communities

Endolithic organisms are microbes that inhabit the interior of rocks thriving in environments previously considered inhospitable to life. These organisms include a diverse array of life forms such as bacteria, fungi, algae, lichens and viruses (Friedmann, 1982; Archer *et al.*, 2017; Ettinger *et al.*, 2023; Coleine *et al.*, 2021, 2024). They have been discovered in various harsh settings, including the hyper-arid Atacama Desert and the McMurdo Dry Valleys in Antarctica. Their ability to survive in such extreme conditions has made them subjects of interest in astrobiology, as they may serve as analogs for potential life on other planets (Meslier and DiRuggiero, 2019).

Endolithic colonization is another survival strategy in extreme cave environments. These microbes have been documented in deep karst systems, lava tubes and even Antarctic ice caves, where they survive by accessing trace amounts of water and nutrients within the rock matrix (Wierzchos *et al.*, 2013; Coleine *et al.*, 2021; Rhind *et al.*, 2024). For example, melanized rock-inhabiting fungi belonging to the genera *Exophiala*, *Devriesia* and *Cladosporium*, were widely recorded in subterranean habitats across multiple substrates such as sediments, walls and speleothems and also recognized among the dominant taxa in caves worldwide (Liu *et al.*, 2022; Biagioli *et al.*, 2024). These microbes exhibit slow growth rates but high resilience to extreme environmental conditions (Ertekin *et al.*, 2021; Perez-Fernandez *et al.*, 2022). Recent studies have revealed that many of these endolithic microorganisms can oxidize trace atmospheric gases such as hydrogen (H₂) and carbon monoxide (CO) to obtain energy, a process known as atmospheric chemosynthesis (Greening and Grinter, 2022). This metabolic strategy enables them to survive in nutrient-poor environments by utilizing high-affinity enzymes to extract energy from these gases (Coleine *et al.*, 2024; Williams *et al.*, 2024). Their ability to survive in nutrient-poor, radiation-exposed habitats (Sajjad *et al.*, 2022; Coleine *et al.*, 2024; Williams *et al.*, 2024) makes them ideal analogs for potential microbial life in Martian subsurface environments, where endolithic niches could provide refuge from surface radiation and desiccation (McNamara *et al.*, 2006; Popović *et al.*, 2021).

Biofilms

Another crucial adaptation seen in cave microbiomes is the formation of biofilms and microbial mats, which allow microbial communities to adhere to mineral surfaces and collectively withstand environmental stresses (Banerjee *et al.*, 2013; Popović *et al.*, 2015; Figure 5). Biofilms create microenvironments that can retain moisture, concentrate nutrients and offer protection against desiccation, radiation and temperature fluctuations. In terrestrial caves, biofilms contribute to the weathering of rock surfaces, often leading to the precipitation of minerals that can serve as biosignatures over geological timescales (Pfendler *et al.*, 2018). The presence of similar biomineralized structures in Martian lava tubes could indicate past or present microbial activity, making biofilms a key target for future astrobiological missions.

Resistance to desiccation, radiation and limited nutrients

Microbial resistance to desiccation is a well-established survival strategy in cave environments. Although resistance to ionizing radiation is not typically selected for in Earth's caves, given their natural shielding, it is highly relevant when considering potential microbial life in Martian caves. Such resistance may be necessary in shallow or partially exposed systems or retained as an ancestral trait from more irradiated environments. Many microbes exhibit robust DNA repair mechanisms and protective cellular structures that enable them to endure prolonged periods of dryness and exposure to high-energy radiation. On Earth, extremotolerant and extremophilic organisms like *Deinococcus radiodurans*, known for their resistance to radiation, desiccation and other harsh conditions, exemplify the resilience of life in extreme environments. Studies show that desiccated and frozen cells of *D. radiodurans* can withstand radiation doses up to 140 kGy, suggesting potential survival strategies for microbes in Martian subsurface environments (Slade *et al.*, 2011; Sudharsan *et al.*, 2022; Liu *et al.*, 2023). Certain cryptoendolithic fungi such as *Cryomyces antarcticus* and *C. minteri*, found in Antarctic deserts, display high resistance to UV radiation and desiccation (Pacelli *et al.*, 2019; Gomez-Gutierrez *et al.*, 2024). Other extremophiles, like *Halobacterium salinarum*, survive intense UV exposure through efficient DNA repair and protective pigments (Baati *et al.*, 2022; Bouhamed *et al.*, 2024). Similarly, desiccation-resistant bacteria found in deep karst systems provide clues about microbial persistence in the dry subsurface of Mars (Coleine and Delgado-Baquerizo, 2022). These findings suggest that if microbial life exists on Mars, it may employ similar strategies to survive in subsurface caves, where water availability is intermittent, and radiation exposure is reduced.

In addition to desiccation and radiation tolerance, oligotrophy-resistance represents a crucial microbial adaptation for survival in nutrient-poor cave systems. Many cave-dwelling microbes have evolved metabolic flexibility, slow growth rates and efficient nutrient recycling mechanisms that allow them to persist under persistent energy limitations. For instance, microbial communities colonizing weathered rocks in subsurface karst caves exhibit syntrophic interactions and specialized metabolic pathways to exploit scarce resources (Wang *et al.*, 2022). Similarly, oligotrophic fungi isolated from carbonate caves, including newly described *Cephalotrichum* species, demonstrate the ability to thrive in carbon- and nitrogen-limited environments (Jiang *et al.*, 2017). These adaptations are highly relevant when considering the possibility of life in Martian caves, where both radiation protection and extreme oligotrophy would be defining constraints. The convergence of desiccation, radiation and oligotrophy-resistance strategies reinforces the potential for microbial survival in extraterrestrial subsurface niches.

Anthropogenic impacts such as tourism, artificial lighting and pollution have also been shown to alter microbial diversity, introducing opportunistic species and disrupting native microbial assemblages (Biagioli *et al.*, 2023; Hathaway *et al.*, 2024; Zerboni *et al.*, 2022). Understanding how microbial communities respond to these stressors is crucial, as human exploration of Mars could introduce Earth microbes that may compete with or obscure indigenous Martian life.

Preservation of biosignatures in caves

Caves provide an exceptional environment for the long-term preservation of biosignatures, acting as natural time capsules that protect microbial fossils from external degradation. The most suitable systems for biosignature preservation are fossil/inactive karst systems, where water flowing and percolation are limited or absent, and lava tubes, which display minimal interaction with the outer environment. Due to their microclimatic conditions, including minimal exposure to ultraviolet radiation, low rates of sedimentary disturbance and geochemically dynamic mineral surfaces, caves offer one of the most promising terrestrial environments for studying biosignature formation. On Mars, subsurface caves may serve a similar role by shielding potential organic material and microbial remnants from the harsh surface conditions, including cosmic radiation, oxidation and extreme temperature fluctuations (Dartnell *et al.*, 2007). Understanding the processes of biosignature preservation in Earth's caves is therefore critical for developing life-detection strategies in Martian lava tubes.

One of the primary mechanisms through which biosignatures are preserved in caves is mineral entrapment and biomineralization, particularly through the deposition of silica, sulfates and iron oxides. These mineral matrices can encapsulate microbial cells, biofilms and extracellular polymeric substances, creating fossilized structures that can persist for thousands to millions of years (Dubois *et al.*, 2014; Melim *et al.*, 2016). For instance, the stromatolite-like speleothems of Figure 4 likely formed in the Neogene, several million years ago (Zerboni *et al.*, 2015), but at the microscale still highlights the occurrence of features that can be interpreted as extracellular iron oxide biominerals like those described in different environmental contexts (Cockell *et al.*, 2011; Chiu *et al.*, 2017). Fe- and Mn-coating along cave walls entomb and shelter trace of biological activities as much as rock varnish do in open air environments (Zerboni, 2008). Silica precipitation, for example, has been shown to rapidly entomb microbial filaments and organic compounds, preserving fine-scale morphological and chemical features that can serve as indicators of past life. Similar silica-rich deposits have been identified in Martian surface sediments and hydrothermal regions, making silica an important target for biosignature detection (Brolly *et al.*, 2019). Sulfate deposits, particularly those associated with sulfidic caves and hydrothermal systems, provide another critical medium for microbial fossilization. Sulfur-metabolizing bacteria, such as those found in Cueva de Villa Luz, interact with dissolved sulfates and hydrogen sulfide to form gypsum and other sulfate minerals, effectively embedding microbial structures within the rock record (Jones *et al.*, 2023). On Mars, sulfate-rich outcrops identified by orbital spectrometers suggest that similar biomineralization processes may have occurred in ancient aqueous environments, providing potential analogs for preserved biosignatures (Antunes *et al.*, 2023).

Iron oxides and iron-rich sediments are also significant contributors to biosignature preservation in caves. In systems such as the Rio Tinto caves in Spain, microbial communities involved in iron oxidation generate extensive iron-rich biofilms, which can become lithified over time, preserving microbial textures and molecular biosignatures (Olías *et al.*, 2015; Nicholas *et al.*, 2022). Given that Mars' surface is dominated by iron oxides, it is plausible that microbial life, if it existed in Martian caves, could have left similar biogeochemical signatures in iron-rich deposits (Kelly *et al.*, 2023). The identification of hematite and goethite on Mars further supports the possibility that microbial iron oxidation may have played a role in the planet's early geochemical cycles (Chen *et al.*, 2021; Jiang *et al.*, 2022).

In the context of extinct life, microbial fossils and preserved biosignatures in Earth's caves serve as models for potential evidence of past life on Mars (Uckert *et al.*, 2017). Microbial filaments preserved in speleothems, iron oxide deposits and stromatolitic structures in terrestrial caves indicate that past microbial activity can leave long-lasting traces in mineral formations. The presence of silica-rich deposits in Martian caves, like those found in Yellowstone's hot spring-associated caves and hydrothermal systems (Brolly *et al.*, 2019; Çolak *et al.*, 2022), suggests that mineralized biosignatures could be preserved within Martian lava tubes and subsurface cavities.

Taken together, these examples illustrate how mineral interactions with microbial communities in caves create lasting records of biological activity. Future exploration efforts should prioritize the

investigation of silica, sulfate and iron-rich deposits within Martian lava tubes, as these materials have demonstrated a strong capacity for preserving microbial structures and molecular biosignatures over geological timescales. The integration of spectroscopic, microscopic and isotopic techniques in upcoming Mars missions will be essential for distinguishing abiotic mineral formations from potential biosignatures, ultimately guiding the search for extraterrestrial life.

Yet, in addition to morphological and mineralogical traces, recent developments in metagenomics, metabolomics and lipidomics offer promising avenues for identifying molecular biosignatures in low-biomass or highly mineralized environments (Azua-Bustos *et al.*, 2023). Integrating these -omics approaches into planetary instrumentation could greatly enhance our ability to detect subtle or cryptic forms of life in Martian subsurface habitats (Rutter *et al.*, 2020).

Future research and exploration strategies

Despite significant advances in understanding terrestrial cave microbiomes and their astrobiological implications, several critical knowledge gaps remain. One of the main unresolved questions concerns the true extent of microbial diversity within these ecosystems, particularly in oligotrophic environments where microbial life is thought to persist at the limits of habitability. Current studies have provided insights into chemolithotrophic metabolism, biofilm formation and endolithic colonization, yet a comprehensive characterization of microbial functional diversity, community interactions and long-term survival mechanisms in caves is still lacking. Additionally, while research has demonstrated that microbial life can thrive in extreme cave environments on Earth, it remains uncertain whether Martian caves possess comparable environmental conditions that would allow for the persistence of life. Factors such as microclimatic stability, humidity retention and subsurface geochemical gradients on Mars require further investigation to assess their potential for supporting microbial ecosystems.

Given that multiple lines of evidence suggest that the Martian surface underwent several warm cycles, characterized by increases and decreases in water availability and consequent phases of hydration of the surface and adjacent areas (Wordsworth *et al.*, 2021; Rapin *et al.*, 2023; Kite and Conway, 2024; Adams *et al.*, 2025), it is important to highlight the potentially special role of subsurface cavities. While overall increases in surface water availability may have acted as a booster for biological activity, at least in specific regions, the transition to hyperarid conditions must instead have imposed severe ecological constraints on life. By contrast, because subsurface cavities are environments more resilient to surface climatic fluctuations (Mammola *et al.*, 2019), they may also have functioned on Mars as ecological refugia or even as true oases, where the concentration and persistence of residual moisture could have enabled microbial communities to develop adaptive strategies. For this reason, efforts should focus on the potential to identify biological activity or its biomineralized traces in subaerial ‘oasis’ environments. Given the challenges of exploring subterranean environments on Mars – which is even more difficult than exploring its surface –, robotic missions for subsurface life detection are becoming a key focus of planetary exploration (Olsson-Francis *et al.*, 2023; Morrell *et al.*, 2024). Several recent and upcoming missions aim to develop the technology necessary to explore Martian caves and lava tubes (e.g., Domínguez *et al.*, 2025), providing crucial data on their habitability and potential for biosignature preservation (Pires *et al.*, 2023; Morrell *et al.*, 2024). The NASA BRAILLE (Biologic and Resource Analog Investigations in Low Light Environments) project has tested autonomous robotic systems in terrestrial caves, demonstrating the feasibility of machine learning-assisted exploration and biosignature detection in dark and complex environments (Blank, 2023). Similarly, ESA’s ExoMars rover, equipped with a subsurface drill, aims to search for organic molecules and potential biosignatures in the Martian regolith, paving the way for future cave exploration missions (Altieri *et al.*, 2023). While current technology is primarily designed for surface and near-subsurface investigations, future missions will need to incorporate advanced autonomous navigation, robotic mobility and subsurface sampling capabilities to access deeper cave systems on Mars.

Beyond navigation challenges, the detection and characterization of biosignatures within Martian caves will require sophisticated instrumentation capable of distinguishing biological materials from abiotic geochemical signatures. Traditional life-detection methods, such as gas chromatography-mass spectrometry (GC-MS) and Raman spectroscopy, have proven effective for organic molecule detection (Baque *et al.*, 2022; Rull *et al.*, 2022; Ansari *et al.*, 2023) but may not be sufficient for identifying subtle or fossilized biosignatures in mineral-rich cave environments. Novel analytical techniques, including amino acid chirality analysis, isotopic fractionation studies and in situ fluorescence-based biofilm detection, could enhance the ability of future missions to identify potential biosignatures in Martian lava tubes. Additionally, the development of drilling and sampling technologies that minimize contamination risks will be essential for ensuring the integrity of collected samples, particularly for future sample return missions. This point is of critical importance, as it is highly probable that Martian caves also preserve vestiges of a previous phase of biological colonization preserved as biomineralization. It is therefore necessary to expand current capabilities to distinguish between products of microbial metabolism and those of authigenic precipitation, starting from the study of terrestrial analogues.

In the coming decades, a multidisciplinary approach combining microbiology, geology, robotics and planetary science will be necessary to advance our understanding of cave environments as potential extraterrestrial habitats. Future Mars exploration missions should prioritize subsurface investigations, with a focus on detecting and characterizing biosignatures within Martian lava tubes and other protected subsurface structures (Çolak *et al.*, 2022; Wynne *et al.*, 2022). By leveraging insights gained from terrestrial cave research, we can refine exploration strategies and improve the likelihood of detecting signs of life beyond Earth. The integration of autonomous robotic exploration, high-resolution life-detection instruments and comparative studies of terrestrial analogs will be crucial in shaping the future of astrobiological investigations on Mars.

Concluding remarks

The study of terrestrial caves provides critical insights into the potential habitability of Martian subsurface environments, highlighting their role as natural refuges for microbial life and reservoirs for biosignatures. Research on cave microbiomes has demonstrated that extremophilic organisms can thrive in conditions characterized by nutrient scarcity, darkness and geochemical gradients, employing metabolic strategies such as chemolithotrophy, endolithic colonization and biofilm formation. These adaptations offer compelling analogs for potential life in Martian caves, where similar environmental conditions suggest that subsurface microbial ecosystems could exist. Furthermore, mineralogical processes observed in Earth's caves, including the biomineralization of silica, sulfates and iron oxides, provide key mechanisms for biosignature preservation, reinforcing the importance of cave environments in the search for past or present extraterrestrial life.

Given their astrobiological significance, Martian caves should be prioritized in life-detection strategies for future exploration missions. Unlike the planet's surface, where organic molecules are rapidly degraded by radiation and oxidative processes, subsurface environments offer greater protection and a higher likelihood of preserving biosignatures over geological timescales. By studying how microbial communities interact with mineral substrates in terrestrial caves, researchers can refine their approaches for detecting and interpreting potential biosignatures in Martian lava tubes.

The exploration of subsurface environments represents a new frontier in astrobiology, offering one of the most promising opportunities to answer the fundamental question of whether life exists or ever existed, beyond our planet. By leveraging the knowledge gained from terrestrial cave research, we can refine our strategies for planetary exploration, improving our ability to detect and interpret biosignatures on Mars and, ultimately, across the Solar System.

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References

- Adams D, Scheucher M, Hu R, Ehlmann BL, Thomas TB, Wordsworth R, Scheller E, Lillis R, Smith K, Rauer H and Yung YL (2025) Episodic warm climates on early Mars primed by crustal hydration. *Nature Geoscience* **18**(2), 133–139. <https://doi.org/10.1038/s41561-024-01626-8>
- Altieri F, Frigeri A, Lavagna M, Le Gall A, Nikiforov SY, Stoker C ... and Vago JL (2023) Investigating the Oxia Planum subsurface with the ExoMars rover and drill. *Advances in Space Research*, **71**(11), 4895–4903.
- Amils R, González-Toril E, Aguilera A, Rodríguez N, Fernández-Remolar D, Gómez F, ... and Sanz JL (2011). From Río Tinto to Mars: the terrestrial and extraterrestrial ecology of acidophiles. In *Advances in applied microbiology* (Vol. **77**, pp. 41–70). Academic Press.
- Antunes A, Lau Vetter MC, Flannery D and Li Y (2023) Mars analogs: environment, habitability and biodiversity. *Frontiers in Astronomy and Space Sciences*, **10**, 1208367.
- Ansari AH (2023) Detection of organic matter on Mars, results from various Mars missions, challenges, and future strategy: a review. *Frontiers in Astronomy and Space Sciences*, **10**, 1075052.
- Archer SD, De los Ríos A, Lee KC, Niederberger TS, Cary SC, Coyne KJ, ... and Pointing SB (2017) Endolithic microbial diversity in sandstone and granite from the McMurdo Dry Valleys, Antarctica. *Polar Biology*, **40**, 997–1006.
- Azua-Bustos A, Fairén AG, González-Silva C, Prieto-Ballesteros O, Carrizo D, Sánchez-García L, Parro V, Fernández-Martínez MÁ, Escudero C, Muñoz-Iglesias V, Fernández-Sampedro M, Molina A, García Villadangos M, Moreno-Paz M, Wierczos J, Ascaso C, Fornaro T, Brucato J R, Poggiali G, Manrique JA, Veneranda M, López-Reyes G, Sanz-Arranz A, Rull F, Ollila AM, Wiens RC, Reyes-Newell A, Clegg SM, Millan M, Johnson SS, McIntosh O, Szopa C, Freissinet C, Sekine Y, Fukushima K, Morida K, Inoue K, Sakuma H and Rampe E (2023) Dark microbiome and extremely low organics in Atacama fossil delta unveil Mars life detection limits. *Nature Communications*, **14**, 808. <https://doi.org/10.1038/s41467-023-36482-8>
- Baati H, Siala M, Azri C, Ammar E, Dunlap C and Trigui M (2022) Genomic analysis of heavy metal-resistant *Halobacterium salinarum* isolated from Sfax solar saltern sediments. *Extremophiles*, **26**(2), 25.
- Baioni D (2018) Karst landforms as markers of recent climate change on Mars: An example from a Late Amazonian Epoch evaporate-karst within a trough in western Noctis Labyrinthus. In *Dynamic Mars* (pp. 411–429). Elsevier.
- Baqué M, Backhaus T, Meeßen J, Hanke F, Böttger U, Ramkissoon N, ... and de Vera JPP (2022) Biosignature stability in space enables their use for life detection on Mars. *Science advances* **8**(36), eabn7412.
- Banerjee S and Joshi SR (2013) Insights into cave architecture and the role of bacterial biofilm. *Proceedings of the National Academy of Sciences, India Section B: Biological Sciences*, **83**, 277–290.
- Barker WW, Welch SA, Banfield JF (1997) Biogeochemical weathering of silicate minerals. In Banfield JF, Nealson KH (eds), *Geomicrobiology: Interactions between Microbes and Minerals*. De Gruyter, Berlin, pp. 391–428.
- Bay SK, Ni G, Lappan R, Leung PM, Wong WW, Holland S, ... and Greening C (2024) Microbial aerotrophy enables continuous primary production in diverse cave ecosystems. *bioRxiv*, 2024–05.
- Benison KC (2016) Alunite on Mars. *American Mineralogist* **101**, 1499–1500.
- Berthelin J (1988) Microbial Weathering Processes in Natural Environments. In Lerman A, Meybeck M (eds), *Physical and Chemical Weathering in Geochemical Cycles*. NATO ASI Series, 251, Springer, Dordrecht.
- Biagioli F, Coleine C, Piano E, Nicolosi G, Poli A, Prigione V, ... and Selbmann L (2023) Microbial diversity and proxy species for human impact in Italian karst caves. *Scientific Reports*, **13**(1), 689.
- Biagioli F, Coleine C, Delgado-Baquerizo M, Feng Y, Saiz-Jimenez C and Selbmann L (2024) Outdoor climate drives diversity patterns of dominant microbial taxa in caves worldwide. *Science of the Total Environment*, **9**
- Bin L, Ye C, Lijun Z, Ruidong Y (2008) Effect of microbial weathering on carbonate rocks. *Earth Science Frontiers* **15**, 90–99.
- Blank JG, Roush TL, Stoker CL, Colaprete A, Datta S, Wong U and Wynne JJ (2018) Planetary caves as astrobiology targets. *A white paper submitted to the Space Studies Board of the National Academy of Sciences*, 5
- Blank JG (2023) Highlights of the NASA BRAILLE Project: A Mars Analog Astrobiology Study Focused on Volcanic Caves at Lava Beds National Monument (N. California USA). *LPI Contributions*, 2697, 1072.
- Bontemps Z, Alonso L, Pommier T, Hugoni M and Moënné-Loccoz Y (2022) Microbial ecology of tourist Paleolithic caves. *Science of the total environment* **816**, 151492.
- Boston PJ, Spilde MN, Northup DE, Melin LA, Soroka DS, Kleina LG, Lavoie KH, Hose LD, Mallory LM, Dahm CM, Crossey LJ, Schelble RT (2004). Cave biosignature suites: microbes, minerals, and Mars. *Astrobiology* **1**, 25–55.

- Bouhamed SBH, Chaari M, Baati H, Zouari S and Ammar E (2024) Extreme halophilic Archaea: *Halobacterium salinarum* carotenoids characterization and antioxidant properties. *Heliyon* **10**(17).
- Brolly C, Parnell J and Bowden S (2019) Surface mineral crusts: a potential strategy for sampling for evidence of life on Mars. *International Journal of Astrobiology* **18**(2), 91–101.
- Burow K, Grawunder A, Harpke M, Pietschmann S, Ehrhardt R, Wagner L, ... and Kothe E (2019) Microbiomes in an acidic rock–water cave system. *FEMS Microbiology Letters* **366**(13), fnz167.
- Carmichael AK, Bräuer SL (2015) Microbial Diversity and Manganese Cycling: A Review of Manganese-oxidizing Microbial Cave Communities. In Summers Engel A (eds), *Microbial Life of Cave Systems*. De Gruyter, Berlin, 137–160.
- Carmichael MJ, Carmichael SK, Santelli CM, Strom A, Bräuer SL (2013) Mn(II)-oxidizing bacteria are abundant and environmentally relevant members of ferromanganese deposits in caves of the Upper Tennessee River Basin. *Geomicrobiology Journal* **30**, 779–800.
- Chen Y *et al* (2009) Life without light: microbial diversity and evidence of sulfur- and ammonium-based chemolithotrophy in Movile Cave. *ISME Journal* **3**, 1093–1104.
- Chen SA, Heaney PJ, Post JE, Fischer TB, Eng PJ and Stubbs JE (2021) Superhydrous hematite and goethite: a potential water reservoir in the red dust of Mars?. *Geology* **49**(11), 1343–1347.
- Cheng X-Y *et al* (2021) USC γ dominated community composition and cooccurrence network of methanotrophs and bacteria in subterranean Karst Caves. *Microbiol Spectr* **9**.
- Cheng X, Xiang X, Yun Y, Wang W, Wang H and Bodelier PL (2023) Archaea and their interactions with bacteria in a karst ecosystem. *Frontiers in Microbiology* **14**, 1068595.
- Chiu BK, Kato S, McAllister SM, Field EK, Chan CS (2017) Novel pelagic iron-oxidizing zetaproteobacteria from the Chesapeake Bay oxic–anoxic transition zone. *Frontiers in Microbiology* **18**, 1280.
- Cockell CS, Pybus D, Olsson-Francis K, Kelly L, Petley D, Rosser N, Howard K, Mosselmans F (2011) Molecular characterization and geological microenvironment of a microbial community inhabiting weathered receding shale cliffs. *Microb Ecol* **61**, 166–181.
- Çolak B and Güngör ND (2022) The astrobiological significance of caves on Earth and on Mars. *International Journal of Environment and Geoinformatics* **9**(3), 57–64.
- Coleine C, Biagioli F, de Vera JP, Onofri S and Selbmann L (2021) Endolithic microbial composition in Helliwell Hills, a newly investigated Mars-like area in Antarctica. *Environmental Microbiology* **23**(7), 4002–4016.
- Coleine C and Delgado-Baquerizo M (2022) Unearthing terrestrial extreme microbiomes for searching terrestrial-like life in the Solar System. *Trends in Microbiology* **30**(11), 1101–1115.
- Coleine C, Albanese D, Ray AE, Delgado-Baquerizo M, Stajich JE, Williams TJ ... and Selbmann L (2024) Metagenomics untangles potential adaptations of Antarctic endolithic bacteria at the fringe of habitability. *Science of the Total Environment* **917**, 170290.
- Connell L and Staudigel H (2013) Fungal diversity in a dark oligotrophic volcanic ecosystem (DOVE) on Mount Erebus, Antarctica. *Biology* **2**(2), 798–809.
- Constantin S, Toulkeridis T, Moldovan OT, Villacís M and Addison A (2019) Caves and karst of Ecuador—state-of-the-art and research perspectives. *Physical Geography* **40**(1), 28–51.
- Cushing GE, Okubo CH and Titus TN (2015) Atypical pit craters on Mars: new insights from THEMIS, CTX, and HiRISE observations. *Journal of Geophysical Research: Planets* **120**(6), 1023–1043.
- Dartnell LR, Desorger L, Ward JM and Coates AJ (2007) Martian sub-surface ionising radiation: biosignatures and geology. *Biogeosciences* **4**(4), 545–558.
- D'Auria G, Artacho A, Rojas RA, Bautista JS, Méndez R, Gamboa MT, ... and Gómez-Cruz R. (2018). Metagenomics of bacterial diversity in Villa Luz caves with sulfur water springs. *Genes* **9**(1), 55.
- Domínguez R, Pérez-del-Pulgar C, Paz-Delgado GJ, Polisanò F, Babel J, Germa T, Dragomir I, Carlietti V, Berthet A-C, Danté LC and Kirchner F (2025) Cooperative robotic exploration of a planetary skylight surface and lava cave. *Science Robotics* **10**(89), eadj6999. <https://doi.org/10.1126/scirobotics.adj6999>
- Dubois C, Quinif Y, Baele JM, Barriquand L, Bini A, Bruxelles L, ... and Vergari A (2014) The process of ghost-rock karstification and its role in the formation of cave systems. *Earth-Science Reviews*, **131**, 116–148.
- Ehlmann BL, Swayze GA, Milliken RE, Mustard JF, Clark RN, Murchie SL, Breit GN, Wray JJ, Gondet B, Poulet F, Carter J, Calvin WM, Benzel WM, Seelos KD (2016) Discovery of alunite in cross crater, terra sirenum, mars: evidence for acidic, sulfurous waters. *American Mineralogist* **101**, 1527–1542.
- Ertekin E, Meslier V, Browning A, Treadgold J and DiRuggiero J (2021) Rock structure drives the taxonomic and functional diversity of endolithic microbial communities in extreme environments. *Environmental Microbiology* **23**(7), 3937–3956.
- Ettinger CL, Saunders M, Selbmann L, Delgado-Baquerizo M, Donati C, Albanese D ... and Coleine C (2023). Highly diverse and unknown viruses may enhance Antarctic endoliths' adaptability. *Microbiome* **11**(1), 103.
- Gomez-Gutierrez SV, Sic-Hernandez WR, Haridas S, LaButti K, Eichenberger J, Kaur N ... and Grigoriev IV (2024). Comparative genomics of the extremophile *Cryomyces antarcticus* and other psychrophilic Dothideomycetes. *Frontiers in Fungal Biology* **5**, 1418145.
- Fernández-Remolar DC and Knoll AH (2008) Fossilization potential of iron-bearing minerals in acidic environments of Rio Tinto, Spain: Implications for Mars exploration. *Icarus* **194**(1), 72–85.
- Forti P (2001) Biogenic speleothems: an overview. *Int. J. Speleol.* **30**, 39–56.
- Friedmann EI (1982) Endolithic microorganisms in the Antarctic cold desert. *Science* **215**(4536), 1045–1053.

- Gallinaro M, Zerboni A (2021) Rock, pigments, and weathering. A preliminary assessment of the challenges and potential of physical and biochemical studies on rock art from southern Ethiopia. *Quaternary International* **572**, 106–119.
- Gomez-Gutierrez SV, Sic-Hernandez WR, Haridas S, LaButti K, Eichenberger J, Kaur N, ... and Grigoriev IV (2024) Comparative genomics of the extremophile *Cryomyces antarcticus* and other psychrophilic Dothideomycetes. *Frontiers in Fungal Biology* **5**, 1418145.
- Gonzalez-Henao S and Schrenk MO (2025) An astrobiological perspective on microbial biofilms: their importance for habitability and production of detectable and lasting biosignatures. *Applied and Environmental Microbiology* e01778–24.
- Greening C and Grinter R (2022) Microbial oxidation of atmospheric trace gases. *Nature Reviews Microbiology* **20**(9), 513–528.
- Hajna NZ, Baioni D and Tramontana M (2017) Karst landforms within Noctis Labyrinthus, Mars. *Acta Carsologica* **46**(1).
- Hays LE, Graham HV, Des Marais DJ, Hausrath EM, Horgan B, McCollom TM, ... and Lynch KL (2017) Biosignature preservation and detection in Mars analog environments. *Astrobiology* **17**(4), 363–400.
- Hallsworth JE, Koop T, Dallas TD, Zorzano MP, Burkhardt J, Golyshina OV, ... and McKay CP (2021) Water activity in Venus's uninhabitable clouds and other planetary atmospheres. *Nature Astronomy* **5**(7), 665–675.
- Hathaway JJM, Spilde MN, Northup DE (2024) *Looking for Microbial Biosignatures in All the Right Places: Clues for Identifying Extraterrestrial Life in Lava Tubes*. Applied Sciences.
- Hedrich S and Schippers A (2021) Distribution of acidophilic microorganisms in natural and man-made acidic environments. *Current issues in molecular biology* **40**(1), 25–48.
- Jiang JR, Cai L and Liu F (2017) Oligotrophic fungi from a carbonate cave, with three new species of *Cephalotrichum*. *Mycology* **8**(3), 164–177.
- Jiang Z, Liu Q, Roberts AP, Dekkers MJ, Barrón V, Torrent J and Li S (2022) The magnetic and color reflectance properties of hematite: from Earth to Mars. *Reviews of Geophysics* **60**(1), e2020RG000698.
- Jones DS, Schaperdorth I and Macalady JL (2014) Metagenomic evidence for sulfide oxidation in extremely acidic cave biofilms. *Geomicrobiology Journal* **31**(3), 194–204.
- Jones DS, Schaperdorth I and Macalady JL (2016) Biogeography of sulfur-oxidizing *Acidithiobacillus* populations in extremely acidic cave biofilms. *The ISME journal* **10**(12), 2879–2891.
- Jones WK, White WB (2019) Karst. In White BW, Culver DC, Pipan T (eds), *Encyclopedia of Caves*. Third Edition. Academic Press, 609–618.
- Jones D. S, Schaperdorth I, Northup DE, Gómez-Cruz R and Macalady JL (2023) Convergent community assembly among globally separated acidic cave biofilms. *Applied and environmental microbiology* **89**(1), e01575–22.
- Jurado V, Northup DE and Saiz-Jimenez C (2024) Microbial roles in caves. *Frontiers in Microbiology* **15**, 1411535.
- Kelly LC, Rivett DW, Pakostova E, Creer S, Cotterell T and Johnson DB (2023) Mineralogy affects prokaryotic community composition in an acidic metal mine. *Microbiological Research* **266**, 127257.
- Kite ES and Conway S (2024) Geological evidence for multiple climate transitions on Early Mars. *Nat. Geosci.* **17**, 10–19. <https://doi.org/10.1038/s41561-023-01349-2>
- Kosznik-Kwaśnicka K, Golec P, Jaroszewicz W, Lubomska D and Piechowicz L (2022) Into the unknown: microbial communities in caves, their role, and potential use. *Microorganisms* **10**(2), 222.
- Lauritzen SE (2018) Physiography of the Caves. In Moldovan O, Kováč L, Halse S (eds), *Cave Ecology. Ecological Studies*, vol 235. Springer, Cham. https://doi.org/10.1007/978-3-319-98852-8_2
- Léveillé RJ and Datta S (2010) Lava tubes and basaltic caves as astrobiological targets on Earth and Mars: a review. *Planetary and Space Science* **58**(4), 592–598.
- Léveillé RJ, Fyfe WS, Longstaffe FJ (2000) Geomicrobiology of carbonate-silicate microbialites from Hawaiian basaltic sea caves. *Chemical Geology* **169**, 339.
- Liu F, Li N, & Zhang Y (2023). The radioresistant and survival mechanisms of *Deinococcus radiodurans*. *Radiation Medicine and Protection*, **4**(02), 70–79.
- Mammola S, Piano E, Cardoso P, Vernon P, Domínguez-Villar D, Culver D. C, Pipan T, & Isaia M (2019). Climate change going deep: the effects of global climatic alterations on cave ecosystems. *The Anthropocene Review* **6**(1-2), 98–116. <https://doi.org/10.1177/2053019619851594>
- Martin-Pozas T, Cuezva S, Fernandez-Cortes A, Cañaveras JC, Benavente D, Jurado V, ... and Sanchez-Moral S (2022). Role of subterranean microbiota in the carbon cycle and greenhouse gas dynamics. *Science of the Total Environment* **831**, 154921.
- Martin-Sanchez P. M, Jurado V, Porca E, Bastian F, Lacanette D, Alabouvette C and Saiz-Jimenez C (2014) Airborne microorganisms in Lascaux cave (France).
- Martínez G and Renno NO (2013) Water and brines on Mars: current evidence and implications for MSL. *Space Science Reviews* **175**, 29–51.
- McNamara CJ, Perry TD, Bearce KA, Hernandez-Duque G and Mitchell R (2006). Epilithic and endolithic bacterial communities in limestone from a Maya archaeological site. *Microbial Ecology* **51**, 51–64.
- Melim LA, Northup DE, Boston PJ and Spilde MN (2016) Preservation of fossil microbes and biofilm in cave pool carbonates and comparison to other microbial carbonate environments. *Palaaios* **31**(4), 177–189.
- Meslier V and DiRuggiero J (2019) Endolithic microbial communities as model systems for ecology and astrobiology. In *Model ecosystems in extreme environments* (pp. 145–168). Academic Press.
- Miller AZ, Dionísio A, Jurado V, Cuezva S, Sanchez-Moral S, Cañaveras JC, Saiz-Jimenez C (2013) Biomineralization by cave dwelling microorganisms. In Sanjurjo Sanchéz J (ed), *Advances in Geochemistry Research*. Nova Science Publishers, 77–105.

- Morrell BJ, da Silva MS, Kaufmann M, Moon S, Kim T, Lei X, ... and Blank JG (2024). Robotic exploration of Martian caves: evaluating operational concepts through analog experiments in lava tubes. *Acta Astronautica* **223**, 741–758.
- Nayeli Luis-Vargas M, Webb J, White S and Bay SK (2024) Linking surface and subsurface: the biogeochemical basis of cave microbial ecosystem services. *Journal of Sustainable Agriculture and Environment* **3**(4), e70031.
- Nguyễn-Thủy D *et al* (2017) Subterranean microbial oxidation of atmospheric methane in cavernous tropical karst. *Chem Geol* **466**, 229–238.
- Nicholas L, & Thomas S (2022). Case Study: Rio Tinto destruction of Juukan Gorge cave system, Western Australia. In *Rural Victims of Crime* (pp. 189–191). Routledge.
- Nicolosi G, Galdenzi S, Messina MA, Miller AZ, Petralia S, Sarbu SM and Isaia M (2022) Sulfidic habitats in the gypsum karst system of Monte Conca (Italy) host a chemoautotrophically supported invertebrate community. *International journal of environmental research and public health* **19**(5), 2671.
- Northup DE (2011) Managing Microbial Communities in Caves. In van Beynen P (eds), *Karst Management*. Springer, Dordrecht. https://doi.org/10.1007/978-94-007-1207-2_10
- Northup DE, Lavoie KH (2001) Geomicrobiology of caves: a review. *Geomicrobiology journal* **18**(3), 199–222.
- Northup DE, Melim LA, Spilde MN, Hathaway JJM, Garcia MG, Moya M, ... and Riquelme C (2011). Lava cave microbial communities within mats and secondary mineral deposits: implications for life detection on other planets. *Astrobiology* **11**(7), 601–618.
- Northup DE, Hathaway JJM, Snider JR *et al.* (2012). Life in Earth's lava caves: Implications for life detection on other planets. *Life on Earth and Other Planets*, Springer.
- Ojha L, Wilhelm MB, Murchie SL, McEwen AS, Wray JJ, Hanley J, ... and Chojnacki M (2015) Spectral evidence for hydrated salts in recurring slope lineae on Mars. *Nature Geoscience* **8**(11), 829–832.
- Olías M and Nieto JM (2015) Background conditions and mining pollution throughout history in the Río Tinto (SW Spain). *Environments* **2**(3), 295–316.
- Olsson-Francis K, Doran PT, Ilyin V, Raulin F, Rettberg P, Kminek G, ... and Zaitsev M (2023) The COSPAR planetary protection policy for robotic missions to Mars: a review of current scientific knowledge and future perspectives. *Life Sciences in Space Research* **36**, 27–35.
- Ortiz M, Legatzki A, Neilson JW, Fryslye B, Nelson WM, Wing RA, ... and Maier RM (2014) Making a living while starving in the dark: metagenomic insights into the energy dynamics of a carbonate cave. *The ISME Journal* **8**(2), 478–491.
- Pacelli C, Selbmann L, Zucconi L, Coleine C, de Vera JP, Rabbow E, ... and Onofri S (2019). Responses of the black fungus *Cryomyces antarcticus* to simulated mars and space conditions on rock analogs. *Astrobiology* **19**(2), 209–220.
- Papike JJ, Kanner JM, Spilde MN, Shearer CK (2006) Terrestrial analogs of martian sulfates: Major and minor element systematics of alunite–jarosite from Goldfield, Nevada. *American Mineralogist* **91**, 1197–1200.
- Perez-Fernandez CA, Wilburn P, Davila A and DiRuggiero J (2022) Adaptations of endolithic communities to abrupt environmental changes in a hyper-arid desert. *Scientific Reports* **12**(1), 20022.
- Pfendler S, Karimi B, Maron PA, Ciadamidaro L, Valot B, Bousta F, ... and Aleya L (2018) Biofilm biodiversity in French and Swiss show caves using the metabarcoding approach: First data. *Science of the Total Environment* **615**, 1207–1217.
- Pipan T, Culver DC (2019) Shallow subterranean habitats. In White BW, Culver DC, Pipan T (Eds), *Encyclopedia of Caves*. Third Edition. Academic Press, 896–908.
- Pires AC, Moura R, Nunes JC, Barcelos P, Caetano P, Quinteiro P, ... and Chaminé HI (2023) Azorean lava tube systems: a proposal for a new planetary analog site towards future lunar and martian exploration research. *International Multidisciplinary Scientific GeoConference: SGEM*, **23**(1.1), 533–540.
- Poli A, Zanellati A, Piano E, Biagioli F, Coleine C, Nicolosi G, ... and Varese GC (2024) Cultivable fungal diversity in two karstic caves in Italy: under-investigated habitats as source of putative novel taxa. *Scientific Reports* **14**(1), 4164.
- Popović S, Subakov Simić G, Stupar M, Unković N, Predojević D, Jovanović J and Ljaljević Grbić M (2015) Cyanobacteria, algae and microfungi present in biofilm from Božana Cave (Serbia). *International Journal of Speleology* **44**(2), 4.
- Popović S, Nikolić N, Predojević D, Jakovljević O, Rađa T, Anđelković A and Subakov Simić G (2021) Endolithic phototrophs: Examples from cave-like environments. *Kragujevac Journal of Science* **43**, 123–137.
- Preston LJ, Melim LA, Polyak VJ, Asmerom Y and Southam G (2014) Infrared spectroscopic biosignatures from Hidden Cave, New Mexico: possible applications for remote life detection. *Geomicrobiology Journal* **31**(10), 929–941.
- Qiu X and Ding C (2023) Radar observation of the lava tubes on the Moon and Mars. *Remote Sensing* **15**(11), 2850.
- Rapin W, Dromart G, Clark BC, Schieber J, Kite ES, Kah LC, Thompson LM, Gasnault O, Lasue J, Meslin P-Y, Gasda PJ and Lanza NL (2023) Sustained wet–dry cycling on early Mars. *Nature* **620**, 299–302. <https://doi.org/10.1038/s41586-023-06220-3>
- Rasmussen TB, Noell SE, Herbold CW, Dickie IA, Richards-Babbage R, Stott MB, ... and McDonald IR (2024) Geothermal ecosystems on Mt. Erebus, Antarctica, support diverse and taxonomically novel biota. *FEMS Microbiology Ecology* **100**(11), fiae128.
- Rhind T, Ronholm J, Berg B, Mann P, Applin D, Stromberg J, ... and Cloutis EA (2014) Gypsum-hosted endolithic communities of the Lake St. Martin impact structure, Manitoba, Canada: spectroscopic detectability and implications for Mars. *International Journal of Astrobiology* **13**(4), 366–377.
- Rull F, Veneranda M, Manrique-Martinez J. A, Sanz-Arranz A, Saiz J, Medina J, ... and Lopez-Reyes G (2022). Spectroscopic study of terrestrial analogues to support rover missions to Mars—A Raman-centred review. *Analytica Chimica Acta* **1209**, 339003.
- Rusakova DA, Sidorenko ML and Kim AV (2024) Characteristics of Psychrotolerant Pseudomonads Isolated from Organogenic Clay Deposits of the Mramornaya Cave (Primorskii Krai). *Microbiology* **93**(1), 92–103.

- Rutter L, Barker R, Bezdan D, Cope H, Costes SV, Degoricija L, ... and Muratani M (2020) A new era for space life science: international standards for space omics processing. *Patterns* **1**(9).
- Sajjad W, Ilahi N, Kang S, Bahadur A, Zada S and Iqbal A (2022) Endolithic microbes of rocks, their community, function and survival strategies. *International Biodeterioration & Biodegradation* **169**, 105387.
- Saiz-Jimenez C (2012) Microbiological and environmental issues in show caves. *World Journal Microbiol Biotechnol* **28**, 2453–2464.
- Sarbu SM, Kane TC and Kinkle BK (1996) A chemoautotrophically based cave ecosystem. *Science* **272**(5270), 1953–1955.
- Sauro F, Pozzobon R, Massironi M, De Berardinis P, Santagata T and De Waele J (2020) Lava tubes on Earth, Moon and Mars: a review on their size and morphology revealed by comparative planetology. *Earth-Science Reviews* **209**, 103288.
- Slade D and Radman M (2011) Oxidative stress resistance in *Deinococcus radiodurans*. *Microbiology and molecular biology reviews* **75**(1), 133–191.
- Stibal M, Wadham JL, Lis GP, Telling J, Pancost RD, Dubnick A, ... and Anesio AM (2012) Methanogenic potential of Arctic and Antarctic subglacial environments with contrasting organic carbon sources. *Global Change Biology* **18**(11), 3332–3345.
- Sudharsan M, Prasad R, Chakraborty A and Rajendrasozhan S (2022) Proteomic profiling of *Deinococcus radiodurans* with response to thioredoxin reductase inhibitor and ionizing radiation treatment. *Journal of Proteomics* **267**, 104697.
- Tebo BM, Davis RE, Anitori RP, Connell LB, Schiffman P and Staudigel H (2015) Microbial communities in dark oligotrophic volcanic ice cave ecosystems of Mt. Erebus, Antarctica. *Frontiers in microbiology*, **6**, 179.
- Tetu SG *et al.* (2013) Life in the dark: metagenomic evidence that a microbial slime community is driven by inorganic nitrogen metabolism. *ISME J* **7**, 1227–1236.
- Tomezyk-Żak K and Zielenkiewicz U (2016) Microbial diversity in caves. *Geomicrobiology Journal* **33**(1), 20–38.
- Tuttle MD and Stevenson DE (2011) Variation in the cave environment and its biological implications. In Tyburec J, Chengler J, Snow T, Geiselman C (eds), Bat conservation and management workshop - Arizona, Bat Conservation International, 19–35.
- Uckert K, Chanover NJ, Getty S, Voelz DG, Brinckerhoff WB, McMillan N, ... and Chavez A (2017) The characterization of biosignatures in caves using an instrument suite. *Astrobiology* **17**(12), 1203–1218.
- Viles HA (1984) Biokarst: review and prospect. *Progress in Physical Geography* **8**, 523–542.
- Villa F, Wu Ying-Li, Zerboni A, Cappitelli F (2022) In living color: Pigments-based microbial ecology at the mineral-air interface. *BioScience* **72**, 1156–1175.
- Wang Y, Cheng X, Wang H, Zhou J, Liu X and Tuovinen OH (2022) The characterization of microbiome and interactions on weathered rocks in a subsurface karst cave, Central China. *Frontiers in Microbiology* **13**, 909494.
- Waring CL *et al.* (2017) Seasonal total methane depletion in limestone caves. *Sci Rep* **7**, 1–12.
- Watson TH and Baldini JU (2024) Martian cave detection via machine learning coupled with visible light imagery. *Icarus* **411**, 115952.
- Weng M.M, Zaikova E, Millan M *et al.* (2022) Life underground: Investigating microbial communities and their biomarkers in Mars-analog lava tubes at Craters of the Moon National Monument and Preserve. *Journal of Geophysical Research: Planets*.
- White BW and Culver DC (2019) Definition of cave. In: White BW, Culver DC and Pipan T. (Eds.), *Encyclopedia of Caves*. Third Edition. Academic Press, 255–259. <https://doi.org/10.1016/B978-0-12-814124-3.00028-5>
- Wierzchos J, Davila A. F, Artieda O, Cámara-Gallego B, de los Ríos A, Neelson KH, ... and Ascaso C (2013) Ignimbrite as a substrate for endolithic life in the hyper-arid Atacama Desert: implications for the search for life on Mars. *Icarus* **224**(2), 334–346.
- Williams TJ, Allen MA, Ray AE, Benaud N, Chelliah DS, Albanese D, ... and Ferrari BC (2024) Novel endolithic bacteria of phylum Chloroflexota reveal a myriad of potential survival strategies in the Antarctic desert. *Applied and Environmental Microbiology* **90**(3), e02264–23.
- Wordsworth RD (2016) The climate of early Mars. *Annual Review of Earth and Planetary Sciences* **44**(1), 381–408.
- Wordsworth R, Knoll AH, Hurowitz J, Baum M, Ehlmann BL, Head JW, Steakley K (2021) A coupled model of episodic warming, oxidation and geochemical transitions on early Mars. *Nat. Geosci.* **14**, 127–132. <https://doi.org/10.1038/s41561-021-00701-8>
- Wray RAL (1997) A global review of solutional weathering forms on quartz sandstone. *Earth-Science Reviews* **42**, 137–160.
- Wynne JJ, Titus TN, Agha-Mohammadi AA, Azua-Bustos A, Boston PJ, de León P, ... and Williams KE (2022) Fundamental science and engineering questions in planetary cave exploration. *Journal of Geophysical Research: Planets* **127**(11), e2022JE007194.
- Young RW, Wray RAL and Young ARM (2009) *Sandstone Landforms*. Cambridge: Cambridge University Press.
- Zerboni A (2008) Holocene rock varnish on the Messak plateau (Libyan Sahara): Chronology of weathering processes. *Geomorphology* **102**, 640–651. <https://doi.org/10.1016/j.geomorph.2008.06.010>
- Zerboni A, Perego A and Cremaschi M (2015) Geomorphological map of the Tadrart Acacus massif and the Erg Uan Kasa (Libyan Central Sahara). *Journal of Maps* **11**, 772–787. <https://doi.org/10.1080/17445647.2014.955891>
- Zerboni A, Villa F, Wu Y-L, Solomon T, Trentini A, Rizzi A, Cappitelli F and Gallinaro M (2022) The sustainability of rock art: Preservation and research. *Sustainability* **14**, 6305. <https://doi.org/10.3390/su14106305>
- Zhao J, Xiong K and Yan N (2024) Research progress on algal biokarst and its implications for carbonate rock dissolution and weathering. *Environment, Resources and Ecology Journal* **8**, 128–136. <https://doi.org/10.23977/erej.2024.080116>
- Zhu H-Z, Jiang C-Y and Liu S-J (2024) Microbial roles in cave biogeochemical cycling. *Frontiers in Microbiology* **13**, 950005. <https://doi.org/10.3389/fmicb.2022.950005>